



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>6</sup> :</b> <b>A61K 47/48, 39/00</b>	<b>A2</b>	<b>(11) International Publication Number:</b> <b>WO 98/36779</b> <b>(43) International Publication Date:</b> 27 August 1998 (27.08.98)
<b>(21) International Application Number:</b> PCT/US98/02945 <b>(22) International Filing Date:</b> 18 February 1998 (18.02.98) <b>(30) Priority Data:</b> 08/801,263 19 February 1997 (19.02.97) US <b>(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application</b> US 08/801,263 (CON) Filed on 19 February 1997 (19.02.97) <b>(71) Applicant (for all designated States except US):</b> UNIVERSITY OF NORTH CAROLINA AT CHAPEL HILL [US/US]; 308 Bynum Hall, Campus Box 4105, Chapel Hill, NC 27599-4105 (US). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> JOHNSTON, Robert, E. [US/US]; 101 Marin Place, Chapel Hill, NC 27516 (US). DAVIS, Nancy, L. [US/US]; 132 New Castle Drive, Chapel Hill, NC 27514 (US). SIMPSON, Dennis, A. [US/US]; 19A Deer Mountain Road, Pittsboro, NC 27312 (US).		<b>(74) Agents:</b> MAGRI, Karen, A. et al.; Myers, Bigel, Sibley & Sajovec, L.L.P., P.O. Box 37428, Raleigh, NC 27627 (US). <b>(81) Designated States:</b> AL, AM, AT, AT (Utility model), AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, CZ (Utility model), DE, DE (Utility model), DK, DK (Utility model), EE, EE (Utility model), ES, FI, FI (Utility model), GB, GE, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SK (Utility model), TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). <b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i>
<b>(54) Title:</b> SYSTEM FOR THE <i>IN VIVO</i> DELIVERY AND EXPRESSION OF HETEROLOGOUS GENES IN THE BONE MARROW <b>(57) Abstract</b> <p>The present invention provides a method of delivering immunogenic or therapeutic proteins to bone marrow cells using alphavirus vectors. The alphavirus vectors disclosed herein target specifically to bone marrow tissue, and viral genomes persist in bone marrow for at least three months post-infection. No or very low levels of virus were detected in quadriceps, brain, and sera of treated animals. The sequence of a consensus Sindbis cDNA clone, pTR339, and infectious RNA transcripts, infectious virus particles, and pharmaceutical formulations derived therefrom are also disclosed. The sequence of the genomic RNA of the Girdwood S.A. virus, and cDNA clones, infectious RNA transcripts, infectious virus particles, and pharmaceutical formulations derived therefrom are also disclosed.</p>		

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# SYSTEM FOR THE *IN VIVO* DELIVERY AND EXPRESSION OF HETEROLOGOUS GENES IN THE BONE MARROW

## 5                    FEDERALLY SPONSORED RESEARCH

This invention was made with Government support under Grant Number 5 RO1 AI22186 from the National Institutes of Health. The Government has certain rights to this invention.

## FIELD OF THE INVENTION

10            The present invention relates to recombinant DNA technology, and in particular to introducing and expressing foreign DNA in a eukaryotic cell.

## BACKGROUND OF THE INVENTION

15            The Alphavirus genus includes a variety of viruses all of which are members of the Togaviridae family. The alphaviruses include Eastern Equine Encephalitis virus (EEE), Venezuelan Equine Encephalitis virus (VEE), Everglades virus, Mucambo virus, Pixuna virus, Western Equine Encephalitis virus (WEE), Sindbis virus, South African Arbovirus No. 86 (S.A.AR 86), Girdwood S.A. virus, Ockelbo virus, Semliki Forest virus, Middelburg virus, Chikungunya virus, O'Nyong-Nyong virus, Ross River virus, Barmah Forest virus, Getah virus,  
20            Sagiyama virus, Bebaru virus, Mayaro virus, Una virus, Aura virus, Whataroa virus, Babanki virus, Kyzylagach virus, Highlands J virus, Fort Morgan virus, Ndumu virus, and Buggy Creek virus.

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The alphavirus genome is a single-stranded, messenger-sense RNA, modified at the 5'-end with a methylated cap, and at the 3'-end with a variable-length poly (A) tract. The viral genome is divided into two regions: the first encodes the nonstructural or replicase proteins (nsP1-nsP4) and the second encodes the viral structural proteins. Strauss and Strauss, *Microbiological Rev.* 58, 491-562, 494 (1994). Structural subunits consisting of a single viral protein, C, associate with themselves and with the RNA genome in an icosahedral nucleocapsid. In the virion, the capsid is surrounded by a lipid envelope covered with a regular array of transmembranal protein spikes, each of which consists of a heterodimeric complex of two glycoproteins, E1 and E2. See Paredes et al., *Proc. Natl. Acad. Sci. USA* 90, 9095-99 (1993); Paredes et al., *Virology* 187, 324-32 (1993); Pedersen et al., *J. Virol.* 14:40 (1974).

Sindbis virus, the prototype member of the alphavirus genus of the family *Togaviridae*, and viruses related to Sindbis are broadly distributed throughout Africa, Europe, Asia, the Indian subcontinent, and Australia, based on serological surveys of humans, domestic animals and wild birds. Kokernot et al., *Trans. R. Soc. Trop. Med. Hyg.* 59, 553-62 (1965); Redaksie, *S. Afr. Med. J.* 42, 197 (1968); Adekolu-John and Fagbami, *Trans. R. Soc. Trop. Med. Hyg.* 77, 149-51 (1983); Darwish et al., *Trans. R. Soc. Trop. Med. Hyg.* 77, 442-45 (1983); Lundström et al., *Epidemiol. Infect.* 106, 567-74 (1991); Morrill et al., *J. Trop. Med. Hyg.* 94, 166-68 (1991). The first isolate of Sindbis virus (strain AR339) was recovered from a pool of *Culex* sp. mosquitoes collected in Sindbis, Egypt in 1953 (Taylor et al., *Am. J. Trop. Med. Hyg.* 4, 844-62 (1955)), and is the most extensively studied representative of this group. Other members of the Sindbis group of alphaviruses include South African Arbovirus No. 86, Ockelbo82, and Girdwood S.A. These viruses are not strains of the Sindbis virus; they are related to Sindbis AR339, but they are more closely related to each other based on nucleotide sequence and serological comparisons. Lundström et al., *J. Wildl. Dis.* 29, 189-95 (1993); Simpson et al., *Virology* 222, 464-69 (1996). Ockelbo82, S.A.AR86 and Girdwood S.A. are all associated with human disease, whereas Sindbis is not. The clinical symptoms of human infection with Ockelbo82,

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S.A.AR86, or Girdwood S.A. are a febrile illness, general malaise, macropapular rash, and joint pain that occasionally progresses to a polyarthralgia sometimes lasting from a few months to a few years.

5 The study of these viruses has led to the development of beneficial techniques for vaccinating against the alphavirus diseases, and other diseases through the use of alphavirus vectors for the introduction of foreign DNA. See United States Patent No. 5,185,440 to Davis et al., and PCT Publication WO 92/10578. It is intended that all United States patent references be incorporated in their entirety by reference.

10 It is well known that live, attenuated viral vaccines are among the most successful means of controlling viral disease. However, for some virus pathogens, immunization with a live virus strain may be either impractical or unsafe. One alternative strategy is the insertion of sequences encoding immunizing antigens of such agents into a vaccine strain of another virus. One such system  
15 utilizing a live VEE vector is described in United States Patent No. 5,505,947 to Johnston et al.

Sindbis virus vaccines have been employed as viral carriers in virus constructs which express genes encoding immunizing antigens for other viruses. See United States Patent No. 5,217,879 to Huang et al. Huang et al. describes  
20 Sindbis infectious viral vectors. However, the reference does not describe the cDNA sequence of Girdwood S.A. and TR339, nor clones or viral vectors produced therefrom.

Another such system is described by Hahn et al., *Proc. Natl. Acad. Sci. USA* 89:2679 (1992), wherein Sindbis virus constructs which express a  
25 truncated form of the influenza hemagglutinin protein are described. The constructs are used to study antigen processing and presentation *in vitro* and in mice. Although no infectious challenge dose is tested, it is also suggested that

such constructs might be used to produce protective B- and T-cell mediated immunity.

London et al., *Proc. Natl. Acad. Sci. USA* 89, 207-11 (1992), disclose a method of producing an immune response in mice against a lethal Rift Valley Fever (RVF) virus by infecting the mice with an infectious Sindbis virus containing an RVF epitope. London does not disclose using Girdwood S.A. or TR339 to induce an immune response in animals.

Viral carriers can also be used to introduce and express foreign DNA in eukaryotic cells. One goal of such techniques is to employ vectors that target expression to particular cells and/or tissues. A current approach has been to remove target cells from the body, culture them *ex vivo*, infect them with an expression vector, and then reintroduce them into the patient.

PCT Publication No. WO 92/10578 to Garoff and Liljeström provide a system for introducing and expressing foreign proteins in animal cells using alphaviruses. This reference discloses the use of Semliki Forest virus to introduce and express foreign proteins in animal cells. The use of Girdwood S.A. or TR339 is not discussed. Furthermore, this reference does not provide a method of targeting and introducing foreign DNA into specific cell or tissue types.

Accordingly, there remains a need in the art for full-length cDNA clones of positive-strand RNA viruses, such as Girdwood S.A and TR339. In addition, there is an ongoing need in the art for improved vaccination strategies. Finally, there remains a need in the art for improved methods and nucleic acid sequences for delivering foreign DNA to target cells.

#### SUMMARY OF THE INVENTION

A first aspect of the present invention is a method of introducing and expressing heterologous RNA in bone marrow cells, comprising: (a) providing

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a recombinant alphavirus, the alphavirus containing a heterologous RNA segment, the heterologous RNA segment comprising a promoter operable in bone marrow cells operatively associated with a heterologous RNA to be expressed in bone marrow cells; and then (b) contacting the recombinant alphavirus to the bone marrow cells so that the heterologous RNA segment is introduced and expressed therein.

As a second aspect, the present invention provides a helper cell for expressing an infectious, propagation defective, Girdwood S.A. virus particle, comprising, in a Girdwood S.A.-permissive cell: (a) a first helper RNA encoding (i) at least one Girdwood S.A. structural protein, and (ii) not encoding at least one other Girdwood S.A. structural protein; and (b) a second helper RNA separate from the first helper RNA, the second helper RNA (i) not encoding the at least one Girdwood S.A. structural protein encoded by the first helper RNA, and (ii) encoding the at least one other Girdwood S.A. structural protein not encoded by the first helper RNA, and with all of the Girdwood S.A. structural proteins encoded by the first and second helper RNAs assembling together into Girdwood S.A. particles in the cell containing the replicon RNA; and wherein the Girdwood S.A. packaging segment is deleted from at least the first helper RNA.

A third aspect of the present invention is a method of making infectious, propagation defective, Girdwood S.A. virus particles, comprising: transfecting a Girdwood S.A.-permissive cell with a propagation defective replicon RNA, the replicon RNA including the Girdwood S.A. packaging segment and an inserted heterologous RNA; producing the Girdwood S.A. virus particles in the transfected cell; and then collecting the Girdwood S.A. virus particles from the cell. Also disclosed are infectious Girdwood S.A. RNAs, cDNAs encoding the same, infectious Girdwood S.A. virus particles, and pharmaceutical formulations thereof.

As a fourth aspect, the present invention provides a helper cell for expressing an infectious, propagation defective, TR339 virus particle, comprising,

in a TR339-permissive cell: (a) a first helper RNA encoding (i) at least one TR339 structural protein, and (ii) not encoding at least one other TR339 structural protein; and (b) a second helper RNA separate from the first helper RNA, the second helper RNA (i) not encoding the at least one TR339 structural protein encoded by the first helper RNA, and (ii) encoding the at least one other TR339 structural protein not encoded by the first helper RNA, and with all of the TR339 structural proteins encoded by the first and second helper RNAs assembling together into TR339 particles in the cell containing the replicon RNA; and wherein the TR339 packaging segment is deleted from at least the first helper RNA.

A fifth aspect of the present invention is a method of making infectious, propagation defective, TR339 virus particles, comprising: transfecting a TR339-permissive cell with a propagation defective replicon RNA, the replicon RNA including the TR339 packaging segment and an inserted heterologous RNA; producing the TR339 virus particles in the transfected cell; and then collecting the TR339 virus particles from the cell. Also disclosed are infectious TR339 RNAs, cDNAs encoding the same, infectious TR339 virus particles, and pharmaceutical formulations thereof.

As a sixth aspect, the present invention provides a recombinant DNA comprising a cDNA coding for an infectious Girdwood S.A. virus RNA transcript, and a heterologous promoter positioned upstream from the cDNA and operatively associated therewith. The present invention also provides infectious RNA transcripts encoded by the above-mentioned cDNA and infectious viral particles containing the infectious RNA transcripts.

As a seventh aspect, the present invention provides a recombinant DNA comprising a cDNA coding for a Sindbis strain TR339 RNA transcript, and a heterologous promoter positioned upstream from the cDNA and operatively associated therewith. The present invention also provides infectious RNA transcripts encoded by the above-mentioned cDNA and infectious viral particles containing the infectious RNA transcripts.

The foregoing and other aspects of the present invention are described in the detailed description set forth below.

### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 presents the cDNA sequence (SEQ ID NO:1) of S.A.AR86. The RNA sequence of the 5' 40 nucleotides was obtained by direct sequencing of the genomic RNA. The rest of the genome was sequenced by RT-PCR of fragments amplified from virion RNA. Nucleotides 1 through 59 represent the 5' UTR, the non-structural polyprotein is encoded by nucleotides 60 through 7559 (nsP1--nt60 through nt1679; nsP2--nt1680 through nt4099; nsP3--nt4100 through nt5729; nsP4--nt5730 through nt7559), the structural polyprotein is encoded by nucleotides 7608 through 11342 (capsid--nt7608 through nt8399; E3--nt8400 through nt8591; E2--nt8592 through nt9860; 6K--nt9861 through nt10025; E1--nt10026 through nt11342), and the 3' UTR is represented by nucleotides 11346 through 11663.

Figure 1A shows nucleotides 1 through 3800 of the cDNA sequence of S.A.AR86.

Figure 1B shows nucleotides 3801 through 7900 of the cDNA sequence of S.A.AR86.

Figure 1C shows nucleotides 7901 through 11663 of the cDNA sequence of S.A.AR86.

Figure 2 presents the putative amino acid sequences of the S.A.AR86 polyproteins (SEQ ID NO:2 and SEQ ID NO:3). The amino acids were derived from the S.A.AR86 cDNA sequence given in Figure 1 (SEQ ID NO:1).

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Figure 2A shows the amino acid sequence of the non-structural polyprotein of S.A.AR86 (SEQ ID NO:2).

Figure 2B shows the amino acid sequence of the structural polyprotein of S.A.AR86 (SEQ ID NO:3).

5                   Figure 3 presents the cDNA sequence (SEQ ID NO:4) of Girdwood S.A. The RNA sequence of the 5' 40 nucleotides was obtained by direct sequencing of the genomic RNA. The rest of the genome sequence was obtained by sequencing of fragments amplified by RT-PCR from virion RNA. An "N" in the sequence indicates that the identity of the nucleotide at that position is  
10                   unknown. Nucleotides 1 through 59 represent the 5' UTR, the non-structural polyprotein is encoded by nucleotides 60 through 7613 (nsP1--nt60 through nt1679; nsP2--nt1680 through nt4099; nsP3--nt4100 through nt5762 or nt5783; nsP4--nt5784 through nt7613), the structural polyprotein is encoded by nucleotides  
15                   7662 through 11396 (capsid--nt7662 through nt8453; E3--nt8454 through nt8645; E2--nt8646 through nt9914, 6K--9915 through nt10079; E1--nt10080 through nt11396), and the 3' UTR is represented by nucleotides 11400 through 11717. There is an opal termination codon at nucleotides 5763 through 5765.

Figure 3A shows nucleotides 1 through 3800 of the cDNA sequence of Girdwood S.A.

20                   Figure 3B shows nucleotides 3801 through 7900 of the cDNA sequence of Girdwood S.A.

Figure 3C shows nucleotides 7901 through 11717 of the cDNA sequence of Girdwood S.A.

25                   Figure 4 illustrates the putative amino acid sequences of the Girdwood S.A. polyproteins (SEQ ID NO:5 and SEQ ID NO:6). The amino

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acids were derived from the Girdwood S.A. cDNA sequence given in Figure 3 (SEQ ID NO:4).

Figure 4A shows the amino acid sequence of the non-structural polyprotein of Girdwood S.A. The sequence terminates at the opal termination codon. The complete amino acid sequence is presented in SEQ ID NO:5.

Figure 4B shows the amino acid sequence of the structural polyprotein of Girdwood S.A. (SEQ ID NO:6).

Figure 5 illustrates the nucleotide sequence (SEQ ID NO:7) of clone pS55, a cDNA clone of the S.A.AR86 genomic RNA.

Figure 5A shows nucleotides 1 through 6720 of the cDNA sequence of pS55.

Figure 5B shows nucleotides 6721 through 11663 of the cDNA sequence of pS55.

Figure 6 presents the cDNA sequence (SEQ ID NO:8) of clone pTR339. The TR339 virus is derived from this clone. Nucleotides 1 through 59 represent the 5' UTR, the non-structural polyprotein is encoded by nucleotides 60 through 7598 (nsP1--nt60 through nt1679; nsP2--nt1680 through nt4099; nsP3--nt4100 through nt5747 or 5768; nsP4--nt5769 through nt7598), the structural polyprotein is encoded by nucleotides 7647 through 11381 (capsid--nt7647 through nt8438; E3--nt8439 through nt8630; E2--nt8631 through nt9899; 6K--nt9900 through nt10064; E1--nt10065 through nt11381), and the 3' UTR is represented by nucleotides 11382 through 11703. There is an opal termination codon at nucleotides 5748 through 5750.

Figure 6A shows nucleotides 1 through 6720 of the cDNA sequence of pTR339.

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Figure 6B shows nucleotides 6721 through 11703 of the cDNA sequence of pTR339.

### DETAILED DESCRIPTION OF THE INVENTION

5 The production and use of recombinant DNA, vectors, transformed host cells, selectable markers, proteins, and protein fragments by genetic engineering are well-known to those skilled in the art. *See, e.g.*, United States Patent No. 4,761,371 to Bell et al. at Col. 6 line 3 to Col. 9 line 65; United States Patent No. 4,877, 729 to Clark et al. at Col. 4 line 38 to Col. 7 line 6; United States Patent No. 4,912,038 to Schilling at Col 3 line 26 to Col 14 line 12; and  
10 United States Patent No. 4,879,224 to Wallner at Col. 6 line 8 to Col. 8 line 59.

The term "alphavirus" has its conventional meaning in the art, and includes the various species of alphaviruses such as Eastern Equine Encephalitis virus (EEE), Venezuelan Equine Encephalitis virus (VEE), Everglades virus, Mucambo virus, Pixuna virus, Western Encephalitis virus (WEE), Sindbis virus,  
15 South African Arbovirus No. 86, Girdwood S.A. virus, Ockelbo virus, Semliki Forest virus, Middelburg virus, Chikungunya virus, O'Nyong-Nyong virus, Ross River virus, Barmah Forest virus, Getah virus, Sagiya virus, Bebaru virus, Mayaro virus, Una virus, Aura virus, Whataroa virus, Babanki virus, Kyzlagach virus, Highlands J virus, Fort Morgan virus, Ndumu virus, Buggy Creek virus,  
20 and any other virus classified by the International Committee on Taxonomy of Viruses (ICTV) as an alphavirus. The preferred alphaviruses for use in the present invention include Sindbis virus strains (*e.g.*, TR339), Girdwood S.A., S.A.AR86, and Ockelbo82.

An "Old World alphavirus" is a virus that is primarily distributed throughout the Old World. Alternately stated, an Old World alphavirus is a virus that is primarily distributed throughout Africa, Asia, Australia and New Zealand, or Europe. Exemplary Old World viruses include SF group alphaviruses and SIN group alphaviruses. SF group alphaviruses include Semliki Forest virus, Middelburg virus, Chikungunya virus, O'Nyong-Nyong virus, Ross River virus,  
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Barmah Forest virus, Getah virus, Sagiyama virus, Bebaru virus, Mayaro virus, and Una virus. SIN group alphaviruses include Sindbis virus, South African Arbovirus No. 86, Ockelbo virus, Girdwood S.A. virus, Aura virus, Whataroa virus, Babanki virus, and Kyzylagach virus.

5           Acceptable alphaviruses include those containing attenuating mutations. The phrases "attenuating mutation" and "attenuating amino acid," as used herein, mean a nucleotide sequence containing a mutation, or an amino acid encoded by a nucleotide sequence containing a mutation, which mutation results in a decreased probability of causing disease in its host (*i.e.*, a loss of virulence),  
10           in accordance with standard terminology in the art, whether the mutation be a substitution mutation or an in-frame deletion mutation. *See, e.g.*, B. DAVIS ET AL., MICROBIOLOGY 132 (3d ed. 1980). The phrase "attenuating mutation" excludes mutations or combinations of mutations which would be lethal to the virus.

15           Appropriate attenuating mutations will be dependent upon the alphavirus used. Suitable attenuating mutations within the alphavirus genome will be known to those skilled in the art. Exemplary attenuating mutations include, but are not limited to, those described in United States Patent No. 5,505,947 to Johnston et al., copending United States application 08/448,630 to Johnston et al.,  
20           and copending United States application 08/446,932 to Johnston et al. It is intended that all United States patent references be incorporated in their entirety by reference.

25           Attenuating mutations may be introduced into the RNA by performing site-directed mutagenesis on the cDNA which encodes the RNA, in accordance with known procedures. *See*, Kunkel, *Proc. Natl. Acad. Sci. USA* 82, 488 (1985), the disclosure of which is incorporated herein by reference in its entirety. Alternatively, mutations may be introduced into the RNA by replacement of homologous restriction fragments in the cDNA which encodes for the RNA, in accordance with known procedures.

I. Methods for Introducing and Expressing Heterologous RNA in Bone Marrow Cells.

5 The present invention provides methods of using a recombinant alphavirus to introduce and express a heterologous RNA in bone marrow cells. Such methods are useful as vaccination strategies when the heterologous RNA encodes an immunogenic protein or peptide. Alternatively, such methods are useful in introducing and expressing in bone marrow cells an RNA which encodes a desirable protein or peptide, for example, a therapeutic protein or peptide.

10 The present invention is carried out using a recombinant alphavirus to introduce a heterologous RNA into bone marrow cells. Any alphavirus that targets and infects bone marrow cells is suitable. Preferred alphaviruses include Old World alphaviruses, more preferably SF group alphaviruses and SIN group alphaviruses, more preferably Sindbis virus strains (*e.g.*, TR339), S.A.AR86 virus, Girdwood S.A. virus, and Ockelbo virus. In a more preferred embodiment,  
15 the alphavirus contains one or more attenuating mutations, as described hereinabove.

20 Two types of recombinant virus vector are contemplated in carrying out the present invention. In one embodiment employing "double promoter vectors," the heterologous RNA is inserted into a replication and propagation competent virus. Double promoter vectors are described in United States Patent No. 5,505,947 to Johnston et al. With this type of viral vector, it is preferable that heterologous RNA sequences of less than 3 kilobases are inserted into the viral vector, more preferably those less than 2 kilobases, and more preferably still those less than 1 kilobase. In an alternate embodiment, propagation-defective "replicon  
25 vectors," as described in copending United States application 08/448,630 to Johnston et al., will be used. One advantage of replicon viral vectors is that larger RNA inserts, up to approximately 4-5 kilobases in length can be utilized. Double promoter vectors and replicon vectors are described in more detail hereinbelow.

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The recombinant alphaviruses of the claimed method target the heterologous RNA to bone marrow cells, where it expresses the encoded protein or peptide. Heterologous RNA can be introduced and expressed in any cell type found in the bone marrow. Bone marrow cells that may be targeted by the recombinant alphaviruses of the present invention include, but are not limited to, polymorphonuclear cells, hemopoietic stem cells (including megakaryocyte colony forming units (CFU-M), spleen colony forming units (CFU-S), erythroid colony forming units (CFU-E), erythroid burst forming units (BFU-E), and colony forming units in culture (CFU-C), erythrocytes, macrophages (including reticular cells), monocytes, granulocytes, megakaryocytes, lymphocytes, fibroblasts, osteoprogenitor cells, osteoblasts, osteoclasts, marrow stromal cells, chondrocytes and other cells of synovial joints. Preferably, marrow cells within the endosteum are targeted, more preferably osteoblasts. Also preferred are methods in which cells in the endosteum of synovial joints (*e.g.*, hip and knee joints) are targeted.

By targeting to the cells of the bone marrow, it is meant that the primary site in which the virus will be localized *in vivo* is the cells of the bone marrow. Alternately stated, the alphaviruses of the present invention target bone marrow cells, such that titers in bone marrow two days after infection are greater than 100 PFU/g crushed bone, preferably greater than 200 PFU/g crushed bone, more preferably greater than 300 PFU/g crushed bone, and more preferably still greater than 500 PFU/g crushed bone. Virus may be detected occasionally in other cell or tissue types, but only sporadically and usually at low levels. Virus localization in the bone marrow can be demonstrated by any suitable technique known in the art, such as *in situ* hybridization.

Bone marrow cells are long-lived and harbor infectious alphaviruses for a prolonged period of time, as demonstrated in the Examples below. These characteristics of bone marrow cells render the present invention useful not only for the purpose of supplying a desired protein or peptide to skeletal tissue, but also for expressing proteins or peptides *in vivo* that are needed by other cell or tissue types.

The present invention can be carried out *in vivo* or with cultured bone marrow cells *in vitro*. Bone marrow cell cultures include primary cultures

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of bone marrow cells, serially-passaged cultures of bone marrow cells, and cultures of immortalized bone marrow cell lines. Bone marrow cells may be cultured by any suitable means known in the art.

The recombinant alphaviruses of the present invention carry a heterologous RNA segment. The heterologous RNA segment encodes a promoter and an inserted heterologous RNA. The inserted heterologous RNA may encode any protein or a peptide which is desirably expressed by the host bone marrow cells. Suitable heterologous RNA may be of prokaryotic (*e.g.*, RNA encoding the *Botulinus* toxin C), or eukaryotic (*e.g.*, RNA encoding malaria *Plasmodium* protein cs1) origin. Illustrative proteins and peptides encoded by the heterologous RNAs of the present invention include hormones, growth factors, interleukins, cytokines, chemokines, enzymes, and ribozymes. Alternately, the heterologous RNAs encode any therapeutic protein or peptide. As a further alternative, the heterologous RNAs of the present invention encode any immunogenic protein or peptide.

An immunogenic protein or peptide, or "immunogen," may be any protein or peptide suitable for protecting the subject against a disease, including but not limited to microbial, bacterial, protozoal, parasitic, and viral diseases. For example, the immunogen may be an orthomyxovirus immunogen (*e.g.*, an influenza virus immunogen, such as the influenza virus hemagglutinin (HA) surface protein or the influenza virus nucleoprotein gene, or an equine influenza virus immunogen), or a lentivirus immunogen (*e.g.*, an equine infectious anemia virus immunogen, a Simian Immunodeficiency Virus (SIV) immunogen, or a Human Immunodeficiency Virus (HIV) immunogen, such as the HIV envelope GP160 protein and the HIV matrix/capsid proteins). The immunogen may also be an arenavirus immunogen (*e.g.*, Lassa fever virus immunogen, such as the Lassa fever virus nucleocapsid protein gene and the Lassa fever envelope glycoprotein gene), a poxvirus immunogen (*e.g.*, vaccinia), a flavivirus immunogen (*e.g.*, a yellow fever virus immunogen or a Japanese encephalitis virus immunogen), a filovirus immunogen (*e.g.*, an Ebola virus immunogen, or a Marburg virus

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immunogen), a bunyavirus immunogen (*e.g.*, RVFV, CCHF, and SFS viruses),  
or a coronavirus immunogen (*e.g.*, an infectious human coronavirus immunogen,  
such as the human coronavirus envelope glycoprotein gene, or a transmissible  
gastroenteritis virus immunogen for pigs, or an infectious bronchitis virus  
immunogen for chickens).

Alternatively, the present invention can be used to express  
heterologous RNAs encoding antisense oligonucleotides. In general, "antisense"  
refers to the use of small, synthetic oligonucleotides to inhibit gene expression by  
inhibiting the function of the target mRNA containing the complementary  
sequence. Milligan, J.F. et al., *J. Med. Chem.* 36(14), 1923-1937 (1993). Gene  
expression is inhibited through hybridization to coding (sense) sequences in a  
specific mRNA target by hydrogen bonding according to Watson-Crick base  
pairing rules. The mechanism of antisense inhibition is that the exogenously  
applied oligonucleotides decrease the mRNA and protein levels of the target gene.  
Milligan, J.F. et al., *J. Med. Chem.* 36(14), 1923-1937 (1993). *See also* Helene,  
C. and Toulme, J., *Biochim. Biophys. Acta* 1049, 99-125 (1990); Cohen, J.S.,  
Ed., OLIGODEOXYNUCLEOTIDES AS ANTISENSE INHIBITORS OF GENE  
EXPRESSION, CRC Press:Boca Raton, FL (1987).

Antisense oligonucleotides may be of any suitable length, depending  
on the particular target being bound. The only limits on the length of the antisense  
oligonucleotide is the capacity of the virus for inserted heterologous RNA.  
Antisense oligonucleotides may be complementary to the entire mRNA transcript  
of the target gene or only a portion thereof. Preferably the antisense  
oligonucleotide is directed to an mRNA region containing a junction between  
intron and exon. Where the antisense oligonucleotide is directed to an intron/exon  
junction, it may either entirely overlie the junction or may be sufficiently close to  
the junction to inhibit splicing out of the intervening exon during processing of  
precursor mRNA to mature mRNA (*e.g.*, with the 3' or 5' terminus of the  
antisense oligonucleotide being positioned within about, for example, 10, 5, 3 or

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2 nucleotides of the intron/exon junction). Also preferred are antisense oligonucleotides which overlap the initiation codon.

When practicing the present invention, the antisense oligonucleotides administered may be related in origin to the species to which it is administered.

5 When treating humans, human antisense may be used if desired.

Promoters for use in carrying out the present invention are operable in bone marrow cells. An operable promoter in bone marrow cells is a promoter that is recognized by and functions in bone marrow cells. Promoters for use with the present invention must also be operatively associated with the heterologous RNA to be expressed in the bone marrow. A promoter is operably linked to a heterologous RNA if it controls the transcription of the heterologous RNA, where the heterologous RNA comprises a coding sequence. Suitable promoters are well known in the art. The Sindbis 26S promoter is preferred when the alphavirus is a strain of Sindbis virus. Additional preferred promoters beyond the Sindbis 26S promoter include the Girdwood S.A. 26S promoter when the alphavirus is Girdwood S.A., the S.A.AR86 26S promoter when the alphavirus is S.A.AR86, and any other promoter sequence recognized by alphavirus polymerases. Alphavirus promoter sequences containing mutations which alter the activity level of the promoter (in relation to the activity level of the wild-type) are also suitable in the practice of the present invention. Such mutant promoter sequences are described in Raju and Huang, *J. Virol.* 65, 2501-2510 (1991), the disclosure of which is incorporated in its entirety by reference.

The heterologous RNA is introduced into the bone marrow cells by contacting the recombinant alphavirus carrying the heterologous RNA segment to the bone marrow cells. By contacting, it is meant bringing the recombinant alphavirus and the bone marrow cells in physical proximity. The contacting step can be performed *in vitro* or *in vivo*. *In vitro* contacting can be carried out with cultures of immortalized or non-immortalized bone marrow cells. In one particular embodiment, bone marrow cells can be removed from a subject, cultured *in vitro*,

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infected with the vector, and then introduced back into the subject. Contacting is performed *in vivo* when the recombinant alphavirus is administered to a subject. Pharmaceutical formulations of recombinant alphavirus can be administered to a subject parenterally (*e.g.*, subcutaneous, intracerebral, intradermal, intramuscular, intravenous and intraarticular) administration. Alternatively, pharmaceutical formulations of the present invention may be suitable for administration to the mucus membranes of a subject (*e.g.*, intranasal administration, by use of a dropper, swab, or inhaler). Methods of preparing infectious virus particles and pharmaceutical formulations thereof are discussed in more detail hereinbelow.

By "introducing" the heterologous RNA segment into the bone marrow cells it is meant infecting the bone marrow cells with recombinant alphavirus containing the heterologous RNA, such that the viral vector carrying the heterologous RNA enters the bone marrow cells and can be expressed therein. As used with respect to the present invention, when the heterologous RNA is "expressed," it is meant that the heterologous RNA is transcribed. In particular embodiments of the invention in which it is desired to produce a protein or peptide, expression further includes the steps of post-transcriptional processing and translation of the mRNA transcribed from the heterologous RNA. In contrast, where the heterologous RNA encodes an antisense oligonucleotide, expression need not include post-transcriptional processing and translation. With respect to embodiments in which the heterologous RNA encodes an immunogenic protein or a protein being administered for therapeutic purposes, expression may also include the further step of post-translational processing to produce an immunogenic or therapeutically-active protein.

The present invention also provides infectious RNAs, as described hereinabove, and cDNAs encoding the same. Preferably the infectious RNAs and cDNAs are derived from the S.A.AR86, Girdwood S.A., TR339, or Ockelbo viruses. The cDNA clones can be generated by any of a variety of suitable methods known to those skilled in the art. A preferred method is the method set

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forth in United States Patent No. 5,185,440 to Davis et al., the disclosure of which is incorporated in its entirety by reference, and Gubler et al., *Gene* 25:263 (1983).

RNA is preferably synthesized from the DNA sequence *in vitro* using purified RNA polymerase in the presence of ribonucleotide triphosphates and cap analogs in accordance with conventional techniques. However, the RNA may also be synthesized intracellularly after introduction of the cDNA.

#### A. Double Promoter Vectors.

In one embodiment of the invention, double promoter vectors are used to introduce the heterologous RNA into the target bone marrow cells. A double promoter virus vector is a replication and propagation competent virus. Double promoter vectors are described in United States Patent No. 5,505,947 to Johnston et al., the disclosure of which is incorporated in its entirety by reference. Preferred alphaviruses for constructing the double promoter vectors are S.A. AR86, Girdwood S.A., TR339 and Ockelbo viruses. More preferably, the double promoter vector contains one or more attenuating mutations. Attenuating mutations are described in more detail hereinabove.

The double promoter vector is constructed so as to contain a second subgenomic promoter (*i.e.*, 26S promoter) inserted 3' to the virus RNA encoding the structural proteins. The heterologous RNA is inserted between the second subgenomic promoter, so as to be operatively associated therewith, and the 3' UTR of the virus genome. Heterologous RNA sequences of less than 3 kilobases, more preferably those less than 2 kilobases, and more preferably still those less than 1 kilobase, can be inserted into the double promoter vector. In a preferred embodiment of the invention, the double promoter vector is derived from Girdwood S.A., and the second subgenomic promoter is a duplicate of the Girdwood S.A. subgenomic promoter. In an alternate preferred embodiment, the double promoter vector is derived from TR339, and the second subgenomic promoter is a duplicate of the TR339 subgenomic promoter.

### B. Replicon Vectors.

Replicon vectors, which are propagation-defective virus vectors can also be used to carry out the present invention. Replicon vectors are described in more detail in copending United States Application 08/448,630 to Johnston et al., the disclosure of which is incorporated in its entirety by reference. Preferred alphaviruses for constructing the replicon vectors are S.A.AR86, Girdwood S.A., TR339, and Ockelbo.

In general, in the replicon system, a foreign gene to be expressed is inserted in place of at least one of the viral structural protein genes in a transcription plasmid containing an otherwise full-length cDNA copy of the alphavirus genome RNA. RNA transcribed from this plasmid contains an intact copy of the viral nonstructural genes which are responsible for RNA replication and transcription. Thus, if the transcribed RNA is transfected into susceptible cells, it will be replicated and translated to give the nonstructural proteins. These proteins will transcribe the transfected RNA to give high levels of subgenomic mRNA, which will then be translated to produce high levels of the foreign protein. The autonomously replicating RNA (*i.e.*, replicon) can only be packaged into virus particles if the alphavirus structural protein genes are provided on one or more "helper" RNAs, which are cotransfected into cells along with the replicon RNA. The helper RNAs do not contain the viral nonstructural genes for replication, but these functions are provided *in trans* by the replicon RNA. Similarly, the transcriptase functions translated from the replicon RNA transcribe the structural protein genes on the helper RNA, resulting in the synthesis of viral structural proteins and packaging of the replicon into virus-like particles. As the packaging or encapsidation signal for alphavirus RNAs is located within the nonstructural genes, the absence of these sequences in the helper RNAs precludes their incorporation into virus particles.

Alphavirus-permissive cells employed in the methods of the present invention are cells which, upon transfection with the viral RNA transcript, are capable of producing viral particles. Preferred alphavirus-permissive cells are

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TR339-permissive cells, Girdwood S.A.-permissive cells, S.A.AR86-permissive cells, and Ockelbo-permissive cells. Alphaviruses have a broad host range. Examples of suitable host cells include, but are not limited to Vero cells, baby hamster kidney (BHK) cells, and chicken embryo fibroblast cells.

5           The phrase "structural protein" as used herein refers to the encoded proteins which are required for encapsidation (*e.g.*, packaging) of the RNA replicon, and include the capsid protein, E1 glycoprotein, and E2 glycoprotein. As described hereinabove, the structural proteins of the alphavirus are distributed among one or more helper RNAs (*i.e.*, a first helper RNA and a second helper RNA). In addition, one or  
10 more structural proteins may be located on the same RNA molecule as the replicon RNA, provided that at least one structural protein is deleted from the replicon RNA such that the resulting alphavirus particle is propagation defective. As used herein, the terms "deleted" or "deletion" mean either total deletion of the specified segment or the deletion of a sufficient portion of the specified segment to render the segment inoperative or  
15 nonfunctional, in accordance with standard usage. *See, e.g.*, U.S. Patent No. 4,650,764 to Temin et al. The term "propagation defective" as used herein, means that the replicon RNA cannot be encapsidated in the host cell in the absence of the helper RNA. The resulting alphavirus replicon particles are propagation defective inasmuch as the replicon RNA in these particles does not include all of the alphavirus structural proteins required  
20 for encapsidation, at least one of the required structural proteins being deleted therefrom, such that the replicon RNA initiates only an abortive infection; no new viral particles are produced, and there is no spread of the infection to other cells.

The helper cell for expressing the infectious, propagation defective alphavirus particle comprises a set of RNAs, as described above. The set of RNAs principally  
25 include a first helper RNA and a second helper RNA. The first helper RNA includes RNA encoding at least one alphavirus structural protein but does not encode all alphavirus structural proteins. In other words, the first helper RNA does not encode at least one alphavirus structural protein; the at least one non-coded alphavirus structural protein being deleted from the first helper RNA.

In one embodiment, the first helper RNA includes RNA encoding the alphavirus E1 glycoprotein, with the alphavirus capsid protein and the alphavirus E2 glycoprotein being deleted from the first helper RNA. In another embodiment, the first helper RNA includes RNA encoding the alphavirus E2 glycoprotein, with the alphavirus capsid protein and the alphavirus E1 glycoprotein being deleted from the first helper RNA. In a third, preferred embodiment, the first helper RNA includes RNA encoding the alphavirus E1 glycoprotein and the alphavirus E2 glycoprotein, with the alphavirus capsid protein being deleted from the first helper RNA.

The second helper RNA includes RNA encoding at least one alphavirus structural protein which is different from the at least one structural protein encoded by the first helper RNA. Thus, the second helper RNA encodes at least one alphavirus structural protein which is not encoded by the first helper RNA. The second helper RNA does not encode the at least one alphavirus structural protein which is encoded by the first helper RNA, thus the first and second helper RNAs do not encode duplicate structural proteins. In the embodiment wherein the first helper RNA includes RNA encoding only the alphavirus E1 glycoprotein, the second helper RNA may include RNA encoding one or both of the alphavirus capsid protein and the alphavirus E2 glycoprotein which are deleted from the first helper RNA. In the embodiment wherein, the first helper RNA includes RNA encoding only the alphavirus E2 glycoprotein, the second helper RNA may include RNA encoding one or both of the alphavirus capsid protein and the alphavirus E1 glycoprotein which are deleted from the first helper RNA. In the embodiment wherein the first helper RNA includes RNA encoding both the alphavirus E1 glycoprotein and the alphavirus E2 glycoprotein, the second helper RNA may include RNA encoding the alphavirus capsid protein which is deleted from the first helper RNA.

In one embodiment, the packaging segment (RNA comprising the encapsidation or packaging signal) is deleted from at least the first helper RNA.

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In a preferred embodiment, the packaging segment is deleted from both the first helper RNA and the second helper RNA.

In the preferred embodiment wherein the packaging segment is deleted from both the first helper RNA and the second helper RNA, the helper cell is co-transfected with a replicon RNA in addition to the first helper RNA and the second helper RNA. The replicon RNA encodes the packaging segment and an inserted heterologous RNA. The inserted heterologous RNA may be RNA encoding a protein or a peptide. In a preferred embodiment, the replicon RNA, the first helper RNA and the second helper RNA are provided on separate molecules such that a first molecule, *i.e.*, the replicon RNA, includes RNA encoding the packaging segment and the inserted heterologous RNA, a second molecule, *i.e.*, the first helper RNA, includes RNA encoding at least one but not all of the required alphavirus structural proteins, and a third molecule, *i.e.*, the second helper RNA, includes RNA encoding at least one but not all of the required alphavirus structural proteins. For example, in one preferred embodiment of the present invention, the helper cell includes a set of RNAs which include (a) a replicon RNA including RNA encoding an alphavirus packaging sequence and an inserted heterologous RNA, (b) a first helper RNA including RNA encoding the alphavirus E1 glycoprotein and the alphavirus E2 glycoprotein, and (c) a second helper RNA including RNA encoding the alphavirus capsid protein so that the alphavirus E1 glycoprotein, the alphavirus E2 glycoprotein and the capsid protein assemble together into alphavirus particles in the host cell.

In an alternate embodiment, the replicon RNA and the first helper RNA are on separate molecules, and the replicon RNA and RNA encoding a structural gene not encoded by the first helper RNA are on another single molecule together, such that a first molecule, *i.e.*, the first helper RNA, including RNA encoding at least one but not all of the required alphavirus structural proteins, and a second molecule, *i.e.*, the replicon RNA, including RNA encoding the packaging segment, the inserted heterologous RNA, and the remaining structural proteins not encoded by the first helper RNA. For example, in one preferred embodiment of

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the present invention, the helper cell includes a set of RNAs including (a) a replicon RNA including RNA encoding an alphavirus packaging sequence, an inserted heterologous RNA, and an alphavirus capsid protein, and (b) a first helper RNA including RNA encoding the alphavirus E1 glycoprotein and the alphavirus E2 glycoprotein so that the alphavirus E1 glycoprotein, the alphavirus E2 glycoprotein and the capsid protein assemble together into alphavirus particles in the host cell, with the replicon RNA packaged therein.

In one preferred embodiment of the present invention, the RNA encoding the alphavirus structural proteins, *i.e.*, the capsid, E1 glycoprotein and E2 glycoprotein, contains at least one attenuating mutation, as described hereinabove. Thus, according to this embodiment, at least one of the first helper RNA and the second helper RNA includes at least one attenuating mutation. In a more preferred embodiment, at least one of the first helper RNA and the second helper RNA includes at least two, or multiple, attenuating mutations. The multiple attenuating mutations may be positioned in either the first helper RNA or in the second helper RNA, or they may be distributed randomly with one or more attenuating mutations being positioned in the first helper RNA and one or more attenuating mutations positioned in the second helper RNA. Alternatively, when the replicon RNA and the RNA encoding the structural proteins not encoded by the first helper RNA are located on the same molecule, an attenuating mutation may be positioned in the RNA which codes for the structural protein not encoded by the first helper RNA. The attenuating mutations may also be located within the RNA encoding non-structural proteins (*e.g.*, the replicon RNA).

Preferably, the first helper RNA and the second helper RNA also include a promoter. It is also preferred that the replicon RNA also includes a promoter. Suitable promoters for inclusion in the first helper RNA, second helper RNA and replicon RNA are well known in the art. One preferred promoter is the Girdwood S.A. 26S promoter for use when the alphavirus is Girdwood S.A. Another preferred promoter is the TR339 26S promoter for use when the alphavirus is TR339. Additional promoters beyond the Girdwood S.A. and TR339

promoters include the VEE 26S promoter, the Sindbis 26S promoter, the Semliki Forest 26S promoter, and any other promoter sequence recognized by alphavirus polymerases. Alphavirus promoter sequences containing mutations which alter the activity level of the promoter (in relation to the activity level of the wild-type) are also suitable in the practice of the present invention. Such mutant promoter sequences are described in Raju and Huang, *J. Virol.* 65, 2501-2510 (1991), the disclosure of which is incorporated herein in its entirety. In the system wherein the first helper RNA, the second helper RNA, and the replicon RNA are all on separate molecules, the promoters, if the same promoter is used for all three RNAs, provide a homologous sequence between the three molecules. It is preferred that the selected promoter is operative with the non-structural proteins encoded by the replicon RNA molecule.

In cases where vaccination with two immunogens provides improved protection against disease as compared to vaccination with only a single immunogen, a double-promoter replicon would ensure that both immunogens are produced in the same cell. Such a replicon would be the same as the one described above, except that it would contain two copies of the 26S RNA promoter, each followed by a different multiple cloning site, to allow for the insertion and expression of two different heterologous proteins. Another useful strategy is to insert the IRES sequence from the picornavirus, EMC virus, between the two heterologous genes downstream from the single 26S promoter of the replicon described above, thus leading to expression of two immunogens from the single replicon transcript in the same cell.

#### C. Uses of the Present Invention.

The alphavirus vectors, RNAs, cDNAs, helper cells, infectious virus particles, and methods of the present invention find use in *in vitro* expression systems, wherein the inserted heterologous RNA encodes a protein or peptide which is desirably produced *in vitro*. The RNAs, cDNAs, helper cells, infectious virus particles, methods, and pharmaceutical formulations of the present invention are additionally useful in a method of administering a protein or peptide to a

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subject in need of the protein or peptide, as a method of treatment or otherwise. In this embodiment of the invention, the heterologous RNA encodes the desired protein or peptide, and pharmaceutical formulations of the present invention are administered to a subject in need of the desired protein or peptide. In this manner,  
5 the protein or peptide may thus be produced *in vivo* in the subject. The subject may be in need of the protein or peptide because the subject has a deficiency thereof, or because the production of the protein or peptide in the subject may impart some therapeutic effect, as a method of treatment or otherwise.

Alternately, the claimed methods provide a vaccination strategy,  
10 wherein the heterologous RNA encodes an immunogenic protein or peptide.

The methods and products of the invention are also useful as antigens and for evoking the production of antibodies in animals such as horses and rabbits, from which the antibodies may be collected and then used in diagnostic assays in accordance with known techniques.

15 A further aspect of the present invention is a method of introducing and expressing antisense oligonucleotides in bone marrow cell cultures to regulate gene expression. Alternately, the claimed method finds use in introducing and expressing a protein or peptide in bone marrow cell cultures.

## II. Girdwood S.A. and TR339 Clones.

20 Disclosed hereinbelow are genomic RNA sequences encoding live Girdwood S.A. virus, live S.A.AR86 virus, and live Sindbis strain TR339 virus, cDNAs derived therefrom, infectious RNA transcripts encoded by the cDNAs, infectious viral particles containing the infectious RNA transcripts, and pharmaceutical formulations derived therefrom.

25 The cDNA sequence of Girdwood S.A. is given herein as SEQ ID NO:4. Alternatively, the cDNA may have a sequence which differs from the cDNA of SEQ ID NO:4, but which has the same protein sequence as the cDNA

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given herein as SEQ ID NO:4. Thus, the cDNA may include one or more silent mutations.

The phrase "silent mutation" as used herein refers to mutations in the cDNA coding sequence which do not produce mutations in the corresponding protein sequence translated therefrom.

Likewise, the cDNA sequence of TR339 is given herein as SEQ ID NO:8. Alternatively, the cDNA may have a sequence which differs from the cDNA of SEQ ID NO:8, but which has the same protein sequence as the cDNA given herein as SEQ ID NO:8. Thus, the cDNA may include one or more silent mutations.

The cDNAs encoding infectious Girdwood S.A. and TR339 virus RNA transcripts of the present invention include those homologous to, and having essentially the same biological properties as, the cDNA sequences disclosed herein as SEQ ID NO:4 and SEQ ID NO:8, respectively. Thus, cDNAs that hybridize to cDNAs encoding infectious Girdwood S.A. or TR339 virus RNA transcripts disclosed herein are also an aspect of this invention. Conditions which will permit other cDNAs encoding infectious Girdwood S.A. or TR339 virus transcripts to hybridize to the cDNAs disclosed herein can be determined in accordance with known techniques. For example, hybridization of such sequences may be carried out under conditions of reduced stringency, medium stringency, or even high stringency conditions (*e.g.*, conditions represented by a wash stringency of 35-40% formamide with 5X Denhardt's solution, 0.5% SDS and 1X SSPE at 37°C; conditions represented by a wash stringency of 40-45% formamide with 5X Denhardt's solution, 0.5% SDS, and 1X SSPE at 42°C; and conditions represented by a wash stringency of 50% formamide with 5X Denhardt's solution, 0.5% SDS and 1X SSPE at 42°C, respectively, to cDNA encoding infectious Girdwood S.A. or TR339 virus RNA transcripts disclosed herein in a standard hybridization assay. *See* J. SAMBROOK ET AL., MOLECULAR CLONING: A LABORATORY MANUAL (2d ed. 1989)). In general, cDNA sequences encoding infectious

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Girdwood S.A. or TR339 virus RNA transcripts that hybridize to the cDNAs disclosed herein will be at least 30% homologous, 50% homologous, 75% homologous, and even 95% homologous or more with the cDNA sequences encoding infectious Girdwood S.A. or TR339 virus RNA transcripts disclosed herein.

Promoter sequences and Girdwood S.A. virus or Sindbis virus strain TR339 cDNA clones are operatively associated in the present invention such that the promoter causes the cDNA clone to be transcribed in the presence of an RNA polymerase which binds to the promoter. The promoter is positioned on the 5' end (with respect to the virion RNA sequence), of the cDNA clone. An excessive number of nucleotides between the promoter sequence and the cDNA clone will result in the inoperability of the construct. Hence, the number of nucleotides between the promoter sequence and the cDNA clone is preferably not more than eight, more preferably not more than five, still more preferably not more than three, and most preferably not more than one.

Examples of promoters which are useful in the cDNA sequences of the present invention include, but are not limited to T3 promoters, T7 promoters, cytomegalovirus (CMV) promoters, and SP6 promoters. The DNA sequence of the present invention may reside in any suitable transcription vector. The DNA sequence preferably has a complementary DNA sequence bound thereto so that the double-stranded sequence will serve as an active template for RNA polymerase. The transcription vector preferably comprises a plasmid. When the DNA sequence comprises a plasmid, it is preferred that a unique restriction site be provided 3' (with respect to the virion RNA sequence) to the cDNA clone. This provides a means for linearizing the DNA sequence to allow the transcription of genome-length RNA *in vitro*.

The cDNA clones can be generated by any of a variety of suitable methods known to those skilled in the art. A preferred method is the method set forth in United States Patent No. 5,185,440 to Davis et al., the disclosure of which

is incorporated in its entirety by reference, and Gubler et al., *Gene* 25:263 (1983).

RNA is preferably synthesized from the DNA sequence *in vitro* using purified RNA polymerase in the presence of ribonucleotide triphosphates and cap analogs in accordance with conventional techniques. However, the RNA may also be synthesized intracellularly after introduction of the cDNA.

The Girdwood S.A. and TR339 cDNA clones and the infectious RNAs and infectious virus particles produced therefrom of the present invention are useful for the preparation of pharmaceutical formulations, such as vaccines. In addition, the cDNA clones, infectious RNAs, and infectious viral particles of the present invention are useful for administration to animals for the purpose of producing antibodies to the Girdwood S.A. virus or the Sindbis virus strain TR339, which antibodies may be collected and used in known diagnostic techniques for the detection of Girdwood S.A. virus or Sindbis virus strain TR339. Antibodies can also be generated to the viral proteins expressed from the cDNAs disclosed herein. As another aspect of the present invention, the claimed cDNA clones are useful as nucleotide probes to detect the presence of Girdwood S.A. or TR339 genomic RNA or transcripts.

### III. Infectious Virus Particles and Pharmaceutical Formulations.

The infectious virus particles of the present invention include those containing double promoter vectors and those containing replicon vectors as described hereinabove. Alternately, the infectious virus particles contain infectious RNAs encoding the Girdwood S.A. or TR339 genome. When the infectious RNA comprises the Girdwood S.A. genome, preferably the RNA has the sequence encoded by the cDNA given as SEQ ID NO:4. When the infectious RNA comprises the TR339 genome, preferably the RNA has the sequence encoded by the cDNA given as SEQ ID NO:8.

The infectious, alphavirus particles of the present invention may be prepared according to the methods disclosed herein in combination with techniques

known to those skilled in the art. These methods include transfecting an alphavirus-permissive cell with a replicon RNA including the alphavirus packaging segment and an inserted heterologous RNA, a first helper RNA including RNA encoding at least one alphavirus structural protein, and a second helper RNA including RNA encoding at least one alphavirus structural protein which is different from that encoded by the first helper RNA. Alternately, and preferably, at least one of the helper RNAs is produced from a cDNA encoding the helper RNA and operably associated with an appropriate promoter, the cDNA being stably transfected and integrated into the cells. More preferably, all of the helper RNAs will be "launched" from stably transfected cDNAs. The step of transfecting the alphavirus-permissive cell can be carried out according to any suitable means known to those skilled in the art, as described above with respect to propagation-competent viruses.

Uptake of propagation-competent RNA into the cells *in vitro* can be carried out according to any suitable means known to those skilled in the art. Uptake of RNA into the cells can be achieved, for example, by treating the cells with DEAE-dextran, treating the RNA with LIPOFECTIN® before addition to the cells, or by electroporation, with electroporation being the currently preferred means. These techniques are well known in the art. *See e.g.*, United States Patent No. 5,185,440 to Davis et al., and PCT Publication No. WO 92/10578 to Bioption AB, the disclosures of which are incorporated herein by reference in their entirety. Uptake of propagation-competent RNA into the cell *in vivo* can be carried out by administering the infectious RNA to a subject as described in Section I above.

The infectious RNAs may also contain a heterologous RNA segment, where the heterologous RNA segment contains a heterologous RNA and a promoter operably associated therewith. It is preferred that the infectious RNA introduces and expresses the heterologous RNA in bone marrow cells as described in Section I above. According to this embodiment, it is preferable that the promoter operatively associated with the heterologous RNA is operable in bone

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marrow cells. The heterologous RNA may encode any protein or peptide, preferably an immunogenic protein or peptide, a therapeutic protein or peptide, a hormone, a growth factor, an interleukin, a cytokine, a chemokine, an enzyme, a ribozyme, or an antisense oligonucleotide as described in more detail in Section I above.

The step of facilitating the production of the infectious viral particles in the cells may be carried out using conventional techniques. *See e.g.*, United States Patent No. 5,185,440 to Davis et al., PCT Publication No. WO 92/10578 to Bioption AB, and United States Patent No. 4,650,764 to Temin et al. (although Temin et al., relates to retroviruses rather than alphaviruses). The infectious viral particles may be produced by standard cell culture growth techniques.

The step of collecting the infectious virus particles may also be carried out using conventional techniques. For example, the infectious particles may be collected by cell lysis, or collection of the supernatant of the cell culture, as is known in the art. *See e.g.*, United States Patent No. 5,185,440 to Davis et al., PCT Publication No. WO 92/10578 to Bioption AB, and United States Patent No. 4,650,764 to Temin et al. Other suitable techniques will be known to those skilled in the art. Optionally, the collected infectious virus particles may be purified if desired. Suitable purification techniques are well known to those skilled in the art.

Pharmaceutical formulations, such as vaccines, of the present invention comprise an immunogenic amount of the infectious, virus particles in combination with a pharmaceutically acceptable carrier. An "immunogenic amount" is an amount of the infectious virus particles which is sufficient to evoke an immune response in the subject to which the pharmaceutical formulation is administered. An amount of from about  $10^3$  to about  $10^7$  particles, and preferably about  $10^4$  to  $10^6$  particles per dose is believed suitable, depending upon the age and species of the subject being treated, and the immunogen against which the immune response is desired.

Pharmaceutical formulations of the present invention for therapeutic use comprise a therapeutic amount of the infectious virus particles in combination with a pharmaceutically acceptable carrier. A "therapeutic amount" is an amount of the infectious virus particles which is sufficient to produce a therapeutic effect (e.g., triggering an immune response or supplying a protein to a subject in need thereof) in the subject to which the pharmaceutical formulation is administered. The therapeutic amount will depend upon the age and species of the subject being treated, and the therapeutic protein or peptide being administered. Typical dosages are an amount from about  $10^1$  to about  $10^5$  infectious units.

Exemplary pharmaceutically acceptable carriers include, but are not limited to, sterile pyrogen-free water and sterile pyrogen-free physiological saline solution. Subjects which may be administered immunogenic amounts of the infectious virus particles of the present invention include but are not limited to human and animal (e.g., pig, cattle, dog, horse, donkey, mouse, hamster, monkeys) subjects.

Pharmaceutical formulations of the present invention include those suitable for parenteral (e.g., subcutaneous, intracerebral, intradermal, intramuscular, intravenous and intraarticular) administration. Alternatively, pharmaceutical formulations of the present invention may be suitable for administration to the mucus membranes of a subject (e.g., intranasal administration by use of a dropper, swab, or inhaler). The formulations may be conveniently prepared in unit dosage form and may be prepared by any of the methods well known in the art.

The following examples are provided to illustrate the present invention, and should not be construed as limiting thereof. In these examples, PBS means phosphate buffered saline, EDTA means ethylene diamine tetraacetate, ml means milliliter,  $\mu$ l means microliter, mM means millimolar,  $\mu$ M means micromolar, u means unit, PFU means plaque forming units, g means gram, mg means milligram,  $\mu$ g means microgram, cpm means counts per minute, ic means

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intracerebral or intracerebrally, ip means intraperitoneal or intraperitoneally, iv means intravenous or intravenously, and sc means subcutaneous or subcutaneously.

Amino acid sequences disclosed herein are presented in the amino to carboxyl direction, from left to right. The amino and carboxyl groups are not presented in the sequence. Nucleotide sequences are presented herein by single strand only in the 5' to 3' direction, from left to right. Nucleotides and amino acids are represented herein in the manner recommended by the IUPAC-IUB Biochemical Nomenclature Commission, or (for amino acids) by either one letter or three letter code, in accordance with 37 CFR § 1.822 and established usage. Where one letter amino acid code is used, the same sequence is also presented elsewhere in three letter code.

#### EXAMPLE I

##### Cells and Virus Stocks

S.A.AR86 was isolated in 1954 from a pool of *Culex* sp. mosquitoes collected near Johannesburg, South Africa. Weinbren et al., *S. Afr. Med. J.* 30, 631-36 (1956). Ockelbo82 was isolated from *Culiseta* sp. mosquitoes collected in Edsbyn, Sweden in 1982 and was associated serologically with human disease. Niklasson et al., *Am. J. Trop. Med. Hyg.* 33, 1212-17 (1984). Girdwood S.A. was isolated from a human patient in the Johannesburg area of South Africa in 1963. Malherbe et al., *S. Afr. Med. J.* 37, 547-52 (1963). Molecularly cloned virus TR339 represents the deduced consensus sequence of Sindbis AR339. McKnight et al., *J. Virol.* 70, 1981-89 (1996); William Klimstra, personal communication. TRSB is a laboratory strain of Sindbis isolate AR339 derived from a cDNA clone pTRSB and differing from the AR339 consensus sequence at three codons. McKnight et al., *J. Virol.* 70, 1981-89 (1996). pTR5000 is a full-length cDNA clone of Sindbis AR339 following the SP6 phage promoter and containing mostly Sindbis AR339 sequences.

Stocks of all molecularly cloned viruses were prepared by electroporating genome length *in vitro* transcripts of their respective cDNA clones

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in BHK-21 cells. Heidner et al., *J. Virol.* 68, 2683-92 (1994). Girdwood S.A. (Malherbe et al., *S. Afr. Med. J.* 37, 547-52 (1963)) and Ockelbo82 (Espmark and Niklasson, *Am. J. Trop. Med. Hyg.* 33, 1203-11 (1984); Niklasson et al., *Am. J. Trop. Med. Hyg.* 33, 1212-17 (1984)) were passed one to three times in BHK-21  
5 cells in order to produce amplified stocks of virus. All virus stocks were stored at -70°C until needed. The titers of the virus stocks were determined on BHK-21 cells from aliquots of frozen virus.

## EXAMPLE 2

### Cloning the S.A.AR86 and Girdwood S.A. Genomic Sequences

10 The sequences of S.A.AR86 (Figure 1, SEQ ID NO: 1) and Girdwood S.A. (Figure 3, SEQ ID NO:4) were determined from uncloned reverse transcriptase-polymerase chain reaction (RT-PCR) fragments amplified from virion RNA. Heidner et al., *J. Virol.* 68, 2683-92 (1994). The sequence of the 5' 40 nucleotides was determined by directly sequencing the genomic RNA. Sanger et al., *Proc. Natl. Acad. Sci. USA* 74, 5463-67 (1977); Zimmern and Kaesberg, *Proc. Natl. Acad. Sci. USA* 75, 4257-61 (1978); Ahlquist et al., *Cell* 23, 183-89  
15 (1981).

The S.A.AR86 genome was 11,663 nucleotides in length, excluding the 5' CAP and 3' poly(A) tail, 40 nucleotides shorter than the alphavirus prototype  
20 Sindbis strain AR339. Strauss et al., *Virology* 133, 92-110 (1984). Compared with the consensus sequence of Sindbis virus AR339 (McKnight et al., *J. Virol.* 70 1981-89 (1996)), S.A.AR86 contained two separate 6-nucleotide insertions, and one 3-nucleotide insertion in the 3' half of the nsP3 gene, a region not well conserved among alphaviruses. The two 6-nucleotide insertions were found  
25 immediately 3' of nucleotides 5403 and 5450, and the 3-nucleotide insertion was immediately 3' of nucleotide 5546 compared with the AR339 genome. In addition, S.A.AR86 contained a 54-nucleotide deletion in nsP3 which spanned nucleotides 5256 to 5311 of AR339. As a result of these deletions and insertions, S.A.AR86 nsP3 was 13 amino acids smaller than AR339, containing an 18-amino acid  
30 deletion and a total of 5 amino acids inserted. The 3' untranslated region of

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S.A.AR86 contained, with respect to AR339, two 1-nucleotide deletions at nucleotides 11,513 and 11,602, and one 1-nucleotide insertion following nucleotide 11,664. The total numbers of nucleotides and predicted amino acids comprising the remaining genes of S.A.AR86 were identical to those of AR339.

5                   A notable feature of the deduced amino acid sequence of S.A.AR86 (Figure 2, SEQ ID NO:2 and SEQ ID NO:3) was the cysteine codon in place of an opal termination codon between nsP3 and nsP4. S.A.AR86 is the only alphavirus of the Sindbis group, and one of just three alphavirus isolates sequenced to date, which do not contain an opal termination codon between nsP3 and nsP4.  
10       Takkinen, K., *Nucleic Acids Res.* 14, 5667-5682 (1986); Strauss et al., *Virology* 164, 265-74 (1988).

                  The genome of Girdwood S.A. was 11,717 nucleotides long excluding the 5' CAP and 3' poly(A) tail. The nucleotide sequence (SEQ ID NO:4) of the Girdwood S.A. genome and the putative amino acid sequence (SEQ  
15       ID NO:5 and SEQ ID NO:6) of the Girdwood S.A. gene products are shown in Figure 3 and Figure 4, respectively. The asterisk at position 1902 in SEQ ID NO:5 indicates the position of the opal termination codon in the coding region of the nonstructural polyprotein. The extra nucleotides relative to AR339 were in the nonconserved half of nsP3, which contained insertions totalling 15 nucleotides, and  
20       in the 3' untranslated region which contained two 1-nucleotide deletions and a 1-nucleotide insertion with respect to the consensus Sindbis AR339 genome. The insertions found in the nsP3 gene of Girdwood S.A. were identical in position and content to those found in S.A.AR86, although Girdwood S.A. did not have the large nsP3 deletion characteristic of S.A.AR86. The remaining portions of the  
25       genome contained the same number of nucleotides and predicted amino acids as Sindbis AR339.

                  Overall, Girdwood S.A. was 94.5% identical to the consensus Sindbis AR339 sequence, differing at 655 nucleotides not including the insertions and deletions. These nucleotide differences resulted in 88 predicted amino acid

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changes or a difference of 2.3%. A plurality of amino acid differences were concentrated in the nsP3 gene, which contained 32 of the amino acid changes, 25 of which were in the nonconserved 3' half.

The Girdwood S.A. nucleotides at positions 1, 3, and 11,717 could not be resolved. Because the primer used during the RT-PCR amplification of the 3' end of the genome assumed a cytosine in the 3' terminal position, the identity of this nucleotide could not be determined with certainty. However, in all alphaviruses sequenced to date there is a cytosine in this position. This, combined with the fact that no difficulty was encountered in obtaining RT-PCR product for this region with an oligo(dT) primer ending with a 3'G, suggested that Girdwood S.A. also contains a cytosine at this position. The ambiguity at nucleotide positions 1 and 3 resulted from strong stops encountered during the RNA sequencing.

### EXAMPLE 3

#### Comparison of S.A.AR86 and Girdwood S.A.

#### Sequences With Other Sindbis-Related Virus Sequences

Table 1 examines the relationship of S.A.AR86 and Girdwood S.A. to each other and to other Sindbis-related viruses. This was accomplished by aligning the nucleotide and deduced amino acid sequences of Ockelbo82, AR339 and Girdwood S.A. to those of S.A.AR86 and then calculating the percentage identity for each gene using the programs contained within the Wisconsin GCG package (Genetics Computer Group, 575 Science Drive, Madison WI 53711), as described in more detail in McKnight et al., *J. Virol.* 70, 1981-89 (1996).

The analysis suggests that S.A.AR86 is most similar to the other South African isolate, Girdwood S.A., and that the South African isolates are more similar to the Swedish Ockelbo82 isolate than to the Egyptian Sindbis AR339 isolate. These results also suggest that it is unlikely that S.A.AR86 is a recombinant virus like WEE virus. Hahn et al., *Proc. Natl. Acad. Sci. USA* 85, 5997-6001 (1988).

TABLE 1  
Comparison of the Nucleotide and Amino Acid Sequences  
of S.A.AR86 Virus with Those of Sindbis AR339, Ockelbo82, and Girdwood S.A. Viruses<sup>a</sup>

Regions	Nucleotide Differences <sup>b</sup>			Amino Acid Differences <sup>b</sup>		
	AR339	Ock82	GIRD	AR339	Ock82	GIRD
	Number (%)			Number (%)		
5' untranslated	0 (0.0)	0 (0.0)	1 (1.7)	--	--	--
nsP1	76 (4.7)	37 (2.3)	15 (0.9)	9 (1.7)	6 (1.1)	2 (0.4)
nsP2	137 (5.7)	86 (3.6)	45 (1.9)	15 (1.9)	8 (1.0)	12 (1.5)
nsP3						
Conserved <sup>c</sup>	51 (5.7)	35 (3.9)	13 (1.6)	6 (2.0)	1 (0.3)	1 (0.4)
Nonconserved <sup>d</sup>	116 (6.6)	83 (4.4)	70 (2.2)	45 (9.7)	34 (7.0)	27 (3.7)
nsP4	111 (6.1)	68 (3.7)	19 (1.1)	8 (1.3)	2 (0.3)	4 (0.6)
26s junction	1 (2.1)	0 (0.0)	1 (2.1)	--	--	--
Capsid	36 (4.5)	26 (3.3)	7 (0.9)	1 (0.4)	3 (1.1)	0 (0.0)
E3	17 (8.9)	5 (2.6)	4 (2.1)	1 (1.6)	0 (0.0)	0 (0.0)
E2	71 (5.6)	43 (3.4)	18 (1.4)	12 (2.6)	6 (1.4)	2 (0.5)
6K	10 (6.1)	9 (5.4)	4 (2.4)	2 (3.6)	2 (3.6)	1 (1.8)
E1	49 (3.7)	31 (2.3)	16 (1.2)	7 (1.6)	6 (1.4)	2 (0.9)
3' untranslated	14 (4.5)	8 (2.5)	1 (0.3)	--	--	--
Totals	689 (5.5)	431 (3.3)	214 (1.4)	106 (2.3)	68 (1.4)	51 (0.9)

a. All nucleotide positions and gene boundaries are numbered according to those used for the Sindbis AR339, HR<sub>10</sub> variant Genebank Accession No. J02363; Strauss et al., *Virology* 133, 92-110 (1984).

b. Differences include insertions and deletions.

c. Conserved region nucleotides 4100 to 5000 (aa 1 to aa300).

d. Nonconserved region nucleotides 5001 to 5729 (aa301 to aa542, S.A.AR86 numbering).

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## EXAMPLE 4

Neurovirulence of S.A.AR86 and Girdwood S.A.

Girdwood S.A., Ockelbo82, and S.A.AR86 are related by sequence; in contrast, it has previously been reported that only S.A.AR86 displayed the adult mouse neurovirulence phenotype. Russell et al., *J. Virol.* 63, 1619-29 (1989). These findings were confirmed by the present investigations. Briefly, groups of four female CD-1 mice (3-6 weeks of age) were inoculated ic with  $10^3$  plaque-forming units (PFU) of S.A.AR86, Girdwood S.A., or Ockelbo82. Neither Girdwood S.A. nor Ockelbo82 infection produced any clinical signs of infection. Infection with S.A.AR86 produced neurological signs within four to five days and ultimately killed 100% of the mice as previously demonstrated.

Table 2 lists those amino acids of S.A.AR86 which might explain the neurovirulence phenotype in adult mice. A position was scored as potentially related to the S.A.AR86 adult neurovirulence phenotype if the S.A.AR86 amino acid differed from that which otherwise was absolutely conserved at that position in the other viruses.

TABLE 2

Divergent Amino Acids in S.A.AR86  
Potentially Related to the Adult Neurovirulence Phenotype

	Position in S.A.AR86	S.A.AR86 Amino Acid	Conserved Amino Acid
nsP1	583	Thr	Ile
nsP2	256	Arg	Ala
	648	Ile	Val
	651	Lys	Glu
nsP3	344	Gly	Glu
	386	Tyr	Ser
	441	Asp	Gly
	445	Ile	Met
	537	Cys	Opal
E2	243	Ser	Leu
6K	30	Val	Ile
E1	112	Val	Ala
	169	Leu	Ser

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## EXAMPLE 5

pS55 Molecular Clone of S.A.AR86

As a first step in investigating the unique adult mouse neurovirulence phenotype of S.A.AR86, a full-length cDNA clone of the S.A.AR86 genome was constructed. The sources of cDNA included conventional cDNA clones (Davis et al., *Virology* 171, 189-204 (1989)) as well as uncloned RT-PCR fragments derived from the S.A.AR86 genome. As described previously, these were substituted, starting at the 3' end, into pTR5000 (McKnight et al., *J. Virol.* 70, 1981-89 (1996)), a full-length Sindbis clone from which infectious genomic replicas could be derived by transcription with SP6 polymerase *in vitro*.

The end result was pS55, a molecular clone of S.A.AR86 from which infectious transcripts could be produced and which contained four nucleotide changes (G for A at nt 215; G for C at nt 3863; G for A at nt 5984; and C for T at nt 9113) but no amino acid coding differences with respect to the S.A.AR86 genomic RNA (amino acid sequence of S.A.AR86 presented in Figure 2 (SEQ ID NO:2 and SEQ ID NO:3)). The nucleotide sequence of clone pS55 is presented in Figure 5 (SEQ ID NO:7).

As has been described by Simpson et al., *Virology* 222, 464-69 (1996), neurovirulence and replication of the virus derived from pS55 (S55) were compared with those of S.A.AR86. It was found that S55 exhibits the distinctive adult neurovirulence characteristic of S.A.AR86. Like S.A.AR86, S55 produces 100% mortality in adult mice infected with the virus and the survival times of animals infected with both viruses were indistinguishable. In addition, S55 and S.A.AR86 were found to replicate to essentially equivalent titers *in vivo*, and the profiles of S55 and S.A.AR86 virus growth in the central nervous system and periphery were very similar.

From these data it was concluded that the silent changes found in virus derived from clone pS55 had little or no effect on its growth or virulence, and that this molecularly cloned virus accurately represents the biological isolate, S.A.AR86.

## EXAMPLE 6

Construction of the Consensus AR339 Virus TR339

The consensus sequence of the Sindbis virus AR339 isolate, the prototype alphavirus was deduced. The consensus AR339 sequence was inferred by comparison of the TRSB sequence (a laboratory-derived AR339 strain) with the complete or partial sequences of HR<sub>p</sub> (the Gen Bank sequence; Strauss et al., *Virology* 133, 92-110 (1984)), SV1A, and NSV (AR339-derived laboratory strains; Lustig et al., *J. Virol* 62, 2329-36 (1988)), and SIN (a laboratory-derived AR339 strain; Davis et al., *Virology* 161, 101-108 (1987), Strauss et al., *J. Virol.* 65, 4654-64 (1991)). Each of these viruses was descended from AR339. Where these sequences differed from each other, they also were compared with the amino acid sequences of other viruses related to Sindbis virus: Ockelbo82, S.A.AR86, Girdwood S.A., and the somewhat more distantly related Aura virus. Rumenapf et al., *Virology* 208, 621-33 (1995).

The details of determining a consensus AR339 sequence and constructing the consensus virus TR339 have been described elsewhere. McKnight et al., *J. Virol.* 70, 1981-89 (1996); Klimstra et al., *manuscript in preparation*. The nucleotide (SEQ ID NO:8) sequence of pTR339 is presented in Figure 6. The deduced amino acid sequences of the pTR339 non-structural and structural polyproteins are shown as SEQ ID NO:9 and SEQ ID NO:10, respectively. The asterisk at position 1897 in SEQ ID NO:9 indicates the position of the opal termination codon in the coding region of the nonstructural polyprotein. The consensus nucleotide sequence diverged from the pTRSB sequence at three coding positions (nsP3 528, E2 1, and E1 72). These differences are illustrated in Table 3.

TABLE 3

Amino Acid Differences Between  
Laboratory Strain TRSB and Molecular Clone TR339

	nsP3 528 (nt5683)	E2 1 (nt8633)	E1 72 (nt10279)
TR339	Arg (CGA)	Ser (AGC)	Ala (GCU)
TRSB	Gln (CAA)	Arg (AGA)	Val (GUU)

## EXAMPLE 7

Animals Used for *In Vivo* Localization Studies

Specific pathogen free CD-1 mice were obtained from Charles River Breeding Laboratories (Raleigh, North Carolina) at 21 days of age and maintained under barrier conditions until approximately 37 days of age. Intracerebral (ic) inoculations were performed as previously described, Simpson et al., *Viol.* 222, 464-49 (1996), with 500 PFU of S51 (an attenuated mutant of S55) or  $10^3$  PFU of S55. Animals inoculated peripherally were first anesthetized with METOFANE®. Then, 25  $\mu$ l of diluent (PBS, pH 7.2, 1% donor calf serum, 100 u/ml penicillin, 50  $\mu$ g/ml streptomycin, 0.9 mM  $\text{CaCl}_2$ , and 0.5 mM  $\text{MgCl}_2$ ) containing  $10^3$  PFU of virus were injected either intravenously (iv) into the tail vein, subcutaneously (sc) into the skin above the shoulder blades on the middle of the back, or intraperitoneally (ip) in the lower right abdomen. Animals were sacrificed at various times post-inoculation as previously described. Simpson et al., *Viol.* 222, 464-49 (1996). Brains (including brainstems) were homogenized in diluent to 30% w/v, and right quadriceps were homogenized in diluent to 25% w/v. Homogenates were handled and titered as described previously. Simpson et al., *Viol.* 222, 464-49 (1996). Bone marrow was harvested by crushing both femurs from each animal in sufficient diluent to produce a 30% w/v suspension (calculated as weight of uncrushed femurs in volume of diluent). Samples were stored at  $-70^\circ\text{C}$ . For titration, samples were thawed and clarified by centrifugation at  $1,000 \times g$  for 20 minutes at  $4^\circ\text{C}$  before being titered by conventional plaque assay on BHK-21 cells.

## EXAMPLE 8

Tissue Preparation for *In Situ* Hybridization Studies

Animals were anesthetized by ip injection of 0.5 ml AVERTIN® at various times post-inoculation followed by perfusion with 60 to 75 ml of 4% paraformaldehyde in PBS (pH 7.2) at a flow rate of 10 ml per minute. The entire carcass was decalcified for 8 to 10 weeks in 4% paraformaldehyde containing 8% EDTA in PBS (pH 6.8) at  $4^\circ\text{C}$ . This solution was changed twice during the decalcification period. Selected tissues were cut into blocks approximately 3 mm thick and placed into biopsy cassettes for paraffin embedding and sectioning. Blocks were embedded, sectioned and hematoxylin/eosin stained by Experimental Pathology Laboratories (Research Triangle Park, North Carolina) or North

Carolina State University Veterinary School Pathology Laboratory (Raleigh, North Carolina).

### EXAMPLE 9

#### In Situ Hybridization

5 Hybridizations were performed using a [<sup>35</sup>S]-UTP labeled S.A.AR86 specific riboprobe derived from pDS-45. Clone pDS-45 was constructed by first amplifying a 707 base pair fragment from pS55 by PCR using primers 7241 (5'-CTGCGGCGGATTCATCTTGC-3', SEQ ID NO:11) and SC-3 (5'-CTCCAACTTAAGTG-3', SEQ ID NO:12). The resulting 707 base pair fragment  
10 was purified using a GENE CLEAN® kit (Bio101, CA), digested with *Hha*I, and cloned into the *Sma*I site of pSP72 (Promega). Linearizing pDS-45 with *Eco*RV and performing an *in vitro* transcription reaction with SP6 DNA-dependent, RNA polymerase (Promega) in the presence of [<sup>35</sup>S]-UTP resulted in a riboprobe approximately 500 nucleotides in length of which 445 nucleotides were  
15 complementary to the S.A.AR86 genome (nucleotides 7371 through 7816). A riboprobe specific for the influenza strain PR-8 hemagglutinin (HA) gene was used as a control probe to test non-specific binding. The *in situ* hybridizations were performed as described previously (Charles et al., *Viol.* 208, 662-71 (1995)) using 10<sup>5</sup> cpm of probe per slide.

### EXAMPLE 10

#### Replication of S.A.AR86 in Bone Marrow

20 Three groups of six adult mice each were inoculated peripherally (sc, ip, or iv) with 1200 PFU of S55 (a molecular clone of S.A.AR86) in 25 µl of diluent. Under these conditions, the infection produced no morbidity or  
25 mortality. Two mice from each group were anesthetized and sacrificed at 2, 4 and 6 days post-inoculation by exsanguination. The serum, brain (including brainstem), right quadricep, and both femurs were harvested and titered by plaque assay. Virus was never detected in the quadricep samples of animals inoculated  
30 sc (Table 4). A single animal inoculated ip (two days post-inoculation) and two mice inoculated iv (at four and six days post-inoculation) had detectable virus in the right quadricep, but the titer was at or just above the limit of detection (6.25 PFU/g tissue). Virus was present sporadically or at low levels in the brain and

serum of animals regardless of the route of inoculation. Virus was detected in the bone marrow of animals regardless of the route of inoculation. However, the presence of virus in bone marrow of animals inoculated sc or ip was more sporadic than animals inoculated iv, where five out of six animals had detectable virus.

5 These results suggest that S55 targets to the bone marrow, especially following iv inoculation.

The level and frequency of virus detected in the serum and muscle suggested that virus detected in the bone marrow was not residual virus contamination from blood or connective tissue remaining in bone marrow samples.

10 The following experiment also suggested that virus in bone marrow was not due to tissue or serum contamination. Mice were inoculated ic with 1200 PFU of S55 in 25  $\mu$ l of diluent. Animals were sacrificed at 0.25, 0.5, 1, 1.5, 2, 3, 4, 5, and 6 days post-inoculation, and the carcasses were decalcified as described in Example 8. Coronal sections taken at approximately 3 mm intervals through the  
15 head, spine (including shoulder area), and hips were probed with an S55-specific [<sup>35</sup>S]-UTP labeled riboprobe derived from pDS-45. Positive *in situ* hybridization signal was detected by one day post-inoculation in the bone marrow of the skull (data not shown). Weak signal also was present in some of the chondrocytes of the vertebrae, suggesting that S55 was replicating in these cells as well. Although  
20 the frequency of positive bone marrow cells was low, the signal was very intense over individual positive cells. This result strongly suggests that S55 replicates *in vivo* in a subset of cells contained in the bone marrow.

### EXAMPLE 11

#### Other Sindbis Group Viruses

25 It was of interest to determine if the ability to replicate in the bone marrow of mice was unique to S55 or was a general feature of other viruses, both Sindbis and non-Sindbis viruses, in the Sindbis group. Six 38-day-old female CD-1 mice were inoculated iv with 25  $\mu$ l of diluent containing 10<sup>3</sup> PFU of S55, Ockelbo82, Girdwood S.A., TR339, or TRSB. At 2, 4 and 6 days post-  
30 inoculation two mice from each group were sacrificed and whole blood, serum, brain (including brainstem), right quadricep, and both femurs were harvested for virus titration.

The results of this experiment were similar to those with S55. TRSB infected animals had no virus detectable in serum or whole blood in any animal at any time, and with the other viruses tested, no virus was detected in the serum or whole blood of any animal beyond two days post-inoculation (detection limit, 25 PFU/ml). Neither TRSB nor TR339 was detectable in the brains of infected animals at any time post-inoculation. S55, Girdwood S.A., and Ockelbo82 were present in the brains of infected animals sporadically with the titers being at or near the 75 PFU/g level of detection. All the tested viruses were found sporadically at or slightly above the 50 PFU/g detection limit in the right quadricep of infected animals except for a single animal four days post-inoculation with TRSB which had nearly  $10^5$  PFU/g of virus in its quadricep.

The frequency at which the different viruses were detected in bone marrow varied widely, with S55 and Girdwood S.A. being the most frequently isolated (five out of six animals) and Ockelbo82 and TRSB being the least frequently isolated from bone marrow (one out of six animals and two out of six animals, respectively) (Table 4). Girdwood S.A. and S55 gave nearly identical profiles in all tissues. Girdwood S.A., unlike S.A.AR86, is not neurovirulent in adult mice (Example 4), suggesting that the adult neurovirulence phenotype is distinct from the ability of the virus to replicate efficiently in bone marrow.

TABLE 4  
Titers Following IV Inoculation of Virus

Virus	Tissue Titered							
	Animal	Days Post-Inoculation	Bone Marrow (PFU/g)	Serum (PFU/ml)	Blood (PFU/ml)	Brain (PFU/g)	Quadriceps (PFU/g)	
S55	A	2	1125	N.D.*	N.D.	N.D.	N.D.	
	B		488	50	200	N.D.	N.D.	
	A	4	863	N.D.	N.D.	N.D.	550	
	B		113	N.D.	N.D.	75	N.D.	
	A	6	N.D.	N.D.	N.D.	N.D.	50	
	B		37.5	N.D.	N.D.	N.D.	N.D.	
	Limit of Detection		37.5	25	25	75	50	
	TR339	A	2	N.D.	N.D.	N.D.	N.D.	N.D.
		B		1500	75	700	N.D.	N.D.
		A	4	1050	N.D.	N.D.	N.D.	N.D.
B		1762		N.D.	N.D.	N.D.	400	
A		6	N.D.	N.D.	N.D.	N.D.	N.D.	
B			N.D.	N.D.	N.D.	N.D.	N.D.	
Limit of Detection			37.5	25	25	37.5	50	
TR5B		A	2	N.D.	N.D.	N.D.	N.D.	N.D.
		B		N.D.	N.D.	N.D.	N.D.	N.D.
		A	4	150	N.D.	N.D.	N.D.	1000
	B	N.D.		N.D.	N.D.	N.D.	100000	
	A	6		N.D.	N.D.	N.D.	N.D.	N.D.
	B		37.5	N.D.	N.D.	N.D.	N.D.	
	Limit of Detection		37.5	25	25	37.5	50	

TABLE 4 Continued  
Titers Following IV Inoculation of Virus

Virus	Animal	Days Post-Inoculation	Tissue Titered				
			Bone Marrow (PFU/g)	Serum (PFU/ml)	Blood (PFU/ml)	Brain (PFU/g)	Quadriceps (PFU/g)
Girdwood S.A.	A	2	22000	2325	1450	300	50
	B		2500	1200	2600	N.D.	N.D.
	A	4	788	N.D.	N.D.	N.D.	N.D.
	B		113	N.D.	N.D.	75	N.D.
	A	6	N.D.	N.D.	N.D.	N.D.	N.D.
	B		75	N.D.	N.D.	1700	N.D.
	Limit of Detection		37.5	25	25	75	50
	A	2	N.D.	125	150	N.D.	N.D.
Ockelbo82	B		N.D.	50	500	N.D.	200
	A	4	N.D.	N.D.	N.D.	300	N.D.
	B		300	N.D.	N.D.	N.D.	N.D.
	A	6	N.D.	N.D.	N.D.	100000	N.D.
	B		N.D.	N.D.	N.D.	N.D.	N.D.
	Limit of Detection		37.5	25	25	75	50

\* "N.D." indicates that the virus titers were below the limit of detection.

## EXAMPLE 12

Virus Persistence in Bone Marrow

The next step in our investigations was to evaluate the possibility that S.A.AR86 persisted long-term in bone marrow. S51 is a molecularly cloned, attenuated mutant of S55. S51 differs from S55 by a threonine for isoleucine substitution at amino acid residue 538 of nsP1 and is attenuated in adult mice inoculated intracerebrally. Like S55, S51 targeted to and replicated in the bone marrow of 37-day-old female CD-1 mice following ic inoculation. Mice were inoculated ic with 500 PFU of S51 and sacrificed at 4, 8, 16, and 30 days post-inoculation for determination of bone marrow and serum titers. At no time post-inoculation was virus detected in the serum above the 6.25 PFU/ml detection limit. Virus was detectable in the bone marrow samples of both animals sampled at four days post-inoculation and in one animal eight days post-inoculation (Table 5). No virus was detectable by titration on BHK-21 cells in any of the bone marrow samples beyond eight days post-inoculation. These results suggested that the attenuating mutation present in S51, which reduces the neurovirulence of the virus, did not impair acute viral replication in the bone marrow.

It was notable that the plaque size on BHK-21 cells of virus recovered on day 4 post-inoculation was smaller than the size of plaques produced by the inoculum virus, and that plaques produced from virus recovered from the day 8 post-inoculation samples were even smaller and barely visible. This suggests a strong selective pressure in the bone marrow for virus that is much less efficient in forming plaques on BHK-21 cells.

To demonstrate that S51 virus genomes were present in bone marrow cells long after acute infection, four to six-week-old female CD-1 mice were inoculated ic with 500 PFU of S51. Three months post-inoculation two animals were sacrificed, perfused with paraformaldehyde and decalcified as described in Example 8. The heads and hind limbs from these animals were paraffin embedded, sectioned, and probed with a S.A.AR86 specific [<sup>35</sup>S]-UTP labeled riboprobe derived from clone pDS-45. *In situ* hybridization signal was clearly present in discrete cells of the bone and bone marrow of the legs (data not shown). Furthermore, no *in situ* hybridization signal was detected in an adjacent

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control section probed with an influenza virus HA gene specific riboprobe. As the relative sensitivity of *in situ* hybridization is reduced in decalcified tissues (Peter Charles, personal communication), these cells likely contain a relatively high number of viral sequences, even at three months post-inoculation. No *in situ* hybridization signal was observed in mid-sagittal sections of the heads with the S.A.AR86 specific probe, although focal lesions were observed in the brain indicative of the prior acute infection with S51.

TABLE 5

S51 Titers in Bone Marrow Following IC Inoculation of 500 PFU			
Days Post-Inoculation	Titers (Total PFU/Animal)		Limit of Detection
	Animal A	Animal B	
4	2100	380	62.5
8	62.5	N.D. <sup>a</sup>	62.5
16	N.D.	N.D.	62.5
30	N.D.	N.D.	62.5

<sup>a</sup> "N.D." indicates that the virus titers were below the limit of detection.

## Example 13

Replication of S.A.A.R86 within Bone/Joint Tissue of Adult Mice

Several old world alphaviruses, including Ross River Virus, Chikungunya virus, Okelbo82, and S.A.AR86 are associated with acute and persistent  
5 arthritis/arthralgia in humans. Molecular clones of several Sindbis group viruses, including S.A.AR86, were used to investigate alphavirus replication within bone/joint tissue.

Following intravenous inoculation of S.A.AR86 into adult CD-1 mice, viral replication was observed in bone/joint tissue, but not surrounding muscle tissue of  
10 the hind limbs. Infectious virus was detectable 24 hrs post-infection; however, viral titer within bone/joint tissue was maximal 72 hours post-infection. Fractionation of hind limbs from infected animals revealed that the hip and knee joints were the predominant sites of viral replication. Replication within bone/joint tissue appears to be a common trait of Sindbis-group viruses, since the laboratory strains TR339 and TRSB  
15 also replicated within bone/joint tissue. *In situ* hybridization and S.A.AR86 based double promoter vectors expressing green fluorescent protein were used to further localize S.A.AR86 infected cells within bone/joint tissue. Green fluorescent protein expression was detected in bone/joint tissue for at least one month post-inoculation. These studies demonstrated that cells within the endosteum of synovial joints were the  
20 predominant site of S.AAR86 replication.

SEQUENCE LISTINGS

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## THAT WHICH IS CLAIMED IS:

1. A method of introducing and expressing heterologous RNA in bone marrow cells, comprising:

(a) providing a recombinant alphavirus, said alphavirus containing a heterologous RNA segment, said heterologous RNA segment comprising a promoter operable in said bone marrow cells operatively associated with a heterologous RNA to be expressed in said bone marrow cells; and then

(b) contacting said recombinant alphavirus to said bone marrow cells so that said heterologous RNA segment is introduced and expressed therein.

2. A method according to claim 1, wherein said contacting step is carried out *in vitro*.

3. A method according to claim 1, wherein said contacting step is carried out *in vivo* in a subject in need of such treatment.

4. A method according to claim 1, wherein said heterologous RNA encodes a protein or peptide.

5. A method according to claim 1, wherein said heterologous RNA encodes an immunogenic protein or peptide.

6. A method according to claim 1, wherein said heterologous RNA encodes an antisense oligonucleotide or a ribozyme.

7. A method according to claim 1, wherein said alphavirus is an Old World alphavirus.

8. A method according to claim 1, wherein said alphavirus is selected from the group consisting of SF group and SIN group alphaviruses.

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9. A method according to claim 1, wherein said alphavirus is selected from the group consisting of Semliki Forest virus, Middelburg virus, Chikungunya virus, O'Nyong-Nyong virus, Ross River virus, Barmah Forest virus, Getah virus, Sagiyama virus, Bebaru virus, Mayaro virus, Una virus, Sindbis virus, South African Arbovirus No. 86, Ockelbo virus, Girdwood S.A. virus, Aura virus, Whataroa virus, Babanki virus, and Kyzylagach virus.

10. A method according to claim 1, wherein said alphavirus is South African Arbovirus No. 86.

11. A method according to claim 1, wherein said alphavirus is Girdwood S.A.

12. A method according to claim 1, wherein said alphavirus is Sindbis strain TR339.

13. A helper cell for expressing an infectious, propagation defective, Girdwood S.A. virus particle, comprising, in a Girdwood S.A.-permissive cell:

(a) a first helper RNA encoding (i) at least one Girdwood S.A. structural protein, and (ii) not encoding at least one other Girdwood S.A. structural protein; and

(b) a second helper RNA separate from said first helper RNA, said second helper RNA (i) not encoding said at least one Girdwood S.A. structural protein encoded by said first helper RNA, and (ii) encoding said at least one other Girdwood S.A. structural protein not encoded by said first helper RNA, and with all of said Girdwood S.A. structural proteins encoded by said first and second helper RNAs assembling together into Girdwood S.A. particles in said cell containing said replicon RNA;

and wherein the Girdwood S.A. packaging segment is deleted from at least said first helper RNA.

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14. The helper cell according to claim 13, further containing a replicon RNA;

said replicon RNA encoding said Girdwood S.A. packaging segment and an inserted heterologous RNA;

5 wherein said Girdwood S.A. packaging segment is deleted from at least one of said helper RNA;

and wherein said replicon RNA, said first helper RNA, and said second helper RNA are all separate molecules from one another.

15. The helper cell according to claim 13, further containing a replicon RNA;

said replicon RNA encoding said Girdwood S.A. packaging segment and an inserted heterologous RNA;

wherein said replicon RNA and said first helper RNA are separate molecules;

15 and wherein the molecule containing said replicon RNA further contains RNA encoding said at least one Girdwood S.A. structural protein not encoded by said first helper RNA.

16. The helper cell according to claim 13, wherein said first helper RNA encodes both the Girdwood S.A. E1 glycoprotein and the Girdwood S.A. E2 glycoprotein, and wherein said second helper RNA encodes the Girdwood S.A. capsid protein.

17. A method of making infectious, propagation defective, Girdwood S.A. virus particles, comprising:

25 transfecting a Girdwood S.A.-permissive cell according to claim 13 with a propagation defective replicon RNA, said replicon RNA including said Girdwood S.A. packaging segment and an inserted heterologous RNA;

producing said Girdwood S.A. virus particles in said transfected cell; and then

collecting said Girdwood S.A. virus particles from said cell.

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18. Infectious Girdwood S.A. virus particles produced by the method of Claim 17.

19. Infectious Girdwood S.A. virus particles containing a replicon RNA encoding a promoter, an inserted heterologous RNA, and wherein  
5 RNA encoding at least one Girdwood S.A. structural protein is deleted therefrom so that said Girdwood S.A. virus particle is propagation defective.

20. A pharmaceutical formulation comprising infectious Girdwood S.A. virus particles according to claim 18 or 19 in a pharmaceutically acceptable carrier.

10 21. A helper cell for expressing an infectious, propagation defective, TR339 virus particle, comprising, in a TR339-permissive cell:

(a) a first helper RNA encoding (i) at least one TR339 structural protein, and (ii) not encoding at least one other TR339 structural protein; and

(b) a second helper RNA separate from said first helper RNA,  
15 said second helper RNA (i) not encoding said at least one TR339 structural protein encoded by said first helper RNA, and (ii) encoding said at least one other TR339 structural protein not encoded by said first helper RNA, and with all of said TR339 structural proteins encoded by said first and second helper RNAs assembling together into TR339 particles in said cell containing said replicon  
20 RNA;

and wherein the TR339 packaging segment is deleted from at least said first helper RNA.

22. The helper cell according to claim 21, further containing a replicon RNA;

25 said replicon RNA encoding said TR339 packaging segment and an inserted heterologous RNA;

wherein said TR339 packaging segment is deleted from at least one of said helper RNA;

and wherein said replicon RNA, said first helper RNA, and said  
30 second helper RNA are all separate molecules from one another.

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23. The helper cell according to claim 21, further containing a replicon RNA;

said replicon RNA encoding said TR339 packaging segment and an inserted heterologous RNA;

5 wherein said replicon RNA and said first helper RNA are separate molecules;

and wherein the molecule containing said replicon RNA further contains RNA encoding said at least one TR339 structural protein not encoded by said first helper RNA.

10 24. The helper cell according to claim 21, wherein said first helper RNA encodes both the TR339 E1 glycoprotein and the TR339 E2 glycoprotein, and wherein said second helper RNA encodes the TR339 capsid protein.

15 25. A method of making infectious, propagation defective, TR339 virus particles, comprising:

transfecting a TR339-permissive cell according to claim 21 with a propagation defective replicon RNA, said replicon RNA including said TR339 packaging segment and an inserted heterologous RNA;

20 producing said TR339 virus particles in said transfected cell; and then

collecting said TR339 virus particles from said cell.

26. Infectious TR339 virus particles produced by the method of Claim 25.

25 27. Infectious TR339 virus particles containing a replicon RNA encoding a promoter, an inserted heterologous RNA, and wherein RNA encoding at least one TR339 structural protein is deleted therefrom so that said virus particle is propagation defective.

28. A pharmaceutical formulation comprising infectious TR339 virus particles according to Claim 26 or 27 in a pharmaceutically acceptable carrier.

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29. A recombinant DNA comprising a cDNA coding for an infectious Girdwood S.A. virus RNA transcript and a heterologous promoter positioned upstream from said cDNA and operatively associated therewith.

5 30. An infectious RNA transcript encoded by a cDNA according to claim 29.

31. An infectious RNA according to claim 30, said infectious Girdwood S.A. RNA transcript containing a heterologous RNA segment, said heterologous RNA segment comprising a promoter operably associated with a heterologous RNA.

10 32. Infectious viral particles containing an RNA transcript according to claim 30.

33. A recombinant DNA comprising a cDNA coding for a Sindbis strain TR339 RNA transcript and a heterologous promoter positioned upstream from said cDNA and operatively associated therewith.

15 34. An infectious RNA transcript encoded by a cDNA according to claim 33.

20 35. An infectious RNA according to claim 34, said infectious Girdwood S.A. RNA transcript containing a heterologous RNA segment, said heterologous RNA segment comprising a promoter operably associated with a heterologous RNA.

36. Infectious viral particles containing an RNA transcript according to claim 34.

## Nucleotide Sequence of S.A.AR86

1 ATTGGCGGCG TAGTACACAC TATTGAATCA AACAGCCGAC CAATTGCACT ACCATCACA TGGAGAAGCC AGTAGTTAAC GTAGACGTAG ACCCTCAGAG  
101 TCCGTTTGTG GTGCAACTGC AAAAGAGCTT CCCGCAATT GAGGTAGTAG CACAGCAGGT CACTCCAAAT GACCATGCTA ATGCCAGAGC ATTTTCCGAT  
201 CTGGCCAGTA AACTAATCGA GCTGGAGGTT CCTACCACAG CGACGATTTT GGACATAGGC AGCGCACCAG CTCGTAGAAT GTTTTCCGAG CACCAAGTACC  
301 ATTGCGTTTG CCCCATGCTT AGTCCAGAA GACCGGACCG CATGATGAAA TATGCCAGCA AACTGGCGGA AAAAGCATGT AAGATTACAA ACAAGAACTT  
401 GCATGAGAA GATCAAGGACC TCCGGACCGT ACTTGATACA CCGGATGCTG AAACGCCATC ACTCTGCTTC CACAACGATG TTACTTGCAA CACGCGTGCC  
501 GAGTACTCCG TCATGCAGGA CGGTACATC AACGCTCCCG GAATATTTA CCACCAGGCT ATGAAAGGCG TCCGGACCCT GTACTGGATT GGTTCGACA  
601 CCACCCAGTT CATGTTCTCG GCTATGGCAG GTTCGTACCC TGCATACAAC ACCAATCGG CCGACGAAAA AGTCCTTGA GCGCGTAACA TCGGACTCTG  
701 CAGCACAAG CTGAGTGAAG GCAGGACAGG AAAGTTGTCT ATAATGAGGA AGAAGGAGTT GAAGCCCGGG TCACGGGTTT ATTTCTCCGT TGGATCGACA  
801 CTTTACCAG AACACAGAGC CAGCTTGCA GCTGCGATC TTCCATCGGT GTTCCACTTG AAAGGAAAGC AGTCGTACAC TTGCGGCTGT GATACAGTGG  
901 TGAGCTCGA AGGCTACGTA GTGAAGAAA TCACCATCAG TCCCGGGATC ACGGGAGAAA CCGTGGGATA CGCGGTACA AACAAAGCG AGGGCTTCTT  
1001 GCTATGCAA GTTACCGATA CAGTAAAGG AGAACGGTA TCGTCCCCG TGTGCAGTA TATCCCCGCC ACCATATGCG ATCAGATGAC CGGCATAATG  
1101 GCCACGGATA TCTCACCTGA CGATGCACAA AAATCTCTGG TTGGGTCAA CCAGCGAATC GTCATTAACG GTAAGACTAA CAGGAACACC AATACCATGC  
1201 AAAATTACCT TCTGCCAATC ATTGCACAAG GGTTCAGCAA ATGGGCCAAG GACGCGAAG AAGATCTTGA CAATGAAAA ATGCTGGGCA CCGAGAGGGG  
1301 CAAGCTTACA TATGGCTGCT TGTGGCGTT TCGCACTAAG AAAGTCACT CGTTCTATCG CCCACCTGGA ACGCAGACCA TCGTAAAGT CCCAGCTCT  
1401 TTTAGCGCTT TCCCATGTC ATCCGTATGG ACTACCTCTT TGCCCATGTC GCTGAGGCAG AAGATGAAAT TGGCATTACA ACCAAAAGAG GAGGAAAAAC  
1501 TGCTGCAAGT CCCGGAGGAA TTAGTTATGG AGGCCAAGC TGCTTCGAG GATGCTCAGG AGGAATCCAG AGCGGAGAA GCTCCGAGAAG CACTCCCACC  
1601 ATTAGTGGCA GACAAAGGTA TCGAGGCAGC TCGGGAAGTT GTCTCGAAG TGGAGGGGCT CCAGGCGGAC ACCGAGCAG CACTCGTCGA AACCCCGCGC  
1701 GGTGATGTA GGATAATACC TCAAGCAAT GACCGTATGA TCGGACAGTA TATCGTTGTC TCGCCGATCT CTGTGCTGAA GAACGCTAAA CTCGACCCAG  
1801 CACACCCGCT AGCAGACCAG GTTAAGATCA TAACGCATC CGGAAGATCA GGAAGGTATG CAGTCGAACC ATACGACGCT AAAGTACTGA TGCCAGCAGG  
1901 AAGTGCCGTA CCATGGCCAG AATTCTTAGC ACTGAGTGAG AGCGCCACGC TTGTGTACAA CGAAAGAGAG TTTGTGAACC GCAAGCTGTA CCATATTGCC  
2001 ATGCACGTC CCGTAAGAA TACAGAAGAG GAGCAGTACA AGGTTACAAA GGCAGAGCTC GCAGAAACAG AGTACGTGTT TGACGTGGAC AAGAAGCGAT  
2101 GCGTTAAGAA GGAAGAAGCC TCAGGACTTG TCCTTTCGGG AGAAGTGACC AACCCGCCCT ATCAGGAACT AGCTCTTGAG GGAAGTGAAG CTCGACCCGC  
2201 GGTCCCGTAC AAGGTTGAAA CAATAGGAGT GATAGGCACA CCAGGATCGG GCAAGTCAGC TATCATCAAG TCAACTGTCA CGGCACGTGA TCTGTTACC  
2301 AGCGGAAAG AAGAAAAGT CCGGAAATG GAGGCCGAGC TGCTACGGCT GAGGGGATG CAGATCACGT CGAAGACAGT GGATTCGGTT ATGCTCAAGC  
2401 GATGCCACAA AGCCGTAGAA GTGCTGTATG TTGACGAAGC GTTCCGGTGC CACGACGAG CACTACTTGC CTGATTGCA ATGTCAGAC CCCGTAAGAA  
2501 GGTAGTACTA TCGGAGAGCC CTAAGCAATG CGGATTCTC AACATGATG AACTAAAGGT ACATTTCAAC CACCCTGAAA AAGACATATG TACCAAGACA  
2601 TTCTACAAAT TTATCTCCC ACGTTGCACA CAGCAGTCA CGGCTATTG ATGCACTG CATTACGATG GAAAAAGTAA AACCAAAAC CCGTGCAAGA  
2701 AGAACATCGA AATCGACATT ACAGGGGCGA CGAAGCGGAA GCCAGGGGAC ATCATCTGA CATGTTCCG CCGGTGGGTT AAGCAACTGC AAATCGACTA  
2801 TCCCGGACAT GAGGTAATGA CAGCCGCGG CTCACAAGG CTAACAGAA AAGGAGTATA TGCCGTCCG CAAAAAGTCA ATGAAAAACC GGTGACGGC  
2901 ATCACATCAG AGCATGTGA CGTGTGCTC ACCCGCACTG AGGACAGGCT AGTATGAAA ACTTTACAG GCGACCCATG GATTAAGCAG CTCCTAACG  
3001 TACCTAAAGG AAATTTTCA GGCACCATCG AGGACTGGGA AGCTGAACAC AAGGGAATAA TTGCTGCGAT AAACAGTCCC GCTCCCCGTA CCAATCCGTT  
3101 CAGCTGCAAG ACTAACGTT GCTGGCGAA AGCACTGGAA CCGATACTG CCACGGCCCG TATCGTACTT ACCGTTGCC AGTGAGCGA GGTGTTCCCA  
3201 CAGTTTGGG ATGACAAACC ACACTCGGCC ATCTACGCT TAGACGTAAT TTGCATTAAG TTTTTCGGCA TGGACTGAC AAGCGGCTG TTTTCCAAAC  
3301 AGAGCATCCC GTTAACGTAC CATCTCCCG ACTCAGCGAG GCCAGTAGCT CATTGGGACA ACAGCCAGG AACACGCAAG TATGGGTACG ATCAGCCGCT  
3401 TGCCCGCGAA CTCCTCCGTA GATTCCCGT GTTCCAGTA GCTGGGAAAG GCACACAGCT TGATTGCGAG ACGGGCAGAA CTAGATTAT CTCTGCACAG  
3501 CATAACTGG TCCAGTGAA CCGCAATCT CTCACGCTT TAGTCCCCG GCACAAGGAG AAACAACCCG GCGCGTGA AAAATTCTTG AGCCAGTTCA  
3601 AACACCACTC CGTACTGTG ATCTCAGAGA AAAAAATGA AGTCCCCAC AAGAGAATCG AATGGATCG CCGGATTGG ATAGCCCGCG CAGATAAGAA  
3701 CTACAACCTG GCTTTCGGT TTCCGCGCA GGCAGGTAC GACCTGGTGT TCATCAATAT TGGAACTAAA TACAGAAACC ATCACTTCA ACAGTGCGAA

Fig. 1A

3801 GACCACGGCG CGACCTTGAA AACCTTTTCG CGTTCCGGCC TGAAGTCCCT TAACCCCGGA GGCACCTCG TGGTGAAGTC CTACGGTTAC GCGACCGCA  
3901 ATAGTGAGGA CGTAGTCACC GCTCTTGCCA GAAATTTGT CAGAGTGCTC GCAGCGAGGC CAGAGTGCGT CTCAAGCAAT ACAGAAATGT ACCTGATTTT  
4001 CCGACAACCTA GACAACAGCC GCACACGACA ATTCACCCCG CATCATTTGA ATTGTGTGAT TTCGTCCGTG TACGAGGGTA CAAGACAGCG AGTTGGAGCC  
4101 GCACCGTCTG ACCGTACTAA AAGGGAGAAC ATTGCTGATT GTCAGAGGA AGCAGTTGTC AATGCAGCCA ATCCACTGGG CAGACCAGGA GAAGGAGTCT  
4201 GCCGTGCCAT CTATAAACGT TGGCCGAACA GTTTCACCGA TTCAGCCACA GAGACAGGTA CCGCAAACT GACTGTGTGC CAAGGAAAGA AAGTGATCCA  
4301 CGCGTTGGC CCGTATTTC GGAACACCC AGAGGCAGAA GCGCTGAAT TGCTGCAAAA CGCTACCAT GCAGTGGCAG ACTTAGTAAA TGAACATAAT  
4401 ATCAAGTCTG TCGCCATCCC ACTGCTATCT ACAGGCATTG ACGCAGCCG AAAAGACCG CTGAGGTAT CACTTAACTG CTGACAAAC GCGTAGACA  
4501 GAACTGATGC GGACGTAACC ATCTACTGCC TGGATAAGAA GTGGAAGGAA AGAATCGACG CGGTGCTCCA ACTTAAGGAG TCTGTAACTG AGCTGAAGGA  
4601 TGAGGATATG GAGATCGACG ACGAGTTAGT ATGGATCCAT CCGGACAGTT GCTGAAGGG AAGAAAGGGA TTCAGTACTA CAAAAGGAAA GTTGTATTCG  
4701 TACTTTGAAG GCACAAAAT CCATCAAGCA GCAAAAGATA TGGCGGAGAT AAAGGTCTG TTCCAAAATG ACCAGGAAAG CAACGAACAA CTGTGTCCCT  
4801 ACATATTGGG GGAGACCATG GAAGCAATCC GCGAAAAATG CCGGTGCGC CACAACCGT CGTCTAGGCC GCCAAAAACG CTGCCGTGCC TGTGTATGTA  
4901 TGCCATGACG CCAGAAAGGG TCCACAGACT CAGAAGCAAT AACGCAAAAG AAGTTACAGT ATGCTCTCC ACCCCCCCTT CAAAGTACAA AATCAAGAAT  
5001 GTTCAGAAAG TTCAGTGAC AAAAGTAGTC CTGTTAAACC CGCATACCC CGCATTCGTT CCCGCCGTA AGTACATAGA AGCACCAGAA CAGCCTGCAG  
5101 CTCGCCCTGC ACAGGCCGAG GAGGCCCGG GAGTTGTAGC GACACCAACA CCACCTGCAG CTGATAACAC CTCGCTTGAT GTCACGGACA TCTCACTGGA  
5201 CATGGAAGAC AGTAGCGAAG GCTCACTCTT TTCGAGCTTT AGCGGATCGG ACAACTACCG AAGGCAGGTG GTGGTGGCTG ACGTCCATGC CGTCAAGAG  
5301 CTTGCCCTG TTCACCGCC AAGGCTAAG AAGATGGCC GCTGCGCAG GCAAGAATG CAGGAAGAGC CAATCCACC GGAAGCACC AGCTGTCCGG  
5401 ACGAGTCCCT TCACCTTCT TTTGATGGG TATCTATAT CTCGGATCC CTTTTCGAGC GAGAGATGG CCGTTGGCA GCGGCACAA CCCCCGAAAG  
5501 TACATGCCCT ACGGATGTGC CTATGTCTT CGGATCGTT TCCGACGGAG AGATTGAGGA GTTGAGCCGC AGAGTAAACG AGTCGGAGCC CGTCTGTTT  
5601 GGGTCATTG AACCGGGCA AGTGAACCTA ATTATATCGT CCGGATCAGC CGTATCTTT CCACCACGCA AGCAGAGACG TAGACGCAGG AGCAGGAGGA  
5701 CCGAATAGTG TCTAACCGG GTAGGTGGGT ACATATTTT GACGGACACA GGCCTGGGC ACTTGCAAAA GAAGTCCGT CTGCAGAAC AGCTTACAGA  
5801 ACCGACCTTG GAGCGCAATG TTTGGAAG AATCTACGCC CCGGTGCTG ACAGTTCGAA AGAGGAACAG CTCAACTCA GGTACCAGAT GATGCCACCC  
5901 GAAGCCAACA AAAGCAGGTA CAGTCTCGA AAAGTAGAAA ACCAGAAAGC CATAACCACT GAGCGACTGC TTTAGGGGT ACGACTGTAT AACTCTGCCA  
6001 CAGATCAGCC AGAATGCTAT AAGATCACC ACCCGAAACC ATCGTATTCC AGCAGTGAT CAGCGAACTA CTCTGACCCA AAGTTTGCTG TAGCTGTTG  
6101 TAACAACAT CTGCATGAGA ATTACCGAC GGTAGCATCT TATCAGATCA CCGACGAGTA CGATGCTTAC TTGGATATGG TAGACGGGAC AGTCGCTGC  
6201 CTAGATACTG CAATTTTTT CCGCGCAAG CTAGAAGTT ACCCGAAAAG ACACGAGTAT AGAGCCCCAA ACATCCGCAG TCGGGTTCCA TCAGCGATGC  
6301 AGAACACGTT GCAAAACGTC CTCATTGCCG CGACTAAAAG AAAGTCAAC GTCACACAAA TGGCTGAAT GCCAACACTG GACTCAGCGA CATTCAACGT  
6401 TGAATGCTTT CGAAAAATG CATGCAATGA CGAGTATTGG GAGGAGTTG CCGAAAGCC AATTAGGATE ACTACTGAGT TCGTTACCCG ATACGTGGCC  
6501 AGACTGAAAG GCGTAAGGC CGCGCACTG TTCGCAAGA CGCATAATT GGTCCATTG CAAGAAGTGC CTATGGATAG ATTCGTCATG GACATGAAAA  
6601 GAGACGTGAA AGTTACACCT GGCACGAAAC ACACAGAAGA AAGACCGAAA GTACAAGTGA TACAAGCCGC AGAACCCCTG GCGACCGCTT ACCTATGCGG  
6701 GATCCACCGG GAGTTAGTGC GCAGGCTTAC AGCCGTTTTG CTACCCAACA TTCACACGCT CTTTGACATG TCGCGGAGG ACTTTGATGC AATCATAGCA  
6801 GAACACTTCA AGCAAGGTGA CCGGTACTG GAGACGGATA TCGCTCGTT CGACAAAAGC CAAGACGACG CTATGGCGTT AACCGGCTG ATGATCTGG  
6901 AAGACCTGGG TGTGGACCAA CCACTACTCG ACTTGATCGA GTGCGCTTT GGAGAAATAT CATCCACCCA TCTGCCACG GGTACCGTT TCAATTCCG  
7001 GCGGATGATG AAATCCGGA TTTCTCTCAC GCTCTTTGTC AACACAGTTC TGAATGCTG TATCGCCAGC AGAGTATTGG AGGAGCGGT TAAACGTTCC  
7101 AAATGTGCG CATTATCCG CGACGACAAC ATTATACAGC GAGTAGTATC TGACAAAGAA ATGGCTGAGA GGTGTCCAC CTGGCTCAAC ATGGAGGTTA  
7201 AGATCATTGA CGCAGTATC GCGAGAGAC CACTTACTT CTGCGGTGGA TTCATCTGC AAGATTCCGT TACCTCCACA GCGTGTCCG TGGCGGACCC  
7301 CTTGAAAGG CTGTTTAAAT TGGTAAACC GCTCCAGCC GACGATGAGC AAGACGAAGA CAGAAGACGC GCTCTGCTAG ATGAAACAAA GCGCTGTTT  
7401 AGAGTAGGTA TAACAGACAC CTTAGCAGTG CCGTGCGCAA CTCGTATGA GGTAGACAAC ATCACACCTG TCTGCTGCC ATTGAGAACT TTTGCCAGA  
7501 GCAAAAGAGC ATTCAAGCC ATCAGAGGGG AAATAAGCA TCTCTACGT GTCTCTAAT AGTCAGCATA GTACATTCA TCTGACTAAT ACCACAACAC  
7601 CACCACATG AATAGAGGAT TCTTTAATCAT GCTCGGCCG CGCCCTTCC CAGCCCCAC TGCCATGTG AGGCCGCGGA GAAGGAGGCA GCGCGCCCG  
7701 ATGCTGCCG GCAATGGGT GGTCTCCAA ATCCAGCAAC TGACCACAGC CGTCAGTCC CTAGTCATTG GACAGGCAAC TAGACCTCA ACCCCACGCC  
7801 CACGCCCGCC CCGCGCCAG AAGAAGCAG CGCCAAAGCA ACCACCGAAG CCGAAGAAAC CAAAACACA GGAGAAGAAG AAGAAGCAAC CTGCAAAACC

Fig. 1B

7901 CAAACCCCGA AAGAGACAGC GTATGGCACT TAAGTTGGAG GCCGACAGAC TGTTCGACGT CAAAAATGAG GACGGAGATG TCATCGGCCA CGCACTGGCC  
8001 ATGGAAGGAA AGGTAATGAA ACCACTCCAC GTGAAAGGAA CTATTGACCA CCTGTGCTA TCAAAAGCTCA AATTCACCAA GTCTCAGCA TACGACATGG  
8101 AGTTCGCACA GTTGCCGGTC AACATGAGAA GTGAGGGGTT CACCTACACC AGTGAACACC CTGAAGGGTT CTACAACCTG CACCACGGAG CGGTGCACTA  
8201 TAGTGGAGGC AGATTATCCA TCCCCCGCG AGTAGGAGGC AGAGGAGACA GTGTCTCTCC GATTATGGAT AACTCAGGCC GGGTTGTCCG GATAGTCCTC  
8301 GGAGGGGCTG ATGAGGGAAC AAGAACCACC CTTTCGGTCC TCACCTGGAA TAGCAAAAGG AAGACAATCA AGACAACCCC GGAAGGGACA GAAGAGTGGT  
8401 CTGTGCACC ACTGGTCAGC GCCATGTGCT TGCTTGGAAA CGTGAGCTTC CCATGCAATC GCGCGCCAC ATCTACACC CGCGAACCTC CCAGAGCTCT  
8501 CGACATCCTC GAAGAGAACC TGAACCACGA GGCCTACGAC ACCCTGCTCA ACGCCATATT GCGGTGCGGA TCCTCCGGCA GAAGTAAAG AAGCTCACT  
8601 GACGACTTTA CCTTGACCAG CCCGTAAGTT GGCACATGCT CGTACTGTCA CCATACTGAA CGGTGCTTTA GCGCGATTAA GATCGAGCAG GTCTGGGATG  
8701 AAGCGGACGA CAACACCATA CGCATAAGCA CTTCGGCCCA GTTTGGATAC GACCAAGGCG GAGCAGCAAG CTCAAATAAG TACCGCTACA TGTCGCTGGA  
8801 CGAGGATCAT ACTGTCAAAG AAGGCACCAT GGTGACATC AAGATCAGCA CCTCAGGACC GTGTAGAAGG CTTAGCTACA AAGGATACTT TCTCTCGCG  
8901 AAGTGTCTCC CAGGGGACAG CGTAACGGTT AGCATAGCGA GTAGCAACTC AGCAACGTCA TGCACAATGG CCCGCAAGAT AAAACCAAAA TTCTGGGAG  
9001 GGGAAAAATA TGACCTACCT CCCGTTACG GTAAGAAAGT TCCTTGACCA GTGTACGACC GTCTGAAAGA AACAAACCGC GGCTACATCA CTATGCACAG  
9101 GCGGGGACCG CATGCTTATA CATCTATCT GGAGGAATCA TCAGGGAAAG TTTACCGGAA GCCACCATCC GGAAGAACA TTACGTACGA GTGCAAGTGC  
9201 GCGGATTACA AGACCGGAAC CGTTACGACC CGTACCGAAA TCACGGGCTG CACCGCCATC AAGCAGTGGC TCGCCTATAA GAGCGACCAA ACGAAGTGGG  
9301 TCTTCAACTC GCGGAGCTCG ATCAGACAGC CCGACCACAC GGCCEAAGG AAATTGCATT TGCCTTTCAA GCTGATCCCG AGTACCTGCA TGTCCTCTGT  
9401 TGCCCAACCG CCGAACGTAG TACACGGCTT TAAACACATC AGCCTCCAAT TAGACACAGA CCATCTGACA TTGCTACCA CCAGGAGACT AGGGGCAAA  
9501 CCGGAACCAA CCACTGAATG GATCATCGGA AACACGGTTA GAACTTCAC CCGCAGCCGA GATGGCCTGG AATACATATG GGGCAATCAC GAACCACTAA  
9601 GGGTCTATGC CCAAGAGTCT GCACCAGGAG ACCCTCAGCG ATGGCCACAC GAAATAGTAC AGCATTACTA TCATCGCCAT CCGTGTACA CCATCTTAGC  
9701 CGTCGCATCA GCTGCTGTGG CGATGATGAT TGGCGTAACT GTTGACGAT TATGTGCTG TAAAGCGCGC CGTGAGTGCC TGACGCCATA TGCCCTGGCC  
9801 CCAATGCCG TGATTCCAAC TTGCTGGCA CTTTGTGCT GTGTAGGTC GGCTAATGCT GAAACATTCA CCGAGACCAT GAGTTACTTA TGTGCAACA  
9901 GCCAGCGCTT CTCTGGGTC CAGCTGTGTA TACCTGTGC CGCTGTGTC GTTCTAATGC GCTGTGCTC ATGCTGCTG CTTTCTTTAG TGGTGGCGG  
10001 CGCCTACCTG GCGAAGGTAG ACGCCTACGA ACATGCGACC ACTGTCCAA ATGTGCCACA GATACCGTAT AAGGCACCTG TTGAAAGGGC AGGGTACGGC  
10101 CCGCTCAATT TGGAGATTAC TGTCATGTCC TCGGAGGTTT TGCTTCCAC CAACCAAGAG TACATTACCT GCAATTCAC CACTGTGTC CCTCCCCA  
10201 AAGTCAGATG CTGCGGCTCC TTGGAATGTC AGCCCGCCGC TCACGCAGAC TATACCTGCA AGGTCTTTGG AGGGGTGTAC CCCTTCATGT GGGGAGGAGC  
10301 ACAATGTTTT TGGACAGTGA AGAACAGCCA GATGAGTGAG GCGTACGTCG AATTGTCACT AGATTGCGCG ACTGACCACG CGCAGCGCAT TAAGGTGCAT  
10401 ACTGCGCGGA TGAAGTAGG ACTGCTATA GTGTACGGGA AACTACACAG TTTCTAGAT GTGTACGTGA ACGGAGTCAC ACCAGGAACG TCTAAAGACC  
10501 TGAAGTCAAT AGCTGGACCA ATTTAGCAT TGTATACACC ATTCGATCAC AAGTCTGTTA TCAATCGCGG CCTGGTGTAC AACTATGACT TTCCGGAATA  
10601 CGGAGCGATG AAACCAGGAG CGTTTGAGA CATCAAGCT ACCTCTTGA CTAGCAAAGA CCTCATCGCC AGCAGACAGA TTAGGCTACT CAAGCCTTC  
10701 GCCAAGAACG TGCATGTCCC GTACACGCA GCGCATCTG GATTCGAGAT GTGGAAAAAC AACTCAGGCC GCCCACTGCA GGAACCGCC CTTTGTGGT  
10801 GCAAGATTGC AGTCAATCCG CTTGAGCGG TGGACTGCTC ATACGGGAAC ATTECCATT CTATTGACAT CCGGAACGCT GCCTTTATCA GGACATCAGA  
10901 TGCACCACTG GTCTAACAG TCAAATGTGA TGTCAGTGA TGCACTATT CAGCGGACTT CGGAGGGATG GCTACCTGC AGTATGTATC CGACCGCGAA  
11001 GGACAATGCC CTGTACATTC GATTCGAGC ACAGCAACCC TCCAAGATC GACAGTTCAT GTCTGGAGA AAGGAGCGGT GACAGTACAC TTCAGCACCG  
11101 CGAGCCACA GCGCAACTTC ATTGTATCG TGTGTGTAA GAAGACAACA TGCAATGCAG AATGCAAAAC ACCAGCTGAT CATATCTGA GCACCCCGCA  
11201 CAAAAATGAC CAAGAATTC AAGCCGCCAT CTCAAAACT TCATGGAGT GGTGTGTTGC CTTTTCGGC GCGCCTCTGT CGCTATTAAT TATAGGACTT  
11301 ATGATTTTTT CTGACAGAT GATGCTGACT AGCAGACGAA GATGACCGCT ACGCCCAAT GACCCGACCA GCAAACTCG ATGTACTTCC GAGGAACCTA  
11401 TGTGCAATAT GCATCAGGCT GGTATATTAG ATCCCGCTT ACCCGGGGA ATATAGCAAC ACCAAAACTC GACGTATTTT CGAGGAAGCG CAGTGCATAA  
11501 TGCTGCGCAG TGTGCGCAAA TAATCACTAT ATTAACCAAT TATTCAGCGG ACGCCAAAAC TCAATGTATT TCTGAGGAAG CATGGTGCAT AATGCCATGC  
11601 AGCGTCTGCA TAACTTTTA TTATTTT TATTAATCAA CAAAATTTG TTTTAACAT TTC

Fig. 1c

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## S.A.AR86

## A. Amino Acid Sequence of the Nonstructural Polyprotein

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1    MEKPVVNVVDV DQSPFVVQL QKSFQFEVY AQQVTNDHA NARAFSHLAS KLIELEVFTT ATILDIGSAP ARRMFSEHQY HCVCPMRSP EDPDRMMKYAS
101  KLAEKACKIT NENLHEKID LRTVLDTPDA ETPSLCFHND VTCNTRAESY VMQDVYINAP GTTYHQAMKG VRTLYWIGFD TTQFMFSAMA GSYPAYNTNW
201  ADEKYLEARN IGLCSTKLSE GXTGKLSIMR KKEKLPQSRV YFSVGTLYP EHRSLSQSWH LPSVFHLKGG QSYTCRCDTV VSCGYVVKK ITSPGITGE
301  TVGYAVTNS EGFLCKVTD TVKGERVSFF VCTYPATIC DQMTGIMATO ISPDDAQKLL VGLNQRIVIN GKTNRNTNTM QNYLLPIAQ GFSKWAKERK
401  EDLDNEKMLG TREKRLTYGC LWAFRTKKVH SFYRPPGTQT IVKVPASFA FPMSSVWTTT LPMSLRQKMK LALQPKKEEK LLQVPEELVM EAKAAFEFAQ
501  EESRAEKLR EALPLVADKG IEAAAEVCE VEGLDATGA ALYETPRGHV RIIPQANDRM IGQYIVVSM SVLKNAKLAP AHPLADQVKI ITSGRSGRY
601  AVEPYDAKVL MPAGSAVPPW EFLALSESAT LVYNREFVN RCLYHIAMHG PAJNTEEEQY KYTKAELAEY EYVFDVDDKR CVKKEEASGL VLSGELTNP
701  YHELALGLK TRPAVPPYVE TIGVIGTPG GKSAIDKSTV TARDLVTSKG KENCREIAD VLRLGMQIT SKTVDSVMLN GCHKAVEVLY VDEAFCHAG
801  ALLALIAVR PRKVVLCGD PKQCGFFNMH QLVYHFNHE KDICTKTFYK FISRRCTQPV TAVSTLHYD GKMKTTNPCK KNEIDITGA TKPKPDIL
901  TCFRGWVKQL QIDYPGHEVM TAAASQGLTR KGVYAVRQKV NENPLVATS EHVNVLLTRT EDRLVWKTQ GDPWIKQLTN VPKGNFQATI EDWAEHKG
1001 IAAINSAPR TNPFCKTNV CWAKELEPI ATAGIVLTGC QWSELPQPA DDKPHSAYA LDVICKFFG MDLTSGLFSK QSLPTVHPA DSARPVAHWD
1101 NSPGTRKYGY DHAVAAELSR RFPVFQLAGK GTQLDLQTR TRVISAQHNH VPVNRNLPH LVEHKEKQ GPVEKFLSQ KHHSVLVISE KKEAPHKRI
1201 EWIAPIGIAG ADKYNLAFG FPPQARYDLV FINIGTKYRN HHFQCCEDHA ATLKTLRSR LNCNPGGTL VVKSQYADR NSEDDVYALA RKFVRVSAAR
1301 PECVSSNVS YLFRQLDNG RTQGFPHLH NCVSSVYEG TRDGVGAAPS YRTKRENAD CQEEAVVNAA NPLGRPGEGV CRANYKRWPN SFTDSATETG
1401 TAKLTVCCGK KVIHAYGPDF RKHPAEALK LLQNAVHAVA DLVNEHNMKS VAIPLLSTGI YAAGKDRLEV SLNCLTTALD RTDADVTYIC LDKKWKERD
1501 AVLQKESVT ELKDEDMEID DELVWIHPDS CLKGRKGFTS TKGKLYSYFE GTFKHQAAD MAEKVLFVN DQESNEQLCA YLGETMEAI REKCPVDRNP
1601 SSSPFTKPLC LCMYAMTPR VHLRLSNVVK EYTVCSSTPL PKYKIKNVQK VQCTKVVLFN PHTPAFVPAR KYIEAPEQA APPAQAEAP GVVATPTPPA
1701 ADNTSLDVTD ISLDMEDSE GSLFSSFSGS DNYRQVVA DVHAVQEPAP VPPRLKCKMA RLAAARMQEE PTPASTSSA DESLHLSFDO VSISFQSLFD
1801 GEMARLAAQ PASTCTPDV PMSFGSFDG EIEELSRRTV ESEPLFGSF EPGEVNSIS SRSAVSFPPR KQRRRRRSR TEYCLTGVG YFSTDTGFG
1901 HLQKKSVLQN QLTEPTLERN VLERIYAPVL DTSKEEQLK RYQMMPTAN KSRYQSRKE NQKATITERL LSLRLYNSA TDQPECYKIT YPKPSYSSV
2001 PANYSDPKFA VAVCNVYLHE NYPTVASYQI TDEYDAYLDM VDGTVACLDT ATCPAKLRS YPKRHEYRAP MRSVAVPSAM QNTLQNVLIA ATKRNCHVTO
2101 MRELPTLDSA TPNVECFRY ACNDEYWEF ARKPIRITTE FVTAYVARLK GPKAAALFAK THNLVPLQEV PMDRFVMDMK RDVYVTPGTG HTEERPKVQV
2201 IQAAEPLATA YLCGIRRELV RRLTAVLLPN IHTLFDMSAE DFDAAEHF KQGDVPLETD IASFDKSQDD AMALTGLMIL EDLGVDPQLL DLIECAFGEI
2301 SSTHLPTGTR FKFGAMMKSG MFLTLFVNTV LNVVIASRVL EERLTSKCA AFIGDDNIH GYVSDKEMAE RCATWLNMEV KIIDAVIGER PPYFCGGFIL
2401 QDSVTSTACR VADPLKRLFK LGKPLPADDE QDEDRRALL DETKAWFRVG ITDTLAVAVA TRYEDNTTP VLLALRTFAQ SKRAFAQIRG EDKHLVGGPK

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## B. Amino Acid Sequence of the Structural Polyprotein

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1    MNRGFFNMLO RRPFPAPTAM WRPRRRRQAA PMPARNGLAS QIQQLTAVS ALVIGQATRP QTPRPPPPR QKKQAPKQPP KPKPKPTQEK KKKQPAKPKP
101  GKRQRMALKL EADRLFDVKN EDGDVIGHAL AMEGKVMKPL HVKGTIDHPV LSKLKFTKSS AYDMEFAQLP VNMRSFAPTY TSEHPEGFYN WHHGAQVQYG
201  GRFTIPRGVG GRGDSGRFPM DNSGRVYAV LGGADEGTRT ALSVVTWNSK GKTIKTTPEG TEWEAAPLV TAMCLLGNVS FPCNRPTCY TREPSRALDI
301  LEENVNHEAY DTLNAILRC GSSGRSKRSV TDDFTLTPY LGTCSYCHHT EPCFSPKIE QVWDEADDNT IRIQTSAQFG YDQSGAASN KYRYSLEQD
401  HTVKEGTMDD IKISTSGPCR RLSYKGYFL AKCPGDSVT VSIASSNSAT SCTMARKKP KFGVREKYDL PPVHGKKIPCV TVYDRLEKETT AGYITMHRPG
501  PHAYTSYLEE SSGKYVAKPP SGKNTIYECK CGDYKTOTVT TRTEITGCTA DKQVAYKSD QTKWVFNPSD SIRHADHTAQ GKHLHFKLI PSTCMVPPAH
601  APNVVHGFKH ISLQDTHL TLLITRLGA NPEPTTEWII GNTVRNFTVD RDGLEIYWN HEPVRVYAE SAPGDPHGWP HEIVQHYHR HPVYTLAVA
701  SAAYAMMIGV TYAALCACKA RRECLTPYAL APNAVITSL ALLCCVRSAN AETFTETMSY LWSNSQFFFW VQLCIPLAAY VVLMRCCSCC LPFLVAGAY
801  LAKVDAYEHA TTVPNVQIP YKALVERAGY APLNLETVM SSEVLPTNQ EYTCCKFTTV VSPKVRCCG SLECPAAHA DYTCKVFGGV YPFMWGGAQC
901  FCDSENSQMS EAYVELSVDC ATDHAQAKV HTAAMKVGLR IVYGNTTSFL DVYVNGVTPG TSKDLKVIAG PISALFTFD HKVYVIRGLV VNYDFPEYGA
1001 MKPGAFGDIQ ATSLTSKDLI ASTDIRLLK SAKNVHVPYT QASGFEMWK NNSGRPLQET APFGCKIAVN PLRAVDCSYG NIPISIDPN AAFIRTSAP
1101 LVSTVKDVS ECTYSADFGG MATLQYVSDR EGQCPVHSHS STATLQESTV HVLEKGAIVT HPSTASPAQAN FVSLCGKKT TCNAECKPPA DHIVSTPHKN
1201 DQEFQAASK TSWSWLFALF GGASSLLIG LMIFACSMML TSTR

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FIG. 2

## Nucleotide Sequence of Girdwood S.A.

1 NTTGNCGGCG TAGTATACAC TATTGAATCA AACAGCCGAC CAATTGCACT ACCATCAAA TGGAGAAGCC AGTAGTTAAC GTAGACGTAG ACCCGCAGAG  
101 TCCGTTTGTG GTGCAACTGC AAAAGAGETT CCGCAATTT GAGGTAGTAG CACAGCAGGT CACTCCAAAT GACCATGCTA ATGCCAGAGC ATTTTCGCAT  
201 CTGGCCAGTA AACTAATCGA GCTGGAGGTT CTAACACAG CGACGATTTT GGACATAGGC AGCGCACCGG CTCGTAGAAT GTTTCCGAG CACCAATACC  
301 ATTGCGTTTG CCCCATGCGT AGTCCAGAAG ACCCGGACCG CATGATGAAA TATGCCAGCA AACTGGCGGA AAAAGCATGC AAGATTACGA ATAAGAACT  
401 GCATGAGAA ATCAAGGACC TCCGACCGT ACTTGATACA CCGGATGCTG AAAGCCATC ACTCTGCTT CACAACGATG TTACTTGCAA CAGCGTGCC  
501 GAGTACTCCG TCATGCAGGA CGTGATAC TCAGCTCCCG GAACTATTTA CCATCAGGCT ATGAAAGGCG TCGCGACCGT GTACTGGATT GGCTTCGATA  
601 CCACCCAGTT CATGTTCTCG GCTATGGCAG GTTCGTACCC TCGGTACAA ACCAACTGGG CCGACGAAAA AGTCCTCGAA GCGCGTAACA TCGGACTCTG  
701 CAGCACAAG CTGAGTGAAG GCAGGACAGG AAAGTTGTCG ATAATGAGGA AGAAGGAGTT GAAGCCCGGG TCACGGGTTT ATTTCTCCGT TGGATCGACA  
801 CTTTACCCAG AACACAGAGC CAGCTTGCA AGCTGTCATC TTCCATCGGT GTTCCACCTG AAAGGAAAAG AGTCGTACAC TTGCGCTGT GATACAGTG  
901 TGAGCTCGA AGGCTACGTA GTGAAGAAAA TCACCATCAG TCCCGGATC ACGGGAGAAA CCGTGGGATA CCGCGTTACA AACATAGCG AGGGCTTCTT  
1001 GCTATGAAA GTTACCGATA CAGTAAAAGG AGAACGGGTA TCGTTCCCG TGTGCACGTA TATCCCGGCC ACCATATGCG ATCAGATGAC CGGCATAATG  
1101 GCCACGGATA TCTCACTGA CGATGCACAA AAATCTCTGG TTGGGCTCAA CCAGCGAATC GTCATTAAAG GTAAGACTAA CAGGAACACC AATACCATGC  
1201 AAAATTACCT TCTGCCAATC ATTGCACAAG GGTTCAGCAA ATGGGCCAAG GAGCGCAAG AAGACCTTGA CAATGAAAA ATGCTGGGTA CCAGAGAGCG  
1301 CAAGCTTACA TATGGCTGCT TGTGGGCGTT TCGCACTAAG AAAGTCACT CGTTCTATCG CCCACCTGGA ACGCAGACCA TCGTAAAAAT CCCAGCCTCT  
1401 TTTAGCGCTT TCCCATGTC ATCCGTATGG ACTACCTCTT TGCCCATGTC GCTGAGGCGA AAGATAAAAT TGGCATTACA ACCAAGAAAG GAGGAAAAAC  
1501 TGCTGCAAGT CCGGAGGAA TTAGTCATGG AGGCCAAGGC TGCTTTCGAG GATGCTCAGG AGGAATCCAG AGCGGAGAAG CTCGAGAAAG CACTCCACCC  
1601 ATTAGTGGCA GACAAAGGTA TCGAGGCAGC CCGGAAAGTT GTCTGCGAAG TGGAGGGGCT CCAGGCGGAC ATCGGAGCAG CACTCGTCGA AACCCTCGCG  
1701 GGTCATGTAA GGATAATACC ACAAGCAAAAT GACCGATGTA TCGGACAGTA CATCGTTGTC TCGCCAACCT CTGTGCTGAA GAACGCTAAA CTCGACCCAG  
1801 CACACCCGCT AGCAGACCAG GTTAAGATCA TAACGCACTC CGGAAGATCA GGAAGGTATG CAGTGAACC ATACGACGCT AAAGTACTGA TGCCAGCAGG  
1901 AAGTGGCGTA CCATGGCCAG AATTCCTAGC ACTGAGTGAG AGCGCCACGC TAGGTACAAA CGAAAGAGAG TTTGTGAACC GCAAGCTGTA CCATATTGCC  
2001 ATGCACGGTC CCGCTAAGAA TACAGAAGAG GAGCAGTACA AGGTACAAA GGCAGAGCTC GCAGAAACAG AGTACGTGTT TGACGTGGAC AAGAAGCGAT  
2101 GCGTCAAGAA GGAAGAAGCC TCAGGACTTG TCCTCTCGGG AGAACTGACC AACCCTCCCT ATCAGCAACT AGCTCTTGAG GGAAGTGAAG CTCGACCCGT  
2201 GGTCCCGTAC AAGGTTGAAA CAATAGGAGT GATAGGCGCA CCAGGATCGG GCAAGTCGGC TATCATCAAG TCAACTGTCA CGGCACGTGA TCTGTATTAC  
2301 AGCGGAAAGA AAGAAAACG CCGCAAAAT CAGGCGGATG TGCTACGGCT GAGGGGCAAG CAGATCACTG CGAAGACAGT GGATTCGGTT ATGCTCAACG  
2401 GATGCGCGAA AGCCGTAGAA GTGCTGTATG TTGACGAAGC GTTCGGGTGC CACGACGAG CACTACTTGC CTGATTGCA ATCGTCAGAC CCCGTACATA  
2501 GGTAGTGCTA TCGGAGAGCC CTAAGCAATG CGGATTCTC AACATGATGC AACTAAAGGT ATATTCAAC CACCCGAAA AAGACATATG TACCAAGACA  
2601 TTCTACAAGT TTATCTCCCG ACGTTGCACA CAGCCAGTCA CCGGTATTGT ATCGACACTG CATTACGATG GAAAAATGAA AACCACAAAC CCGTGCAAGA  
2701 AGAACATCGA AATCGACATT ACAGGGGCGA CGAAGCGGAA GCCAGGGGAC ATCATCTGA CATGCTTCG CCGGTGGGTT AAGCAACTGC AAATCGACTA  
2801 TCCCGGACAT GAGGTAATGA CAGCCGGGCG CTCACAAGGG CTAACAGAA AAGGAGTATA TGCCGTCCCG CAAAAAGTCA ATGAAAAACC GCTGTACGG  
2901 ATCACATEAG AGCATGTGAA CGTGCTGCTC ACCCGCACTG AGGACAGGCT AGTATGGAAA ACTTTACAGG GCGACCCATG GATTAAGCAG CTCACCTAAG  
3001 TACCAAAAGG AAATTTTCAA GCCACCATCG AGGACTGGGA AGCTGAACAC AAGGGAATAA TTGCTGCGAT AAACAGTCCC GCTCCCGTA CCAATCCGTT  
3101 CAGCTGCAAG ACTAACGTTT GCTGGGGGAA ACGACTGAAA CCGATACTGG CCACGGCCCG TATCGTACTT ACCGTTTGGC AGTGGAGCGA GCTGTTCCTA  
3201 CAGTTTGCAG ATGACAAAAC AACTCGGCC ATCTACGGCC TGGACGTAAT CTGCTAATAG TTTTCGGCA TGGACTTGAC AAGCGGACTG TTTTCCAAAC  
3301 AGAGCATCCC GTTAACGTAC CATCTGCGCG ATTCAGCGAG GCCAGTAGCT CATTGGGACA ACAGCCACAG AACCCTCAAG TATGGGTACG ATCAGCCGCT  
3401 TGCCCGCGAA CTCCTCCGTA GATTTCGGT GTTCCAGTA GCTGGGAAAG GCACACAGCT TGATTGCGAG ACGGGCAGAA CTAGAGTTAT CTCGACACAG  
3501 CATAACTTGG TCCAGTGAA CCGCAATCTC CCGCACGCTT TAGTCCCGA GCACAAGGAG AAACAACCCG GCGCGGTCAA AAAATTCTTG AGCCAGTTCA  
3601 AACACCACTC CGTACTTGTG GTCTCAGAGG AAAAAATGAA AGTCCCCAC AAGAGAATCG AATGGATCGC CCGGATTGGC ATAGCCGGCG CTGATAAGAA  
3701 CTACAACCTG GCTTTCGGGT TTCCGCGCA GGCACGGTAC GACCTGGTGT TTATCAATAT TGGAACTAAA TACAGAAACC ATCACTTCA GCAGTGCGAA

Fig. 3A

3801 GACCATGCGG CGACCTTGAA AACCTCTCG CGTTCGGCC TGAAGTCCCT TAACCCCGGA GGCACCTCG TGGTGAAGTC CTACGGTTAC GCCGACCGCA  
3901 ATAGTGAGGA CGTAGTCACC GCTCTTGCCA GAAAAATTGT CAGAGTGTCT GCAGCGAGGC CAGAGTGCCT CTCAGCAAT ACAGAAATGT ACCTGATCTT  
4001 CCGACAATA GACAACAGCC GCACAGACA ATTCAACCCG CATCATCTGA ATTGTGTGAT TTCTCCCTG TACGAGGGTA CAAGAGACGG AGTTGGAGCC  
4101 GCACCGTCAT ACCGCACTAA AAGGGAGAAC ATTGCTGATT GTCAAGAGGA AGCAGTTGTC AATGCAGCCA ATCCGCTGGG CAGACCAGGC GAAGGAGTCT  
4201 GCCGTGCCAT CTATAAACGT TGGCCGAACA GTTTCACCGA TTCAGGCACA GAGACCGGCA CCGCAAACT GACTGTGTGC CAAGGAAAGA AAGTGATCCA  
4301 CGCGTTGGC CCGATTTCG GGAACACCC AGAGGCAGAA GCCCTGAAAT TGCTGCAAAA CGCTACCAT GCAGTGGCAG ACTTAGTAAA TGAACATAAT  
4401 ATCAAGTCTG TCGCCATCCC ACTGCTATCT ACAGGCATTT AGCGAGCCGG AAAAGACCCG CTGGAAGTAT CACTTAACCTG CTTGACAACC GCGCTAGATA  
4501 GAACTGATGC GGACGTAAAC ATCTACTGCC TGGATAAGAA GTGGAAGGAA AGAATCGACG CGGTGCTCCA ACTTAAGGAG TGTGTAATAG AGCTGAAGGA  
4601 TGAGGATATG GAGATCGAGC ACGATTAGT ATGGATCCAT CCGGACAGTT GCCTGAAGGG AAGAAAGGGA TTCAGTACTA CAAAAGGAAA GTTGTATTGG  
4701 TACTTTGAAG GCACCAAAAT CCATCAAGCA GCAAAAGATA TGGGGAGAT AAAGGTCTGT TCCCAATG ACCAGGAAAG CAACGAGCAA CTGTGTGCTT  
4801 ACATATTGGG GGAGACCATG GAAGCAATCC GCGAAAAATG CCGGTGCGAC CACAACCCGT CGTCTAGCCC GCCAAAAACG CTGCCGTGCC TCTGCATGTA  
4901 TGCCATGACG CCAGAAAGGG TCCACAGACT CAGAAGCAAC AACGTCAAG AAGTTACAGT ATGCTCCTCC ACCCCCTTC CAAAGTACAA AATCAAGAAC  
5001 GTTCAGAAGG TTCAGTGCAC AAAAGTAGTC CTGTTAAAC CGCATACCCC TGCAATCGTT CCGCCCGTA AGTACATAGA AGCGCCAGAA CAGCCTGCAG  
5101 CTCCGCTGC ACAGGCCGAG GAGGCCCGG AAGTTGCAGC AACACCAACA CCACCTGCAG CTGATAACAC CTCGCTTGAT GTCACGGACA TCTCACTGGA  
5201 CATGGAAGAC AGTAGCGAAG GCTCACTCTT TTCGAGCTT AGCGGATCGG ACAACTCTAT TACTAGTATG GACAGTTGGT CGTCAGGACC TAGTCACTA  
5301 GAGATAGTAG ACCGAAGGCA GGTGGTGGT GCTGACGTCC ATGCCGTCCA AGAGCCTGCC CCGTTCCAC CGCCAAGGCT AAAGAAGATG GCCCGCTGG  
5401 CAGCGGCAAG AATGCAGGA GAGCCAACTC CACCGGCAAG CACCAGCTCT GCGGACGAGT CCGTTCACCT TTCTTTTGGT GGGGTATCCA TGTCCTTCGG  
5501 ATCCCTTTTC GACGGAGAGA TGGGGCCCTT GGCAGCGGCA CAACCCCGG CAAGTACATG CCTACGGAT GTGCTATGT CTTCGGATC GTTTCCGAC  
5601 GGAGAGATTG AGGAGCTGAG CCGCAGAGTA ACCGAGTCTG AGCCCGTCT GTTTGGGTCA TTGAACCGG GCGAAGTGAA CTCATTATA TCGTCCGAT  
5701 CAGTTGTATC TTTCCACCA CGCAAGCAGA GACGTAGACG CAGGAGCAGG AGGACCGAAT ACTGACTAAC CCGGGTAGGT GGGTACATAT TTTCCAGGGA  
5801 CACAGCCCT GGGCACTGC AAATGGAGTC CGTCTGCAG AATCAGCTTA CAGAACCAGC CTGGAGCGC AATGTTCTGG AAAGAATCTA CGCCCGGTG  
5901 CTCGACAGCT CGAAAGAGGA ACAGCTCAA CTCAGGTACC AGATGATGCC CACCGAAGCC AACAAAAGCA GGTACCAGTC TAGAAAAGTA GAAAATCAGA  
6001 AAGCCATAAC CACTGAGGGA CTGCTTACG GGCTACGACT GTATAACTCT GCCACAGATC AGCCAGAATG CTATAAGATC ACCTACCGGA AACCATCTA  
6101 TTCCAGCAGT GTACCGGGA ACTACTCTGA CCCAAAGTTT GCTGTAGCTG TTGCAACAA CTATCTGCAT GAGAATTACC CGACGGTAGC ATCTTATCAG  
6201 ATCACCAGC AGTAGCTGC TTACTTGGAT ATGGTAGACG GGACAGTCCG TTGCTAGAT ACTGCAACTT TTTGCCCGC CAAGCTTAGA AGTTACCGGA  
6301 AAAGACAGGA GTATAGAGCC CCAAACTC GCAGTGGGT TCCATCAGCG ATGCAGAACA CGTTGCAAAA CGTGCTCATT GCGCGGACTA AAAGAAACTG  
6401 CAACGTACA CAAATGCGTG AATTGCCAAC ACTGGACTCA GCGACATTCA ACGTTGAATG CTTTCGAAAA TATGCATGTA ATGACGAGTA TTGGGAGGAG  
6501 TTTGCCCGAA AGCCAATTAG GATCACTACT GAGTTCGTTA CCGCATACGT GCGCAGACTG AAAGGCCCTA AGGCCCGCG ACTGTTCCGA AAGACGATA  
6601 ATTTGGTCCC ATTGCAAGAA GTCCCTATGG ATAGGTTCGT CATGGACATG AAAAGAGACG TGAAGTTAC ACCTGGCAGC AAACACACAG AAGAAAGACC  
6701 GAAAGTACAA GTGCTACAAG CCGCAGAACC CTTGGCGACC GCTTACCTGT CCGGGATCCA CCGGAGTTA GTGCGCAGGC TTACAGCCGT CTGCTACCC  
6801 AACATTACA CGCTTTTGA CATGTCGGG GAGGACTTTG ATGCAATCAT AGCAGAACAC TTCAAGCAAG GTGACCCGT ACTGGAGACG GATATCGCT  
6901 CGTTGACAA AAGCCAAGAC GACGCTATGG CGTTAACTGG CCGTATGATC TTGGAAGACC TGGGTGTGGA CCAACCACTA CTCGACTGA TCGAGTGGC  
7001 CTTTGGAGAA ATATCATCCA CCCATCTGCC CACGGGTACC CGTTTCAAT TCGGGCGAT GATGAAATCC GGAATGTTCC TCACGCTCTT TGTCAACACA  
7101 GTTCTGAATG TCGTTATGCC CAGCAGAGTA TTGGAGGAGC GCGTTAAAA GTCCAAATGT GCAGCATTTA TCGGCGACGA CAACATCATA CACGGAGTAG  
7201 TATCTGACAA AGAAATGGCT GAGAGTGTG CCACCTGGCT CAACATGGAG GTTAAGATCA TTGACGCACT CATEGGCGAG AGACCGCTT ACTTCTGCGG  
7301 TGGATTATC TTGCAAGATT CGGTACCTC CACAGCGGT CGCGTGCGG ACCCTTGAA AAGCTGTTT AAGTTGGTA AACCGTCCC AGCCGACGAC  
7401 GAGCAAGACG AAGACAGAAG ACGCGCTCTG CTAGATGAAA CAAAGGCGTG GTTTAGAGTA GGTATAACAG ACACCTTAGC AGTGGCCGTG GCAACTCGGT  
7501 ATGAGGTAGA CAACATCACA CCGTCTCTG TGGCATTGAG AACTTTTGC CAGAGCAAAA GAGCATTTCA AGCCATCAGA GGGGAAATAA AGCATCTCTA  
7601 CGGTGCTCT AAATAGTCAG CATAGCACAT TTCATCTGAC TAATACCACA ACACCACCAC CATGAATAGA GGATTCTTTA ACATGCTCGG CCGCCGCCCC  
7701 TTCCCGCCCC CCACTGCCAT GTGGAGGCCG CGGAGAAGGA GCGAGCGCG CCGATGCTT GCGCGAATG GGTGGCTTC CCAATCCAG CAACTGACCA  
7801 CAGCCGTCAG TGCCCTAGTC ATTGGACAG CAACTAGACC TCAACCCCA CCGCCACGCC CGCCCGCGG CCAAGAAGA CAGGCGCAA AGCAACCACC

Fig. 3B

7901 GAAGCCGAAG AAACCAAAAA CACAGGAGAA GAAGAAGAAG CAACCTGCAA AACCCAAACE CGGAAAGAGA CAACGTATGG CACTCAAGTT GGAGGCCGAC  
8001 AGACTGTTTC ACGTCAAAAA TGAGGACGGA GATGTTCATCG GGCACGCACT GGCCATGGAA GGAAGGTAA TGAAACCACT CCACGTGAAA GGAAGTATTC  
8101 ACCACCTGT GCTATCAAA GCTAAATCA CCAAGTCCTC AGCATAAGAC ATGGAGTTCC CACAGTTGCC GGTCAACATG AGAAGTGAGG CGTTCACTTA  
8201 CACCAGCGAA CACCTGAAG GGTTTTACAA CTGGCACCAC GGAGCGGTGC AGTATAGTGG AGGTAGATT ACCATCCCC CGGGAOTAGG AGGCAGAGGA  
8301 GACAGTGTTC GTCCGATTAT GGATACTCA GGCCGGGTTG TCGGATAGT CTEGGAGGG GCTGATGAGG GAACAAGAAC TGCCCTTTTC GTCTCACTT  
8401 GGAATAGCAA AGGGAAGACA ATCAAGACAA CCCCGGAAGG GACAGAAGAG TGGTCTGCA CCACTAGTGT CACGGCCATG TGTCTGCTT GAAACGTGAG  
8501 CTTCCTATGC AATGCCCGCC CCACATGCTA CACCCGGBAA CCATCCAGAG CTCTTGACAT CCTTGAAGAG AACGTGAACC ACGAGGCTTA CGACACCTTC  
8601 CTCAACGCCA TATTGCGGTG CGGATCGTCC GGCAGAAGCA AAAGAAGCGT CACTGACGAC TTTACCTTGA CCAGCCCGTA CTGGGCGACA TGCTCGTACT  
8701 GTCAACATAC TGAACCGTGC TTTAGCCCCA TTAAGATCGA GCAGGTCTGG GATGAAGCGG ACGACAACAC CATACGCATA CAGACTTCGG CCCAGTTTGG  
8801 ATACGACCAA AGCGGAGCAG CAAGCTCAAA TAAGTACCGC TACATGTCGC TCGAGCAGGA TCATACCGTC AAAGAAGGCA CTATGGATGA CATCAAGATC  
8901 AGCACCTCAG GACCGTGTAG AAGGCTTAGC TACAAAGGAT ACTTTCTCTC CGCGAAGTGT CCTCCAGGGG ACACCGTAAC GGTAGTATA GCGAGTAGCA  
9001 ACTCAGCAAC GTCATGCACA ATGGCCCGCA AGATAAAACE AAAATTCGTG GGACGGGAAA AATATGACCT ACCTCCCGTT CACGGTAAGA AGATTCTTTG  
9101 CACAGTGTAC GACCGTGTGA AAGAAACAA CGCCGGCTAC ATCACTATGC ACAGGCCGGG ACCGCACGCC TATACGTCT ATCTGGAGGA ATCATCAGGG  
9201 AAAGTCTACG CGAAGCCACC ATCCGGAAG AACATTACGT ACGAGTGCAA GTCCGGCGAT TACAAGACCG GTACCGTTAC GACCCGTACC GAAATCACGG  
9301 GTTGCAACCG CATCAAGCAG TGCCTCGCT ATAAGAGCGA CCAACGAAG TGGGTCTTCA ATTCCCGGA CTGATCAGA CATGCCGACC ACACGGCCCA  
9401 AGGGAATTC CATTACCTT TCAAGTGTAT CCCGAGTACC TGCATGGTCC CTGTGCCCCA CGCCCGCAAC GTAGTACACG GCTTTAAACA CATCAGCCTC  
9501 CAATTAGACA CAGACCCT GACATTGCTC ACCACCAGGA GACTAGGGGC AAATCCGGA CCACTACTG AATGATCAT CGGAAAGACG GTTAGAAACT  
9601 TCACCGTGA CCGAGATGGC CTGGAATACA TATGGGGCAA TCACGAACCG GTAAGGGTCT ATGCCAAGA GTCTGCACCA GGAGACCTTC ACGGATGGCC  
9701 ACACGAAATA GTACAGCATT ACTACCATCG CCATCCTGTG TACACCATCT TAGCCGTGCG ATCAGTGTCT GTGGCGATGA TGATTGGCGT AACTGTGCA  
9801 GCATTATGT CCGTAAAGC GCGCCGTGAG TGCCTGACGC CATATGCCCT GGGCCCAAT GCGTGATTC CACTTCGCT GGCACTTTG TGTGTGTTA  
9901 GGTGGCTAA TGCTGAACA TTCACCGAGA CCATGAGTTA CCTATGTCG AACAGCCAGC CATTCTCTG GGTCCAGCTG TGTATACCC TGGCCGCTGT  
10001 CATCGTTCTA ATGCGCTGTT GCTCATGCTG CCTGCTTTT TTAGTGTTG CGCGCCCTA CCGGCGAAG GTAGACGCTC ACGAACATGC GACCACTGTT  
10101 CCAATGTGC CACAGATACC GTATAAGGA CTTGTTGAAA GGGCAGGTA CGCCCGCTC AATTGGAGA TTAGTGTAT GTCTCGGAG GTTTTGCTT  
10201 CCACCAACCA AGAGTACATC ACCTGCAAT TCACCACTGT GGTCCCCCTC CTAAGTCA AATGCTCGG CTCTTGGA TGTACGCCG CGGCTCACGC  
10301 AGACTATACC TGCAAGTCT TCGAGGGGT GTACCCCTTC ATGTGGGGAG GAGCACAATG TTTTTCGAC AGTGAGAACA GCCAGATGAG TGAGGCGTAC  
10401 GTCGAATTGT CAGCAGATTG CGCGACTGAC CACCGCAGG CGATTAAGGT GCATCTGCC GCGATGAAAG TAGGACTACG TATAGTGTAC GGGAAACCTA  
10501 CCAGTTCTCT AGATGTGATC GTGAACGGAG TCACACCAGG AACGTCTAAA GACGTGAAAG TCATAGCTGG ACCAATTCA GCATCGTTA CACCATGGA  
10601 TCACAAGTCT GTTATCCATC GCGGCTGTGT GTACAACTAT GACTTCCCGG AATACGGAGC GATGAAACCA GGAGCGTTG GAGACATTA AGCTACCTCC  
10701 TTGACTAGCA AAGATCTCAT CGCCAGCACA GACATTAGAC TACTCAAGCC TTCCGCCAAG AACGTGCATG TCCCTACAC GCAGGCCGCA TCTGATTGG  
10801 AGATGTGGA AAACAACCTA GCGCGCCAC TGCAGGAAAC CGCCCTTTC GGTGCAAGA TTGCACTCA TCCGCTTCCA GCGGTGGACT GCTCATACGG  
10901 GAACATTCCT ATCTCTATCG ACATCCGAA CGTGCCTTT ATCAGGACAT CAGATGCACC ACTGGTCTCA ACAGTCAAT GTGATGTCAG TGAGTGCACT  
11001 TACTCAGCG ACTTCGGCG GATGGCTACC CTGCAATG TATCCGACCG CGAAGGACAA TGCCCTGTAC ATTCGCTTC GAGCACAGCA ACCCTCCAAG  
11101 AGTCGACAGT TCATGTCTG GAGAAAGGAG CGGTGACAGT ACATTCAGC ACCCGAGCC CACAGCGAA CTTTATTGTA TCGCTGTGT GTAAGAAGAC  
11201 AACATGCAAT GCAGAATGCA AACCAACAGC TGACCATATC GTGAGCACCC CGCAGAAAAA TGACCAAGAA TTCCAAGCG CCATCTCAAA AACTTCATGG  
11301 AGTTGGCTGT TTGCCCTTT CGCGGGCGCC TCGTCGTAT TAATTATAGG ACTTATGATT TTTGCTTGA GCATGATGCT GACTAGCACA CGAAGATGAC  
11401 CGCTACGCC CAATGACCG ACCAGCAAAA CTCGATGTAC TTCCGAGGAA CTGATGTGCA TAATGCATCA GGCTGGTATA TTAGATCCCC GCTTACCGCG  
11501 GGCAATATAG CAACACAAA ACTCGACGTA TTTCGAGGA AGCGCAGTGC ATAATGCTGC CAGTGTGTC CAAATAATCA CTATATTAA CATTATTTA  
11601 GCGGACGCCA AAACCTAATG TATTTCTGAG GAAGCATGGT GCATAATGCC ATGACGCTC TGCATAACT TTTATTATT CTTTATTAA TCAACAAAT  
11701 TTTGTTTTTA ACATTTN

Fig. 3c

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## Girdwood S.A.

## A. Amino Acid Sequence of the NonStructural Polyprotein

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1      MEKPVVNDV DPQSPFVVL QKSPQFEVY AQQVTPNDHA NARAFSHLAS KLIELEVPTT ATILDIGSAP ARRMFSEHQY HCVCPMRSPE DPDRMMKYAS
101     KLAEKACKIT NKNLREKIKD LRTVLDTFDA ETPSLCFHND VTCNTRAEYS VMQDVYINAP GTTYHQAMKG VRTLYWIGFD TTFMFMSAMA GSYPAYNTNW
201     ADEKYLEARN IGLCTKLESE GRTGKLSMR KKEKPGSRV YFSVGLTLYP EHRSLSQSWH LPSVPHLKCK QSYTCRCDTV VSCGGYVVKK ITSPGITGE
301     TVGYAVTNS EGFLLCKYTD TVKGERVSFP VCTYPATIC DQMTGDMATD ISPDQAQKLL VGLNQRIVIN GKTNRNTNTM QNYLLPIAQ GFSKWAKERK
401     EDLDNEKMLG TREERKLYGC LWAFRTKKVH SFYRPPGTQT IVKVPASFA FPMSSVWTTT LPMSLRQKIK LALQPKKEEK LLQVPEELVM EAKAAFEDAQ
501     EESRAEKLRE ALPLYADKG IEAAAEVYCE VEGLOADIGA ALVETPRGHV RIIPQANDRM IGQYTVVSPT SYLKNAKLAP AHPLADQVKI ITHSGRSGRY
601     AVEPYDAKVL MPAGSAVPWP EFLALSESAT LVYNEREFVN RKLYHIAMHG PAKNTEEEQY KVTKAELAET EYVFDVKKR CVKKEEASGL VLSGELTNP
701     YHELALEGLK TRPVVPYKVE TIGVIGAGS GKSADKSTV TARDLYTSGK KENCREIQAD VLRLRGMQT SKTVDSVMLN GCRKAVEVLY VDEAFACHAG
801     ALLALAIVR PRHKVVLGGD PKQCGFFNMN QLKVYFNHPE KDICTKTFYK FISRCTQPV TAVSTLHYD GKMKTTNPKC KNIEDITGA TKPKPGDIL
901     TCFRGWVKQL QIDYFGHEVM TAAASQGLTR KGVYAVRQKV NENPLYAITS EHVNVLLTRT EDRLVWXTLQ GGPWKQLTN VPKGNFQATI EDWAEHKG
1001    IAADNPAPR TNPPSCKTNY CWAKRLEPIL ATAGVLTGC QWSELFPFA DDKPHSAIYA LDVICKFFO MDLTSGLFSK QSIPLTTHPA DSARPVAHW
1101    NSPOTRYGY DHAVAAELSR RFPVFLQAGK GTQLDLQGR TRVISAGHNL VPVNRNLPHA LVPEHKEKQP GPVKKFLSQF KHHSVLVSE EKIEAPHKRI
1201    EWIAPIGAG ADKNYNLAFG FPPQARYDLV FINIGTKYRN HHFQCCEDHA ATLKTLRSR LNCNPGGTL VVKSYGYADR NSEDVVTALA RKFVRVSAAR
1301    PECVSSNTEM YLIFRQLONS RTRQFTPHIL NCVSSVYEG TRDGVGAAPS YRTKRENIAD CQEEAVVNAA MPLGRPGEGV CRAIYKRWPN SFTDSATETG
1401    TAKLTVCGQK KVIHAVGPDF RKHPAEALK LLQNAVHAVA DLVNEHNKS VAIFLLSTGI YAAAGKDRLEV SLNCLTTALD RTDADVTYTC LDKKWKERID
1501    AVLQLKESVI ELKDEDMEID DELVWTHPDS CLKGRKGFST TKGKLYSYFE GTFKHQAARD MAEKVLPFN DOESNEQLCA YILGETMEAI REKCPVDHNP
1601    SSSPKTLPC LCMYAMTPER VHLRSNNHYK EYTVCSSTPL PKYIKIKVQK VQCTKVVLFN PHTPAFVPAR KYEAPQPCA APPAQAEAP EVAATPTPPA
1701    ADNTSLDVTI ISLDMEDSSE GSLFSSFGS DNSITSMDSW SSGPSSLEIV DRRQVVVADV HAVQEPAPVP PPRLLKMARL AAARMQEPTT PPASTSSADE
1801    SLHLSFGGVS MSFSGSLFDGE MGALAAAQPP ASTCPTDVPM SFGSFDGEI EELSRVYTES EPVLFSGFEP GEVNSISSR SVVSFPPRKQ RRRRASRRT
1901    Y

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## B. Amino Acid Sequence of the Structural Polyprotein

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1      MNRGFFNMLG RRPFFAPTAM WRPRRRRQAA PMPARNGLAS QIQQLTAVS ALVIGQATRP QTPRPPTPPR QKKQAPKQPF KPKKPKTQEK KKKQPAKPKP
101     GKRQRMALKL EADRLFDVKN EDGDVIGHAL AMEGKVMKPL HYKGTIDHPY LSKLKFTKSS AYDMEFAQLP VNMRSFAFTY TSEHPEGFYN WHHGAVQYSG
201     GRPTIPRGVG GRGDSGRPM DNGSRVVAIV LGGADEGTRT ALSVVTWNSK GKTIXTTPEG TEEWSAAPLY TAMCLLGNVS FPCNRPTCTY TREPSRALDI
301     LEENVNHEAY DTLNAILRC GSSGRSKRSV TDDFTLTSY LGTCSYCHHT EPCFSPKIE QWQDEADDNT IRIQTSAQFG YDQSGAASN KYRYMSLEQD
401     HTVKEGTMDD IKISTSGPCR RLSYKGYELL AKCPGDSVT VSIASSNSAT SCTMARKIKP KFGGREKYDL PPVHGKKIPC TVYDRLKETT AGYITMHRPG
501     PHAYTSYLEE SSGKVYAKPP SOKNTTYECK CGDYKTOTVT TRTETGCTA IKQCVAYKSD QTKWVFNSPD LIRHADHTAQ GKLHLFPKLI PSTCMVPVPAH
601     APNVVHGFKH ISLQLDTHL TLLTTRRLGA NPEPTTEWII GKTVRNFTVD RDGLEIYWNH HEPVRVYAE SAPGDPHGWP HEIVQHYTHR HPVYTLAVA
701     SAAYAMMIGV TYAALCACKA RRECLTPYAL APNAVITSL ALLCCVRSAN AETPTETMSY LWSNSQFFFW VQLCIPLAIV ILMKRCSCC LPFLVAGAY
801     LAKVDAYEHA TTVPNVPQIP YKALVERAGY APLNLEITVM SSEVLPSTNQ EYITCKFTTY VSPKVKCCG SLEQPAAHA DYTCKVFGGV YPFMWGQAQC
901     FCDSENSQMS EAYVELSADC ATDHAQAIKV HTAAMKVGLR IVYGNTTSL DYYVNGVTPG TSKDLKVIAG PISASFTPD HKVVIHRLV YNYDPFEGYA
1001    MKPGAFGDIQ ATSLTSDLI ASTDIRLLKP SAKNVHVPYT QAASGFEMWK NNSGRPLQET APFGCKIAYN PLRAVDCSYG NIPISIDIPN AAFIRTSAP
1101    LVSTYKCDVS ECTYSADFGG MATLOVYSDR EGQCPVHSHS STATLQESTV HYLEKGAVTV HFSTASQAN FIVSLCGKKT TCNAECKPPA DHVSTPHKN
1201    DQEFQAISK TSWSWLFAFP GGASSLLIG LMIFACSMML TSTR

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Fig. 4

## Nucleotide Sequence of S55

1 ATTGGCGCGG TAGTACACAC TATTGAATCA AACAGCCGAC CAATTGCACT ACCATCAACA TGGAGAAGCC AGTAGTTAAC GTAGACGTAG ACCCTCAGAG TCCGTTTCTC GTGCAACTGC  
 121 AAAAGAGCTT CCGCAATTT GAGGTAGTAG CACAGCAGGT CACTCCAAAT GACCATCTTA ATGCCAGAGE ATTTTGCAT CTGGCCAGTA AACTGATCGA CCGTGGAGTT CCAACACAG  
 241 CGACGATTTT GGACATAGGC AGCGACACCG CTCGTAGAAAT GTTTTCCGAG CACCAGTACC ATTCGCTTTG CCCCATGCGT AGTCCAGAAAG ACCCGGACCG CATGATGAAA TATGCCAGCA  
 361 AACTGGCCGA AAAAGCATGT AAGATTACAA ACAAGAACTT GCATGAGAAG ATCAAGGACC TCCGGACCGT ACTTGATACA CCGGATGCTG AAACGCCATE ACTGTCTTTC CACAACGATG  
 481 TTACTGCGAA CACCGTCCG GAGTACTCCG TCATCCAGGA CGTGATACAT AACGCTCCCG GAACATTTTA CCACGAGCCT ATGAAGGCGT TCCGAGCCCT GTACTGGATT GCGTTCGACA  
 601 CCACCGAGTT CATGTTCTCG GCTATGCCAG GTTCGTACCC TCATACAAAC ACCAAGTCCG CCGACGAAAA AGTCCTTGAA CCGGCTAACA TCGGACTCTG CAGCACAAGG CTGAGTGAAG  
 721 GCAGGACAGT AAGGTGTTGG ATATGAGGGA AGAAGGAGTT GAAGCCCGGG TACCGGTTT ATTCTCCGCT TGGATGACAA CTTTACCCAG AACACAGAGC CAGCTTGCAAG AGCTGGCATC  
 841 TTCCATCGGT GTTCCACTTG AAAGGAAAGC AGTGTACAC TTGCGCGTGT GATACAGTGG TGAGCTGCGA AGGCTACGTA GTGAAGAAAA TCACCATCAG TCCCGGGATE ACCGGAGAAA  
 961 CCGTGGGATA CCGGTTTACA AACATAGCG AGCGCTTCTT GCTATGCAAA GTTACCGATA CAGTAAAGAG AGAAGCGGTA TCGTTCCTCG TGTGCACTGA TATCCCGGCC ACCATATCGG  
 1081 ATCAGATGAC CCGCATAATG GGCACGGATA TCTACCTGTA CGATGCAACA AACTTCTGCT TTGCGCTTAA CCAGCGAATC GTCAATTAACG GTAAGACTAA CAGGAACACC AATACCATCC  
 1201 AAAATTACCT TCTGCCAATC ATTCACAAAG GGTTCAGCAA ATGGGCCAAG GAGCGCAAGG AAGATCTTGA CAATGAAAAA ATGCTGGCCA CCAGAGAGCG CAAGCTTACA TATGGCTGCT  
 1321 TGTGGCGGTT TCGCACTAAG AAAGTGCACT CTTTCTATCG CCCACCTGGA ACCGAGACCA TGTAAAGAT CCGAGCCTCT TTTAGCGCTT TCCCATGTC ATCCGTATCG ACTACCTCTT  
 1441 TTGCCATGTC CCGTAGCCAG AAGATGAAT TGGCATTACA ACCAAGAAAG GAGGAAAAAC TCGTCAAGT CCGGAGGAA TTAGTTATCG AGCGAAGGC TCGTTTCGAG GATGTCAGG  
 1561 AGGAATCCAG AGCGGAGAAAG TCCGAGAAAG CACTCCACCC ATTACTGGCA GACAAAGGTA TCGAGGCAAC TCGGGAAGTT GTTTCGCAAG TGGAGGAGTT CCGAGCGGAC ACCGGAGCAAG  
 1681 CACTGCTGCA AACCCCGCGC GGTGATGTA GGATAATACC TCAAGCAAT GACCGTATGA TCGGACAGTA TATGTTGTC TCGCGGATCT CTGTGCTGAA GAACGCTAAA CTGCGACCAAG  
 1801 CACACCCGCT AGCAGACGAG GTTAAGATCA TAACGCACTC CGGAAGATCA GGAAGGTATG CAGTGAAGC ATACGAGCCT AAAGTACTGA TCGCAGCAGG AAGTCCGCTA CCAATGCCAG  
 1921 AATTCTTAGC ACTGAGTGAG AGCGCCACCG TTGTGTACAA CGAAAGAGAG TTTGTGAACC GCAGCCTGTA CCATATTGCC ATGCAAGGTC CCGCTAAGAA TACAGAGAGG GAGCAGTACA  
 2041 AGGTACAAA GGCAGAGCTC GCAGAAACAG AGTACGTGTT TGAGTGGAC AAGAAGCGAT CGTTTAAGAA GGAAGAAGCC TGAAGACTTG TCTTTTCGGG AGAAGTGAAC AACCCGCTCT  
 2161 ATCAGCAACT AGCTTTTGA GCACTGAAGA CTGACCCCGC GGTCCGCTAC AAGGTTGAAA CAATAGGAT GATAGGCA CAAGATCCG GCAAGTCAAG TATCATCAAG TCAACTGTCA  
 2281 CGGCACTGTA TTTGTTTACC AGCGGAAAGA AAGAAAACTG CCGCGAAATT GAGCGGACCG TCGTACCGCT GAGCGGCAAT GAGTACAGCT CGAAGCAAGT GATTCGGTT ATGCTAACG  
 2401 GATGCCACAA AGCCGTAGAA GTGCTGTATG TTGACGAAGC GTTCCGCTGC CAGCGAGGAG CACTACTTGC CTTGATTGCA ATGCTGAGAC CCGCTAAGAA GGTAGTACTA TCGGAGACCC  
 2521 CTAAGCAATG CCGATTCTTC AACATGATGC AACTAAAGT ACATTCAAC CACCTGAAA AAGACATATG TACCAAGACA TTCTACAAGT TTATCTCCCG ACCGTGACA CAGCAGTCA  
 2641 CCGCTATTGT ATGACACTG CATTACGATG GAAAAATGAA AACCAACAA CCGTCAAGA AGAATCATGA AATGACATT ACAGGGGCA CGAAGCCGAA GCGAGCGGAC ATCATCTGTA  
 2761 CATGTTTCCG CCGGTGGGTT AAGCACTGC AAATGACTA TCCCGGACAT GAGGTATGA CAGCCGCGCG CTCACAAGGG CTAACAGAA AAGGAGTATA TCCGTCGGG CAAAAAGTCA  
 2881 ATGAAAAACC CGTGTACCGC ATCAGATCAG AGCATGTGAA CGTGTGCTC ACCGCACTG AGGACAGGCT AGTATGAAA ACTTTACAGG GCGACCCATG GATTAAGCAG CTCACTAAGC  
 3001 TACCTAAAGG AAATTTTCA GGCACATCG AGGACTGGGA AGCTGAACAC AAGGAATATA TTGCTCGAT AAACAGTCCC GCTCCCGTA CCAATCGTT CAGCTCAAG ACTAACGTTT  
 3121 GCTGGCGGAA AGCACTGGAA CCGTACTGCG CCACGCGCGG TATCGTACT ACCTGCTGCG AGCTGCGGCA ATGTGAGGCA GCTTTTCCA CAGTTTCCCG ATGACAAAGC ACATCGGCC ATCTACGCT  
 3241 TAGACGTAAT TTGCTAAG TTTTTCGCA TGGACTTAC AAGCGCGCT TTTTCAAC AGAGCATCC GTTAACGTAC CATCTGCGC ACTCAGCGAG GCGAGTAGT CATTGGGACA  
 3361 ACAGCCCAAG AACACGCAAG TATGGTAGC ATCAGCGCT TCCCGCGGAA CTCCTCGTA GATTTCGCT GTTCCAGTA CCGTGGAAAG GCACACAGCT TGATTTCAG ACCGGCAGAA  
 3481 CTAGAGTTAT CTCTGCACAG CATACTTGG TCCAGTGAA CCGCAATCTC CCTCAGCCT TAGTCCCGCA GCACAAGGAG AAACAACCCG CCGCGGTGCA AAAATTCTTG AGCGAGTTCA  
 3601 AACACCACTC CGTACTTGT ATCTCAGAGA AAAAAATGA AGCTCCCAAC AAGAGATGAG AATGATGCG CCGGATTGCG ATAGCCGCGC CAGATAAGAA CTACAACCTG GCTTTCCGCT  
 3721 TTCCCGCGCA GGCACGCTAC GACTGTGTT TCATCAATAT TGGAACTAAA TACGAAACAC ATCACTTTCA AGAGTGGAA GAGCAGCGCG CGACTTTGAA AACCTTTTGG CTTGCGCGC  
 3841 TGAAGTCCCT TAAACCCGGA GGCACCTCTG TGTGAAGTTC TACGGTTTAC CCGACCGCA ATAGTGAAGA CGTAGTCAAC GCTTTTCCA CAGTTTCCCG ATGACAAAGC ACATCGGCC ATCTACGCT  
 3961 CAGAGTGGCT CTCAGCAAT ACAGAAATGT ACCTGATTT CCGACAATA GACAACAGCC GCACACGACA ATTCACCCCG CATEATTGA ATTGTGAT TTGCTCGCTG TACGAGGTA  
 4081 CAAGAGACCG AGTTGAGGCC GCACCTGCT ACCGTACTAA AAGGAGAAC ATTCGTAAT GTCAAGAGA AGCAGTTGTC AATGACGCA ATCACTGGG CAGACAGGA GAAGGAGTCT  
 4201 CCGTGGCAT CTATAACGCT TCGCCGACA GTTTCACCGA TTACGCGACA GAGACAGGTA CCGCAAACT GACTGTGTC CAAGGAAAGA AAGTATCCA CCGGTGCGC CCGTATTCC  
 4321 GGAACACCC AGAGCGAGAA GCGCTGAAT TGTGCAAAA CCGCTACCAT CGAGTGGAG ACTTAGTAAA TGAACATAAT ATCAAGTCTG TCGCATCC CACTGTATCT ACAGGCATT  
 4441 AGCGAGCCCG AAAAGACCGC CTTGAGGAT CACTTAACTG CTTGACAACC GCGTAGACA GAAGTATG GAGGTAAAC ATCTACTGCC TGGATAAGAA GTGGAAGGAA AGAATGAGC  
 4561 CGCTGCTCCA ACTTAAGGAG TGTGTAAGT AGCTGAAGGA TGAGGTATG GAGTGGAGC ACGAGTTAGT ATGATCCAT CCGGACAGTT GCGTGAAGG AAGAAAGGGA TTCACTACTA  
 4681 CAAAAGGAAA GTTGTATTG TACTTTGAG GCACCAATT CCATCAAGCA GCAAAAGATA TCGCGGAGT AAGGTGCTG TTCCCAATG ACCAGGAAAG CAACGAACAA CTGTGTGCT  
 4801 ACATATTGGG GGAGACCATG GAAGCAATCC GCGAAAAAT CCGGTGCGAC CACAACCCGT GGTGAGCCG GCGAAAAAG CCGCGTCC TCTGTATGTA TCCCATGAGC CCGAGAAAGG  
 4921 TCCACAGACT CAGAAAGCAAT AAGTCAAGG AAGTTACAGT ATCTCTCTC ACCCCCTTC CAAAGTACAA AATCAAGAA GTTACAGAGG TTCACTGAC AAAAGTAGTC CTGTTTAAAC  
 5041 CCGTACCCCG CCGATTGCT CCGCCCGCTA AGTACATAGA AGCACCAGAA CAGCTGCGAG CTCCGCTCTC ACAGCGCGAG GAGCGCCCG GAGTTGATG GAGCAGCAAC CCACTGCGAG  
 5161 CTGATAACAC CTCGCTTAT GTACCGGACA TCTCACTGGA CATGGAAGAC AGTAGCGAAG GCTCACTTT TTGAGCTTT AGCGGATCG ACAACTAGC AAGCGAGTG GTGTGCGCTG  
 5281 AGCTCATGCG CCGTCAAGAG CCGTCCCTG TTCCACCGCC AAGGTAAAG AAGATGCCCC GCTGCGAGC GCGAAGAAAT CAGGAAGAGC CAACTGCAAC GCGAAGCACC AGCTCTCGCG  
 5401 ACGAGTCCCT TCACCTTTCT TTTGATGGG TATCTATATC CTTGGAATC CTTTGGAGC GAGAGATGCG CCGTTTCCA CCGGCAAC CCGCGGCAAG TACATGCCCT ACGGATGTC  
 5521 CTATGTTTTT CCGATGCTT TCGACGGAG AGATTGAGGA GTTGAGCGC AGAGTAAACC AGTGGAGCC CCGCTGTTT GGTCAATTG AACCGCGCA AGTGAATCA ATTATATCT  
 5641 CCGGATGAGC CGTATCTTT CCAACACCGA AGCAGAGAGC TAGACCGAGC AGCAGGAGGA CCGAATAGT TCTAACCGGG GTAGGTGGT ACATATTTC CAGCGACACA GCGCTCGCG  
 5761 ACTTGCAAAA GAAGTGGTT CTGCAAGC AGCTTACAGA ACCGACCTTG GAGCGCAATG TTCTGGAAG AATCAACCC CCGGTGCTG ACAGCTGGA AGAGGAACAG CTCAAAATCA  
 5881 GGTACCATG GATGCCACC GAAGCAACA AAGCAGGTA CAGTGTGGA AAGTAGAAA ACCGAAAGC CATAACCACT GAGCGACTCG TTGAGCGCT ACGCTGTAT AACTGTGCA  
 6001 CAGATCAGCC AGAATCTAT AAGTACCTT ACCCGAAAC ATGATATTC AGCACTGAC CCGCAACTA CTGAGCCCA AAGTTTCTG TAGCTTTTG TAACAACTAT CTGATGGA  
 6121 ATTACCCGAC GGTAGCATCT TATCAAGTA CCGAGAGTA CGATGCTTAC TTGATATG TAGACGGAG AGTCCCTTC CTAGATACT CAACTTTTG CCGCGCAAG CTTAGAAGT  
 6241 ACCCGAAAG ACACGAGTAT AGAGCCCAA ACATCCGAG TCGGTTTCA TAGCGATGC AGAACAGCT GCAAAACGTC CTCATTGCG CCACTAAAG AAGTGCAC GTACACAAA  
 6361 TCGGTAACT GCAACACTG CACTACGGA CATTCAAGT TGAATGCTT CCAAAATAT CATGCAATGA CGAGTATTG GAGGAGTTT CCGAAAGCC AATTAGGATC ACTACTGAT  
 6481 TCGTTACCGC ATACGTGCC AGACTGAAAG GCGCTAAGC CCGCGCACT TTGCAAGA CCGTAATTT GGTCCCATG CAAGAAGTC CTATGGATG ATTCGTATG CACATGAAA  
 6601 GAGAGCTGAA AGTTACACT GGCACGAAC ACACAGAGA AAGACCGAAA GTACAAGTA TACAAGCCG AGAACCCCTG GCGACCGCTT ACCTATCGG GATCACCAG GAGTTAGTGC

Fig 5A

6721 GCAGGCTTAC AGCCGTTTT CTACCCAACA TTCACAGCT CTTCGACATG TCGCGGAGG ACTTTCATGC AATCATAGCA GAACACTTCA AGCAAGGTGA CCCGGTACTG GAGACGGATA  
6841 TCGCTCGTT CGACAAAAGC CAAGACGAGC CTATCGCGTT AACCGCGCTG ATGATCTTGC AAGACCTGGG TGTGGACCAA CCACTACTCG ACTTGATCGA GTCCGCTTT GGAGAAATAT  
6961 CATCCACCCA TCTCCCAAGC GGTACCGGTT TCAAAATCGG GCGATGATG AAATCGGAA TGTCTCTAC GCTCTTTGTC AACACAGTTC TGAATGTGCT TATCGCCAGC AGAGTATTGG  
7081 AGGAGCGGCT TAAAACGTCC AAATGTGACG CATTATCGG CGACGACAA ATATATACAGC GAGTAGTATE TGACAAAGAA ATGCGTGAGA GGTGTGCCAC CTGCTCAAC ATGGAGGTGA  
7201 AGATCATTTA CGCAGTCATC GCGAGAGAC CACCTTACTT CTGCGGTGGA TTCACTTGC AAGATTGCGT TACCTECACA GCGTGTGCGG TCGCGGACCC CTTGAAAAGG CTGTTAAAGT  
7321 TGGTAAAACC GCTCCAGCC GACGATGAGC AAGAGGAAGA CAGAAGACGC GCTGTCTAG ATGAACAAA GCGTGCTTT AGAGTAGGTA TAACAGACAC CTTAGCAGTG CCGCTGCCAA  
7441 CTGCGTATGA GGTAGACAAC ATCACACCTG TCTGTCTGCC ATTGAGAACT TTTCGCCAGA GCAAAAAGAGC ATTTCAGCC ATCAGAGGGG AAATAAAGCA TCTCTACGCT GGTCTAAAT  
7561 AGTCAGCATA GTACATTTCA TCTGACTAAT ACCACAACAC CACCACCATG AATAGAGGAT TCTTTAATCAT GCTCGGCGCC CCGCCCTTCC CAGCGCCCACT TCGCATGTGG AGCGCGCGGA  
7681 GAAGGAGGCA GCGCGCCCGG ATCGTCCGC GCAATGGGCT GCTTCCCAA ATCCAGCAAC TGACACAGC GTCAGTGCC CTAGTCATTG GACAGGCAAC TAGACCTCAA ACCCCAGGCC  
7801 CACCGCCGCC CCGCGCCAG AAGAGCAGG CCGCAAGCA ACCACCGAAG CCGAAGAAC CAAAAACACA GGAGAGGAAG AAGAGCAAC CTCGAAAACC CAAACCCGGA AAGAGACAGC  
7921 GTATGGCACT TAAGTGGAG GCGACAGAC TGTTCGAGCT CAAAATGAG GACGGAGATG TCATCGCGCA CCGACTGCC ATGGAAGGAA AGGTAATGAA ACCACTCCAC GTGAAGGAA  
8041 CTATTGACCA CCGTGTCTA TCAAGCTCA AATTCACCA GTGCTACGA TACGACATGG AGTTCGACA GTTCGCGTC AACATGAGAA GTGAGCGCTT CACTACACE AGTGACACC  
8161 CTGAAGGGTT CTACAAGTG CACCAAGGAG CCGTGCAGTA TAGTGGAGCC AGATTACCA TCCCGCCCGG AGTAGGAGCC AGAGGAGACA GTGTCTGCTC GATTATGGAT AACTCAGGCC  
8281 GCGTTGTGCG GATAGTCTC GGAGCGGAGC AAGAGCGAAC AAGACAATCA AGACAATCA AGACAATCA AGACAATCA AGACAATCA AGACAATCA AGACAATCA AGACAATCA AGACAATCA  
8401 CTGCTGACCC ACTGTCTCC GCGATGTCT TCTGTGAAA COTGAGCTT CCACTCAATC CCGCGCCAC ATGCTACACC CCGGAACCAT CAGAGCTCT GACATCTCT GAAGGAGAGC  
8521 TGAACACGCA GCGCTACGAC ACCGTCTCA ACGCATATT CCGGTCCGA TGTCTCCGA GAATAAAAG AAGCTCACT GACGACTTTA CTTGACCGC CCGTACTTG GGCACATGCT  
8641 CGTACTGTCA CCATAGTAA CCGTCTTTA CCGCGATTAA GATGAGGAG GTGTGGATG AAGCGGAGCA CAACACATA CCGATACAGA CTTCGCGCA GTTTGATAC GACCAAGCG  
8761 GAGCAGCAAG CTCAAATAAG TACCGCTACA TGTCTCTGA CAGGATCAT ACTGTCAAG AAGCGACCAT GATGACATC AAGATACGA CCGAGGACC GTTAGAGAGG CTTAGCTACA  
8881 AAGGATACTT TCTCTCGG AAGTGTCTC CAGGGGACAG CGTAACGTT AGCATAGGCA GTAGCAACT AGCAAGCTCA TGCACAATG CCGCAAGAT AAAACCAAAA TTGTTGGGAC  
9001 GGGAAAATA TGACCTACT CCGTTCACG GTAAGAGAT TCTTTCACA GTGTAGGACC GTTGAAGA AACACCGCC GGTACATCA CTATGACAG CCGCGGACCG CACGCTATA  
9121 CATCTATCT GGAGGAATCA TGAGGAAAG TTACCGGAA GCGACATCC GCGAAGAACT TTACTAGCA GTGCAAGTCC GCGATTACA AGACCGGAA CCGTACGACC GTTAGCGAAA  
9241 TCACGGGCTG CACCGCATC AAGCAGTGG TCGCTATAA GAGCGACCAA ACGAAGTGG TTTCAACTC CCGGAGTCC ATCAGACAG CCGACACAC GCGCAAGG AAATGCAAT  
9361 TCGCTTTCAA GCTGATCCG AGTACCTCA TGTCTCTCT TCGCCAGCG CCGAAGTAG TACACGGCTT TAAACACATC AGCTTCAAT TAGACACAGA CCATCTGACA TTGCTACCA  
9481 CCAGGAGACT AGCGGCAAG CCGGAACCAA CCACTGAATG GATCATGGA AACACGGTTA GAACTTCAC CCGTACGGA GATGCGCTGG AATACATATG GCGCAATCAC GAACAGTAA  
9601 GGTCTATG CCAAGAGTCT GCGCAGGAG ACCCTACCG ATGCGCACAC GAAATAGTAC AGCATTACTA TCATCGCAT CCGTGTACA CCATCTTAC CCGTACATA GCTGTGTG  
9721 CGATGATGAT TCGGTAACT GTTCAGCAT TATGTCTCT TAAAGCGCG CCGTGTGCT TGACCGCAT TCGCTGCC CCAATCGCG TCAATCCAC TTGCTGGCA CTTTGTCT  
9841 GTGTTAGTCC GCTAATGCT GAAACATTA CCGAGACCAT GAGTTACTTA TGTGGAACA GCGACCGCTT CTCTGCTC CAGCTGTGTA TACCTCTGC CCGTCTCTC GTTCTAATG  
9961 GCTGTCTC ATGCTGCTC CTTTITTAG TGTGCGCG CCGTACTG GCGAAGTAG ACGCTACGA ACATGCGACC ACTGTTCAA ATGTGCAACA GATACCTAT AAGGCACTG  
10081 TTGAAAGGCG AGCGTACGCC CCGCTCAAT TGGAGATTAC TGTGATGTC TCGGAGTIT TCGCTTCCAC CAACCAAGAG TACATTACCT GCAAAATCAC CACTGTGTC CCGTCCCTA  
10201 AAGTCAGATG CTGCGCTCC TTGGAATGTC AGCGCGCGC TCAGCGAGC TATCTTCA AGGTCTTGG AGCGGTGAC CCGTCTATG GCGGAGGAGC ACAATTTTT TCGACAGTG  
10321 AGACAGCCA GATGATGAG GCGTACGTC AATTGTCACT AGATTGCGG ACTGACAGC CCGAGGCGAT TAAAGTGCAT ACTGCGCGA TGAAGTAGG ACTGCGTATA GTGTACGGGA  
10441 ACACACAGC TTCTAGAT GTGACGTGA ACGGAGTCA ACCAGGAGC TCTAAGAGC TGAAGTCA AGCTGAGCA ATTTAGCAT TGTTCACAC ATTGATCAC AAGGTGTTA  
10561 TCAATCGCG CCGTGTGAC AACTATGACT TTGCGAATA CCGAGCGATG AAACAGGAG CTTTGGAGA CATTCAAGCT ACCTCTTGA CTAGCAAGA CCGTACGCGC AGCAGAGCA  
10681 TTAGCTACT CAAGCTTCC GCGAAGAGG TGCATGTCC GTACAGCGAG CCGCATCTG GATTCGAGAT GTGAAAAC AACTAGGCC CCGCACTGCA GAAACCGCC CTTTGTGGT  
10801 GCAAGATTGC AGTCAATCG CTGAGCGCG TCGACTGCT ATACGGGAA ATTCCTATT CTATTGACAT CCGAAGCGT CCGTTATCA GGACATGGA TCGACCACTG GTCTAACAG  
10921 TCAATGTGA TGTAGTGAG TGCATTAT CAGCGACTT CCGAGGATG GCTACCTGC AGTATGATE CAGCGCGAA GGAATGCG CTGTACATTC GCATTGAGC ACAGAACCC  
11041 TCCAAGATC GACAGTTCAT GTCTGAGA AAGGAGCGGT GACAGTACAC TTACGACCG CCGCCCAACA GCGAATTC ATTGTATCG TGTGTGTA GAAAGACACA TCGAATGAG  
11161 AATCGAAACC ACCAGCTGAT CATATCTGA GCAACCGCA CAAAATGAG CAAGAAATCC AAGCGCCAT CTCAAAACCT TCAATGAGTT GCGTGTTCG CTTTTCGCG GCGCGCTGT  
11281 CCGTATTAAT TATAGACTT ATGATTTTT CTTCAGCAT GATGCTGACT AGCAGAGAA GATGACCGT ACGCCCAAT GACCGAGCA GCAAACTG ATGTACTTC GAGGAATGA  
11401 TGTGATAAT GCATAGGCT GGTATATTAG ATCCCGCTT ACCCGCGCA ATATAGCAAC ACCAAACTE GACGTATTT CAGGGAAGCG CAGTGCATA TGTGCGCAG TTTGCCAAA  
11521 TAATCACTAT ATTAACCAT TATTCAGCG AGCGCAAAAC TCAATGTATT TCTAGGAAG CATGTCAT AATGCCATG AGCGTCTGA TAACTTTTA TTATTTT TATTAATCA  
11641 CAAAATTTG TTTTAAAT TTT

Fig. 5 B

## Nucleotide Sequence of TR339

1 ATTGGCGGCG TAGTACACAC TATTGAATCA AACAGCGGAC CAATTGCACT ACCATCACAA TGGAGAAGCC AGTAGTAAAC GTAGACGTAG ACCCCACAGAG TCCGTTTGTG GTGCAACTGC  
121 AAAAAAGCTT CCGCAATTT GAGGTAGTAG CACAGCAGGT CACTCEAAAT GACCATGCTA ATGCCAGAGC ATTTTGGCAT CTGGCCAGTA AACTAATCGA GCTGAGGTTT CTAACACAG  
241 CGACGATCTT GGACATAGGC AGCGCACCGG CTGTTAGAA GTTTTCCGAG CACCAATGATC ATTGTGTGTG CCCCATGCGT AGTCACAGAG ACCCGGACCG CATGATGAAA TATGCCAGTA  
361 AAAATGCGCGA AAAAGCGTGC AAGATTACAA ACAAGAACTT GCATGAGAAG ATTAAGGATC TCGGACCGT ACTTGATAGG CCGGATGCTG AAACACCATC GCTCTGCTTT CACAACGATG  
481 TTACTGTCAA CATGCTGEC GAATATTCG TCATGACGGA COTGTATATC AACGCTCCG GAATCATCTA TCATGAGCT ATGAAGCGG TCGGACCGT GTACTGATT GCGTTGACA  
601 CCACCCAGTT CATGTTTCTG GCTATGCGAG GTTCGTACCC TCGTACAA ACCTAATGCG CCGACGAGAA AGTCCTTGAA CGCGTAACA TCGGACTTTC CAGCACAAG CTGATGAAAG  
721 GTAGGACAGG AAAATGTGCT ATAATGAGGA AGAAGGAGTT GAAGCGCGG TCGCGGTTT ATTTCTCGT AGGATGACA CTTTATCCAG AACACAGAGC CAGCTTGCAG AGCTGCGATC  
841 TTCCATCGGT GTTCCACTTG AATGGAAGG AGTGTACAC TTGCGCTGT GATACAGTGG TGAGTTGCGA AGCTACGTA GTGAAGAAAA TCACCATCAG TCCCGGATC ACCGGAGAAA  
961 CCGTGGGATA CCGGTTACA CACAATAGCG AGGCTTCTT GCTATGCAA GTTACTGACA CAGTAAAGG AGAACGGTA TCGTCCCTG TGTGACGTA CATCCCGCC ACCATATGCG  
1081 ATCAGATGAC TGTATAATG GGCAGGATA TATCACTGA CGATGACAA AAATCTCTG TTGGCTCAA CCAGCGAAT GTCAATTAAG GTAGGACTAA CAGGAACACC AACACCATG  
1201 AAAATTACCT TCTGCCGATC ATAGCACAAG GCTTCAGCAA ATGGCTTAAG GCGCGAAGG ATGATCTGTA TAACGAGAAA ATGCTGGTA CTAGAGAAAG CAAGCTTACG TATGCTGCT  
1321 TGTGGCGTT TCGCACTAAG AAAGTACATT CTTTTATCG CCCACCTGGA ACCGAGACCA TCGTAAAGT CCCAGCTCT TTAAGCGTT TTECATGTC GTCCGTATGG ACGACTCTT  
1441 TCGCATGTC GCTGAGGAG AAATGAAAC TGGCATGCA ACCAAGAGG GAGGAAAAAC TCGTGCAGT CTGGAGGAA TTAGTCATG AGGCCAAGCG TCGTTTGAAG GATGCTCAGG  
1561 AGGAAGCCAG AGCGGAGAGG CTCGAGAGG CACTTCCACC ATTAGTGGCA GACAAGGCA TCGAGGCGAG CCGAGAGTT GTCTGGAAG TCGAGGGCT CAGGCGGAC ATCGGAGCAG  
1681 CATTAGTTGA AACCCCGCG GGTACGTA GGATAATACC TCAAGCAAT GACCGTATGA TCGGACATA TATGTTGTC TCGCCAACT CTGTCTGAA GAATGCCAAA CTCGACCCAG  
1801 CGCACCCGCT AGCAGATCAG GTTAAGATCA TAACACACTE CGGTAGATCA GGAAGGTAG CGGTGGAAC ATACGACCT AAAGTACTGA TCCGAGCAG AGGTGCGTA CCATGGCCAG  
1921 AATTCTAGC ACTGAGTAG AGCGCCACGT TAGTGTACAA CGAAGAGAG TTTGTGAACC GCAACTATTA CCACATTGCT ATGATGCGC CCGCAAGAA TACAGAGAG GAGCAGTACA  
2041 AGGTACAAA GGCAGAGCTT CCAGAAACAG AGTACGTGTT TGACGTGGAC AAGAAGCTT AGTATAAGAA GGAAGAGGCC TCAGGTCTGG TCCTCTCGG AGAAGTACC AACCTCTCT  
2161 ATCATGAGCT AGCTCTGAG GGAAGTGA CCGACCTGC GTCCCGTAC AAGGTGAAA CAATAGGAGT GATAGGCACA CCGGGTCCG GCAAGTCAG TATTATCAA TCACTGTCA  
2281 CCGCACGGGA TCTGTACC AGCGGAAAG AAGAAAATT TCGCGAATT GAGCGGAGG TCGTAAGACT GAGGGTATG CAGATTAGT CGAAGACAGT AGATTGCGT ATGCTCAAG  
2401 GATGCCACAA AGCGTAGAA GTCTGTATG TTGACGAAG GTTCGCTGC CAGCGAGGAG CACTACTTC CTGATTGCT ATGTCAGGC CCGCAAGAA GGTAGTACTA TCGGAGACC  
2521 CCATGCAATG CGGATTCTT AACATGTC AACATAAGT ACATTCAAT CACCTGAAA AAGACATATG CACCAAGACA TTCTACAAGT ATATCTCCG CCGTGCACA CAGCAGTTA  
2641 CAGCTATTGT ATGACACTG CATTAGATG GAAGATGAA ACCACGAA CCGTCAAGA AGAACATTGA AATGATATT ACAGGGCCA CAAGCGGAA GCGAGGGAT ATCATCTGA  
2761 CATGTTCCG CCGGTGGTT AAGCAATTG AAATGCACTA TCCCGACAT GAAGTAATGA CAGGCGCG CTACAAGG CTACAAGG AAGGAGTGA TCGCTCCG CAAAAGTCA  
2881 ATGAAAACCC ACTGTACCG ATCAGTAC AGCATGTGA COTGTGCT ACCCGACTG AGGACAGCT AGTGTGAAA ACCTTGAGG CGACCCATG GATTAAGCAG CTAECTAACA  
3001 TACCTAAGG AAATTTAG GCTACTATG AGGACTGGA AGCTGAAC ACAGGAAATA TTGCTCAAT AAACAGCCC ACTCCCGTG CCAATCGTT CAGCTGCAAG ACCAAGCTT  
3121 CCGCGGCAA AGCATGGA CCGATACAT CACCGCGCG TATGTAAT ACCGTTCCG AGTGAGCGA ACTGTTCCA CAGTTTCCG ATGACAAA ACATTGCGC ATTTAGCGT  
3241 TAGACGTAAT TTGCAATAG TTTTCGCA TGGACTGAC AAGCGAGT TTTTAAAC AGAGCATCC ACTAAGTAC CATCCCGCG ATTCAGCGG GCGGTAGCT CATTGGACA  
3361 ACAGCCGAG AACCCGCA TATGGTAC ATACGCCAT TCCCGCGAA CTCTCCGTA GATTTCGGT GTTCCAGTA GCTGGGAAG GCACACAAT TGATTGCAG ACGGGAGAA  
3481 CCAGAGTAT CTGTGACAG CATAAGCTG TCCCGTGAA CCGCAATTT CTAAGCGCT TAGTCCCGA GTACAAGGA AAGCAACCG CCGCGTGA AAAATTCTT AACCACTTA  
3601 AACACCACTE AGTACTGT GTATCAGAG AAAAAATTGA AGCTCCCGT AAGAGAAATG AATGATGTC CCGGATTGCT ATAGCGGTG CAGATAAGAA GTACAACCTA CTTTCCGCT  
3721 TCCCGCGCA GGCAGGTAC GACCTGTGT TCATCAACAT TGGAACTAAA TACAGAAAC ACCACTTICA GCAGTGGAA GACCATGCG CGACCTTAAA AACCTTTCT GCTTCCGCG  
3841 TGAATTGCT TAACCCAGGA GGCACCTCG TGTGAAATG CTATGCTAC GCGACCGCA ACAGTAGGA CGTAGTCACC GCTCTGCA GAAAGTTGT CAGGTGTCC GCAGCGAGAC  
3961 CAGATTGCT CTCAGCAAT ACAGAAATG ACCTGATTT CCGACAATA GACAACGCG GTACACGGCA ATTAACCCG CACCATCTGA ATTGCTGAT TCTCTGCTG TATGAGGTA  
4081 CAAGAGATG AGTTGGAGC GCGCGCTAT ACCGACCAA AAGGAGAA ATTTGTAET GTCAAGAGGA AGCAGTTGTC AACCGACCA ATCCGCTGG TAGACAGCG GAAGGAGTCT  
4201 GCGTGCCAT CTATAAGCT TGGCGACCA GTTTACCGA TTCAAGCAG GAGACAGCA CCGCAAGAT GACTGTGTC CTAGGAAAG AAGTATCCA CCGCGTCCG CTTGATTTT  
4321 GGAAGCACCC AGAAGCAGAA CCGTTGAAAT TGTACAAAA CCGCTACCAT GCAGTGGCAG ACTTATGAAA TGAACATAAC ATCAAGTCTG TCGCAATTC ACTGATATC ACAGGCAATT  
4441 ACCGAGCGG AAAAGACCG CTTGAAGTAT CACTTAACT CTTGACAAAC GCGTAGACA GAAGTACCG GGACGTAAAC ATCTATTGCT TGGATAAGAA GTGGAAGGAA AGAATGAGC  
4561 ECGCACTCCA ACTTAAGGAG TCTGTAAAG AGCTGAAGG TGAAGATAT GAGATGAGC ATGAGTTAGT ATGATECAT CCAGACAGT GCTTGAAGG AAGAAAGGAA TTCACTACTA  
4681 CAAAAGGAAA ATTGTATTC TACTTGAAG GCACCAAAAT CCATCAAGCA GCAAAAGACA TCGCGGAGAT AAAGGTCTG TTECTAATG ACCAGGAAAG TAATGAACAA CTGTGTGCT  
4801 ACATATTGG TGAGCAATG GAAGCAATC CGCAAAAGT CCGGTGCG ACATAACCGT CGTATAGCC GCGCAAAAG TTGCGTGC TTTGATGTA TGCATGAGC CCAGAAAGG  
4921 TCCACAGAT TAGAAGCAAT AAGTCAAG AGTTACAGT ATGCTCTCC ACCCGCTTC CTAAGCACA AATTAAGAT GTTCAAGAG TTCAAGTAC GAAAGTATC CTTTAAAT  
5041 CGCACTCC CCGATTGCT CCGCGCTGA AGTACATAGA AGTGCAGAA CAGCTACCG CTCTCTGTC ACAGCGCGAG GAGCGCCCG AAGTTGATG GACACCGTCA CCATCTACAG  
5161 CTGATAACAC CTGCTGAT GTACAGACA TCTCACTGA TATGATGAC AGTAGGAG GCTCACTTT TTGAGCTTT AGCGATCGG ACACTCTAT TACTAGTAT GACAGTGGT  
5281 CGTGAGGAC TAGTCTACTA GAGATAGAG ACCGAAGGCA GGTGTGCTG CTAAGCTTC ATGCGTCA AGAGCTGCT CTAATTCAC CCGCAAGCT AAAGAAGAT GCGCGCTCG  
5401 CAGCGCAAG AAAAGAGCCC ACTCCACCG CAAGCAATAG CTCTGATTC CTCACCTCT CTTTGTGTC GGTATCCATG TCCCTCGAT CAATTTGCA CCGAGAGAG GCGCGCACG  
5521 CAGCGTACA ACCCTGGA ACAGCGCCA CCGATGTC TATGTTTC GATGCTTT CCGACGGAG GATTGATGAG CTAGCGCCA GAGTAACTGA GTCCGAACCC GTCTGTTT  
5641 GATCAATTGA ACCCGCGAA GTGAATCAA TTATATGTC CCGATAGCC GTATCTTTC CACTACGCA CAGAGACGT AGAGCGAGG GAGGAGGAC TGAATCTGA CTAACCGCG  
5761 TAGTGGGTA CATATTTTC CCGGACACAG GCGCTGCGA CTGCAAAAG AATGCGTTC TCGCAACCA GCTTACAGAA CCGACCTTG AGCGCAATG CTTGGAAGA ATTAATGCCC  
5881 CCGTGTGCA CAGCTGAAA GAGGAACA TCAAACTCAG GTACAGATG ATGCGACCG AAGCAACAA AAGTAGTAC CAGTCTGTA AAGTAGAAA TCAGAAAGC ATAACACTG  
6001 AGCGACTACT GTACAGTA CCACTGTATA ACTGCGCAC AGATACGCA GAATGTATA AGATACCTA TCGGAAACA TTGACTCCA GTAGCTACC GCGCAACTAC TCGGATCAC  
6121 AGTTGCTGT AGCTGTCTT AACAACTAT TGCATGAGAA CTATCGGACA GTAGCATTT ATCAGATTAC TGACGAGTAC GATGCTTACT TGGATATGT AGACGGACA GTCCGCTCC  
6241 TGGATATGC AACCTTTC CCGCTAAGC TTAGAAGTA CCGGAAAAA CATGATATA GAGCCCGAA TATCCGAGT CCGGTTCAT CAGCGATGA GAACAGCTA CAAAATGTC  
6361 TCATTGCGC AACTAAAGA AATTGCAAG TCACCGAGT CCGTGAAGT CCAACACTG ACTCAGGAC ATCAATGTC GAATGCTTC GAAATATGC ATGTAATGC GAGTATTGG  
6481 TCGAGTTCC TCGGAGCCA ATTAGATTA CCACTGAGT TGTACCGCA TATGATGTA GACTGAAG CCGTAAGCC CCGCACTAT TCGAAGAG GATAATTG GTCCATTC  
6601 AAGAGTCC TATGATAGA TTGCTATG ACATGAAAG AGAGTGAA GTTACACAG GCACGAAACA CACAGAGAA AGACCGAAG TACAAGTAT ACAAGCGCA GAACCTG

Fig 6A.

6721 CGACTGCTTA CTTATGCGGG ATTACCGGG AATTAGTGG TAGGCTTACG GCCGTCTTC TTCAAACAT TCACAGCTT TTGACATGT CGGCGGAGGA TTTTGATCA ATCATAGCAG  
6841 AACACTTCAA GCAAGGCGAC CCGGTACTGG AGACGGATAT CCGATCATT GACAAAAGCC AAGACGAGC TATGGCGTTA ACCGTTCTGA TGATCTTGA GGACCTGGGT GTGGATCAAC  
6961 CACTACTCGA CTTGATCGAG TCGGCTTTG GAGAAATATC ATCCACCAT CTACCTACGG GTACTCGTT TAAATTCGG GCGATGATGA AATCGGAAT GTTCTCACA CTTTTGTCA  
7081 ACACAGTTTT GAATGTCTT ATCCGAGCA GAGTACTAGA AGACGGCTT AAAAGGTCCA GATGTGCAG GTTCATTGGC GACGACAACA TCATACATG AGTAGTATCT GACAAAAGAA  
7201 TGGCTGAGAG GTGCGGCACC TGGCTCAACA TGGAGGTAA GATCATGAC GCACTCATCG GTGAGAGACC ACCTTACTTC TCGGCGGAT TTATCTTGA AGATTGCTT ACTTCCACAG  
7321 CGTGCGCGCT GCGGAGCCCG CTGAAAAGC TGTTAAGTT GGTAAACCG CTCCAGCCG ACGAGGAGCA AGACGAAGAC AGAAGACCG CTCTGCTAGA TGAACAAAAG CGGTGTTTA  
7441 GAGTAGGTAT AACAGCACT TTAGCATGG CCGTAGGAC CCGGTATGAG GTAGACAATA TTACAGCTGT CTACTGCA TTGAGAACT TTGCCCAGAG CAAAAGAGCA TTCCAGGCA  
7561 TCAGAGGCGA AATAAGCAT CTCTACGTT GTCTAAATA GTACGATAG TACATTTCAT CTGACTAATA CTACAACACC ACCACCATGA ATAGAGGATT CTTTAAACAT GTGCGCGCC  
7681 GCGCTTCCG GCGGCCACT GGCATGTGA GCGCGGAG AAGAGGAGC GCGGCCCGA TGCTGCCCG CAACGGGCTG GCTTCTCAA TCGAGCACT GACCACAGCC GTACGTGCC  
7801 TAGTCATTGG ACAGGCACT AGACETCAAC CCCCAGTCC ACCCGGCCA CCGCGGAGA AGAAGCAGC GCCAAGCAA CCACCGAAGC CGAAGAAAC AAAACCGAG GAGAAGAGA  
7921 AGAAGCAACC TCAAAAACC AAACCGGAA AGAGACAGC CATGCACTT AAGTGGAGG GCGACAGATT GTTCGAGCTC AAGAAGGAG ACCGAGATGT CATCGGGCAC GCACTGGCCA  
8041 TGGAGGAAA GGTAAAGAAA CCTCTGACG TGAAGGAAC CATGACCAAC CCGTGTCTAT CAAAGCTCAA ATTTACCAAG TGTGAGCAT ACGACATGA GTTCGACAG TTGCGAGTCA  
8161 ACATGAGAG TGAGGCATT ACCTACACCA GTGAACACC CGAAGGATC TATAACTGG ACCACGGAG GGTGCAATAT AGTGAGGTA GATTTACCAT CCGTCGCGA GTAGAGGCA  
8281 GAGGAGACAG CCGTCTCCG ATCATGGTA ACTCGGTGG GGTGTGCGG ATAGTCTCG GTGAGCTGA TGAAGGAACA CGAACTGCC TTTCGGTGT CACCTGGAAT AGTAAAGGA  
8401 AGACAAATTA GACGACCCG GAAGGACAG AAGAGTCTC CCGACACCA CTGCTACCG CAATGTGTT GTCTGGAAAT GTGAGCTTC CATGCGACCG CCGGCCACA TGCTATACC  
8521 CGGAACCTTC CAGAGCCCTC GACATCTTG AAGAGACGT GAACATGAG GCCTACGATA CCGTCTCAA TGCCATATT CCGTGGGAT CCGTGGCAG AAGCAAAAGA AGCGTCACTG  
8641 ACAGCTTAC CCGACAGC CCGTACTGG GCACATGCT GTACTGCCA CATACTGAC CCGTCTTAC CCGTCTTAA AGTGCAGCG TCTGGGAGA AGCGGACGAT AACACATCA  
8761 GCATACAGAG TTCCGCCAG TTGGATACG ACCAAGCGG ACCAGCAAGC GCAAAACAAT ACCGTACAT GTCGCTTGA CAGGATCAEA CCGTAAAGA AGCCACCATG GATGACATCA  
8881 AGATTAGCAC CTCAGGACCG TGTAGAAGC TTAGCTACA AGGATACCT CTCTGCCAA AATGCCCTC AGGGGACAG GTAAAGGTTA GCATAGTGA TAGCAACTCA GCAAGCTCAT  
9001 GTACACTGGC CCGCAAGTA AAACCAAAAT TGTGGGAGC GGAATAATAT GATCTACCT CCGTTCACG TAAAAAAT CTTGACAG TGTACGACCG TGTAAAGAA ACAATGAG  
9121 GCTACATCAC TATGACAG CCGGACCCG ACCGTATAC ATCTACTCG GAAGATCAT CAGGGAAGT TTACGCAAG CCGCATCTG GGAAGAACAT TACGTATGAG TCGAAGTGG  
9241 CGCACTACAA GACCGAACG GTTTCAGCC GCAACGAAAT CACTGTTTC ACCGCCATCA AGCAGTGGT CCGCTATAAG AGCGACCAA CGAAGTGGT CTTCAACTCA CCGGACTGA  
9361 TCAGACATGA CGACACAGC GCCAAGGGA AATTGCATT GCTTTCAAG TTGATCCGA GTACTGCAT GGTCCCTGT GCCCAGCGC CGAATGTA ACATGCTTT AAACATCA  
9481 GCTTCAATT AGATACAGC CACTGACAT TGCTACAC CAGGAGCTA GGGCAAAAC CGGAACCAAC CACTGAATG ATCGTCGAA AGACGGTCAG AAACCTCAC GTGACCGAG  
9601 ATGGCTCGA ATACATATG GGAATCATG AGCAGTGAG GGTETATGC CAAGAGTCAG CACCAAGAGA CCGTACCGA TGCCACAGC AAATAGTACA GCATTACTAC CATECCATC  
9721 CTGTATACAC CATCTAGC GTCCATAG CTACCCTGC GATGATGAT GCGTAACCG TTGAGTGT ATGTGCTGT AAAGCGCGC GTGAGTGGT GACCGCATAC GCGCTGGCC  
9841 CAAAGCGCT AATCCAACT TCGTGGCAG TCTGTGCTG CTTAGGTG GCAATGCTG AAGCTTCAC CGAGACCATG AGTTACTGT GTTGAACAG TCAGCCGCTC TTCTGGTCC  
9961 AGTTGTGAT ACCTTTGGC GCTTCATCG TTCTAATCG CTGCTGCTC TGCTGCTCG CTTTTTAT GTTTCGGC GCTACCTG CGAAGGTAGA CGCTACGAA CATGCGACA  
10081 CTGTTCCAAA TGTGACAG ATACCGTATA AGGCACTTGT TGAAGGGGA GGTATGCCC GCGTCAATT GAGATCACT GTCATGCTT CGGAGTTTT GCTTCCACC AACCAAGAT  
10201 ACATTACTCG CAATTCACC ACTGTGCTC CTTCCCAAA AATCAATGC TCGGTCTCT TGAATGTCA GCGGCGCT CATGCACT ATACCTGCAA GGTCTTCCA GGGGTCTACC  
10321 CTTTTATGT GGGAGGAGC CAATGTTTT GCGACAGTA GAACAGCCAG ATGAGTGAG COTACGTGA ACTGTGACA GTTGGCGT CTGACCAGC CGAGCGAT AAGGTGACA  
10441 CTGCGCGAT GAAAGTAGGA CTGCTATAG TGTACGGAA CACTACAGT TTCTAGATG TGTACGTGA CGGATCACA CCAGGAAGT CTAAGACTT GAAAGTATA GGTGACCAA  
10561 TTTGAGCAT GTTACGCCA TTGATCATA AGGTGTTAT CCATGCGCG CTGGTGTACA ACTATGACT CCGGAATAT GAGCGATGA AACAGGAGC GTTGTAGAC ATTCAGCTA  
10681 CCGCTTAC TAGCAAGAT CTCATGCCA GCACAGCAT TAGGCTACT AAGCTTCCG CCAAGAACGT GCATGTCCG TACAGCAGG CCGCATCAG ATTGAGATG TGAAGAACCA  
10801 ACTAGGCGC CCGACTGAG GAAACCGCAC CTTTCGGTG TAAGATTGA GTAAATCCG TCGAGCGGT GAGCTGTCA TACGGGAACA TTCCATTTC TATTGACATE CGGAACGCTG  
10921 CTTTTATAG GACATAGAT GCACACTGG TCTAACAGT CAATGTGAA GTAGTGAT GCACTTATC AGCAGACTT GCGCGGATG CCACCTGCA GTATGTATCC GACCGGAG  
11041 GTCAATGCC CGTACATTG CATTEGACA CAGCACTCT CCAAGAGTG ACAGTACATG TCTGGGAA AGGAGCGGT ACAGTACACT TTAGCAGCG GAGTCCACAG CGGAACCTTA  
11161 TGATATGCT GTTGGGAAG AAGACAACAT GCAATGAGA ATGTAAACA CCACTGACC ATATGCTGAG CACCCGAC AAAAATGACC AAGAATTICA AGCGCCATC TCAAAACAT  
11281 CATGGAGTG GTGTTTGGC CTTTCGGCG GCGCTGCT GCTATTAAT ATAGGACTA TGATTTTTC TTGAGCATG ATGCTGACT GCACACGAAG ATGACCGCTA CCGCCCAATG  
11401 ATCCGACAG CAAACTCGA TGTACTTCC AGGAAGTAT GTGCATAAT CATAGGCTG GTACATTAGA TCCCGCTTA CCGCGGCAA TATAGCAACA CTAACAACT GATGTACTTC  
11521 CGAGGAAGCG CAGTGCATA TCTGCGCAG TTTGCEACA TAACCACTAT ATTAACCAAT TATCTAGCG AGCCCAAAA CTAATGTAT TTCTAGGAA CGGTGTGCA TAATGCCAG  
11641 CAGCTGCTG ATAACCTTA TATTCTTT TATTAATCAA CAAATTTTG TTTTAAACAT TTC

FIG. 6B

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